



## INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

<b>(51) International Patent Classification 5 :</b> C12N 15/82, 15/32, 15/67 C12N 15/40, 5/10, A01H 5/00	<b>A1</b>	<b>(11) International Publication Number:</b> WO 90/10076 <b>(43) International Publication Date:</b> 7 September 1990 (07.09.90)
<b>(21) International Application Number:</b> PCT/US90/00778 <b>(22) International Filing Date:</b> 13 February 1990 (13.02.90)  <b>(30) Priority data:</b> 315,355 24 February 1989 (24.02.89) US 476,661 12 February 1990 (12.02.90) US  <b>(71) Applicant:</b> MONSANTO COMPANY [US/US]; 800 North Lindbergh Boulevard, St. Louis, MO 63167 (US).  <b>(72) Inventors:</b> FISCHHOFF, David, Allen ; 28 North Maple Avenue, Webster Groves, MO 63119 (US). PERLAK, Frederick, Joseph ; 11035 Crimson Drive, St. Louis, MO 63146 (US).		<b>(74) Agent:</b> COLE, Arnold, Harvey; Monsanto Company, 800 North Lindbergh Boulevard, St. Louis, MO 63167 (US).  <b>(81) Designated States:</b> AT (European patent), AU, BE (European patent), BG, BR, CA, CH (European patent), DE (European patent), DK (European patent), ES (European patent), FI, FR (European patent), GB (European patent), HU, IT (European patent), JP, LU (European patent), NL (European patent), NO, RO, SE (European patent), SU.  <b>Published</b> <i>With international search report. Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</i>
<b>(54) Title:</b> SYNTHETIC PLANT GENES AND METHOD FOR PREPARATION  <b>(57) Abstract</b>  A method for modifying structural gene sequences to enhance the expression of the protein product is disclosed. Also disclosed are novel structural genes which encode insecticidal proteins of <i>B.t.k.</i> HD-1, <i>B.t.k.</i> HD-73, <i>B.t. tenebrionis</i> , <i>B.t. entomocidus</i> , 2 protein of <i>B.t.k.</i> HD-1, and the coat protein of potato leaf roll virus.		

BEST AVAILABLE COPY

***FOR THE PURPOSES OF INFORMATION ONLY***

Codes used to identify States party to the PCT on the front pages of pamphlets publishing international applications under the PCT.

AT	Austria	ES	Spain	MG	Madagascar
AU	Australia	FI	Finland	ML	Mali
BB	Barbados	FR	France	MR	Mauritania
BE	Belgium	GA	Gabon	MW	Malawi
BF	Burkina Faso	GB	United Kingdom	NL	Netherlands
BG	Bulgaria	HU	Hungary	NO	Norway
BJ	Benin	IT	Italy	RO	Romania
BR	Brazil	JP	Japan	SD	Sudan
CA	Canada	KP	Democratic People's Republic of Korea	SE	Sweden
CF	Central African Republic	KR	Republic of Korea	SN	Senegal
CG	Congo	LJ	Liechtenstein	SU	Soviet Union
CH	Switzerland	LK	Sri Lanka	TD	Chad
CM	Cameroon	LU	Luxembourg	TG	Togo
DE	Germany, Federal Republic of	MC	Monaco	US	United States of America
DK	Denmark				

-1-

SYNTHETIC PLANT GENES AND METHOD FOR PREPARATIONBACKGROUND OF THE INVENTION

5

The present invention relates to genetic engineering and more particularly to plant transformation in which a plant is transformed to express a heterologous gene.

10

Although great progress has been made in recent years with respect to transgenic plants which express foreign proteins such as herbicide resistant enzymes and viral coat proteins, very little is known about the major factors affecting expression of foreign

15 genes in plants. Several potential factors could be responsible in varying degrees for the level of protein expression from a particular coding sequence. The level of a particular mRNA in the cell is certainly a critical factor.

20

The potential causes of low steady state levels of mRNA due to the nature of the coding sequence are many. First, full length RNA synthesis might not occur at a high frequency. This could, for example, be caused by the premature termination of RNA during transcription or due to unexpected mRNA processing

25 during transcription. Second, full length RNA could be produced but then processed (splicing, polyA addition) in the nucleus in a fashion that creates a nonfunctional mRNA. If the RNA is properly synthesized, terminated and polyadenylated, it then

30 can move to the cytoplasm for translation. In the

-2-

cytoplasm, mRNAs have distinct half lives that are determined by their sequences and by the cell type in which they are expressed. Some RNAs are very short-lived and some are much more long-lived. In addition, there is an effect, whose magnitude is uncertain, of translational efficiency on mRNA half-life. In addition, every RNA molecule folds into a particular structure, or perhaps family of structures, which is determined by its sequence. The particular structure of any RNA might lead to greater or lesser stability in the cytoplasm. Structure per se is probably also a determinant of mRNA processing in the nucleus. Unfortunately, it is impossible to predict, and nearly impossible to determine, the structure of any RNA (except for tRNA) in vitro or in vivo. However, it is likely that dramatically changing the sequence of an RNA will have a large effect on its folded structure. It is likely that structure per se or particular structural features also have a role in determining RNA stability.

Some particular sequences and signals have been identified in RNAs that have the potential for having a specific effect on RNA stability. This section summarizes what is known about these sequences and signals. These identified sequences often are A+T rich, and thus are more likely to occur in an A+T rich coding sequence such as a B.t. gene. The sequence motif ATTTA (or AUUUA as it appears in RNA) has been implicated as a destabilizing sequence in mammalian cell mRNA (Shaw and Kamen, 1986). No analysis of the function of this sequence in plants has been done.



-3-

Many short lived mRNAs have A+T rich 3' untranslated regions, and these regions often have the ATTTA sequence, sometimes present in mutiple copies or as multimers (e.g., ATTTATTTA...). Shaw and Kamen showed that the transfer of the 3' end of an unstable mRNA to a stable RNA (globin or VA1) decreased the stable RNA's half life dramatically. They further showed that a pentamer of ATTTA had a profound destabilizing effect on a stable message, and that this signal could exert its effect whether it was located at the 3' end or within the coding sequence. However, the number of ATTTA sequences and/or the sequence context in which they occur also appear to be important in determining whether they function as destabilizing sequences. Shaw and Kamen showed that a trimer of ATTTA had much less effect than a pentamer on mRNA stability and a dimer or a monomer had no effect on stability (Shaw and Kamen, 1987). Note that multimers of ATTTA such as a pentamer automatically create an A+T rich region. This was shown to be a cytoplasmic effect, not nuclear. In other unstable mRNAs, the ATTTA sequence may be present in only a single copy, but it is often contained in an A+T rich region. From the animal cell data collected to date, it appears that ATTTA at least in some contexts is important in stability, but it is not yet possible to predict which occurences of ATTTA are destabiling elements or whether any of these effects are likely to be seen in plants.

Some studies on mRNA degradation in animal cells also indicate that RNA degradation may begin in some cases with nucleolytic attack in A+T rich regions. It

-4-

is not clear if these cleavages occur at ATTTA sequences. There are also examples of mRNAs that have differential stability depending on the cell type in which they are expressed or on the stage within the cell cycle at which they are expressed. For example, histone mRNAs are stable during DNA synthesis but unstable if DNA synthesis is disrupted. The 3' end of some histone mRNAs seems to be responsible for this effect (Pandey and Marzluff, 1987). It does not appear to be mediated by ATTTA, nor is it clear what controls the differential stability of this mRNA. Another example is the differential stability of IgG mRNA in B lymphocytes during B cell maturation (Genovese and Milcarek, 1988). A final example is the instability of a mutant beta-thalassaemic globin mRNA. In bone marrow cells, where this gene is normally expressed, the mutant mRNA is unstable, while the wild-type mRNA is stable. When the mutant gene is expressed in HeLa or L cells in vitro, the mutant mRNA shows no instability (Lim et al., 1988). These examples all provide evidence that mRNA stability can be mediated by cell type or cell cycle specific factors. Furthermore this type of instability is not yet associated with specific sequences. Given these uncertainties, it is not possible to predict which RNAs are likely to be unstable in a given cell. In addition, even the ATTTA motif may act differentially depending on the nature of the cell in which the RNA is present. Shaw and Kamen (1987) have reported that activation of protein kinase C can block degradation mediated by ATTTA.

-5-

The addition of a polyadenylate string to the 3' end is common to most eucaryotic mRNAs, both plant and animal. The currently accepted view of polyA addition is that the nascent transcript extends beyond the mature 3' terminus. Contained within this transcript are signals for polyadenylation and proper 3' end formation. This processing at the 3' end involves cleavage of the mRNA and addition of polyA to the mature 3' end. By searching for consensus sequences near the polyA tract in both plant and animal mRNAs, it has been possible to identify consensus sequences that apparently are involved in polyA addition and 3' end cleavage. The same consensus sequences seem to be important to both of these processes. These signals are typically a variation on the sequence AATAAA. In animal cells, some variants of this sequence that are functional have been identified; in plant cells there seems to be an extended range of functional sequences (Wickens and Stephenson, 1984; Dean et al., 1986). Because all of these consensus sequences are variations on AATAAA, they all are A+T rich sequences. This sequence is typically found 15 to 20 bp before the polyA tract in a mature mRNA. Experiments in animal cells indicate that this sequence is involved in both polyA addition and 3' maturation. Site directed mutations in this sequence can disrupt these functions (Conway and Wickens, 1988; Wickens et al., 1987). However, it has also been observed that sequences up to 50 to 100 bp 3' to the putative polyA signal are also required; i.e., a gene that has a normal AATAAA but has been replaced or disrupted

-6-

5 downstream does not get properly polyadenylated (Gil and Proudfoot, 1984; Sadofsky and Alwine, 1984; McDevitt et al., 1984). That is, the polyA signal itself is not sufficient for complete and proper processing. It is not yet known what specific downstream sequences are required in addition to the polyA signal, or if there is a specific sequence that has this function. Therefore, sequence analysis can  
10 only identify potential polyA signals.

In naturally occurring mRNAs that are normally polyadenylated, it has been observed that disruption of this process, either by altering the polyA signal or other sequences in the mRNA, profound effects can  
15 be obtained in the level of functional mRNA. This has been observed in several naturally occurring mRNAs, with results that are gene specific so far. There are no general rules that can be derived yet from the study of mutants of these natural genes, and no rules that can be applied to heterologous genes. Below are  
20 four examples:

1. In a globin gene, absence of a proper polyA site leads to improper termination of transcription. It is likely, but not proven, that the improperly  
25 terminated RNA is nonfunctional and unstable (Proudfoot et al., 1987).

2. In a globin gene, absence of a functional polyA signal can lead to a 100-fold decrease in the level of mRNA accumulation (Proudfoot et al., 1987).

3. A globin gene polyA site was placed into the  
30 3' ends of two different histone genes. The histone genes contain a secondary structure (stem-loop) near

-7-

their 3' ends. The amount of properly polyadenylated histone mRNA produced from these chimeras decreased as the distance between the stem-loop and the polyA site increased. Also, the two histone genes produced greatly different levels of properly polyadenylated mRNA. This suggests an interaction between the polyA site and other sequences on the mRNA that can modulate mRNA accumulation (Pandy and Marzluff, 1987).

4. The soybean leghemoglobin gene has been cloned into HeLa cells, and it has been determined that this plant gene contains a "cryptic" polyadenylation signal that is active in animal cells, but is not utilized in plant cells. This leads to the production of a new polyadenylated mRNA that is nonfunctional. This again shows that analysis of a gene in one cell type cannot predict its behavior in alternative cell types (Wiebauer et al., 1988).

From these examples, it is clear that in natural mRNAs proper polyadenylation is important in mRNA accumulation, and that disruption of this process can effect mRNA levels significantly. However, insufficient knowledge exists to predict the effect of changes in a normal gene. In a heterologous gene, where we do not know if the putative polyA sites (consensus sequences) are functional, it is even harder to predict the consequences. However, it is possible that the putative sites identified are disfunctional. That is, these sites may not act as proper polyA sites, but instead function as aberrant sites that give rise to unstable mRNAs.

-8-

5 In animal cell systems, AATAAA is by far the most  
 common signal identified in mRNAs upstream of the  
 polyA, but at least four variants have also been found  
 (Wickens and Stephenson, 1984). In plants, not nearly  
 so much analysis has been done, but it is clear that  
 multiple sequences similar to AATAAA can be used. The  
 plant sites below called major or minor refer only to  
 the study of Dean et al. (1986) which analyzed only  
 10 three types of plant gene. The designation of  
 polyadenylation sites as major or minor refers only to  
 the frequency of their occurrence as functional sites  
 in naturally occurring genes that have been analyzed.  
 In the case of plants this is a very limited database.  
 15 It is hard to predict with any certainty that a site  
 designated major or minor is more or less likely to  
 function partially or completely when found in a  
 heterologous gene such as *B.t.*

20	PA	AATAAA	Major consensus site
	P1A	AATAAT	Major plant site
	P2A	AACCAA	Minor plant site
	P3A	ATATAA	"
	P4A	AATCAA	"
	P5A	ATACTA	"
25	P6A	ATAAAA	"
	P7A	ATGAAA	"
	P8A	AAGCAT	"
	P9A	ATTAAT	"
	P10A	ATACAT	"
30	P11A	AAAATA	"

-9-

	P12A	ATTAAA	Minor animal site
	P13A	AATTAA	"
	P14A	AATACA	"
5	P15A	CATAAA	"

Another type of RNA processing that occurs in the nucleus is intron splicing. Nearly all of the work on intron processing has been done in animal cells, but  
10 some data is emerging from plants. Intron processing depends on proper 5' and 3' splice junction sequences. Consensus sequences for these junctions have been derived for both animal and plant mRNAs, but only a few nucleotides are known to be invariant. Therefore,  
15 it is hard to predict with any certainty whether a putative splice junction is functional or partially functional based solely on sequence analysis. In particular, the only invariant nucleotides are GT at the 5' end of the intron and AG at the 3' end of the intron. In plants, at every nearby position, either  
20 within the intron or in the exon flanking the intron, all four nucleotides can be found, although some positions show some nucleotide preference (Brown, 1986; Hanley and Schuler, 1988).

A plant intron has been moved from a patatin gene  
25 into a GUS gene. To do this, site directed mutagenesis was performed to introduce new restriction sites, and this mutagenesis changed several nucleotides in the intron and exon sequences flanking the GT and AG. This intron still functioned properly,  
30 indicating the importance of the GT and AG and the flexibility at other nucleotide positions. There are

-10-

of course many occurrences of GT and AG in all genes that do not function as intron splice junctions, so there must be some other sequence or structural features that identify splice junctions. In plants, one such feature appears to be base composition per se. Wiebauer et al. (1988) and Goodall et al. (1988) have analyzed plant introns and exons and found that exons have ~50% A+T while introns have ~70% A+T. Goodall et al. (1988) also created an artificial plant intron that has consensus 5' and 3' splice junctions and a random A+T rich internal sequence. This intron was spliced correctly in plants. When the internal segment was replaced by a G+C rich sequence, splicing efficiency was drastically reduced. These two examples demonstrate that intron recognition in plants may depend on very general features -- splice junctions that have a great deal of sequence diversity and A+T richness of the intron itself. This, of course, makes it difficult to predict from sequence alone whether any particular sequence is likely to function as an active or partially active intron for RNA processing.

B.t. genes being A+T rich contain numerous stretches of various lengths that have 70% or greater A+T. The number of such stretches identified by sequence analysis depends on the length of sequence scanned.

As for polyadenylation described above, there are complications in predicting what sequences might be utilized as splice sites in any given gene. First, many naturally occurring genes have alternative



-11-

splicing pathways that create alternative combinations of exons in the final mRNA (Gallega and Nadal-Ginard, 1988; Helfman and Ricci, 1988; Tsurushita and Korn, 1989). That is, some splice junctions are apparently recognized under some circumstances or in certain cell types, but not in others. The rules governing this are not understood. In addition, there can be an interaction between processing paths such that utilization of a particular polyadenylation site can interfere with splicing at a nearby splice site and vice versa (Adami and Nevins, 1988; Brady and Wold, 1988; Marzluff and Pandey, 1988). Again no predictive rules are available. Also, sequence changes in a gene can drastically alter the utilization of particular splice junctions. For example, in a bovine growth hormone gene, small deletions in an exon a few hundred bases downstream of an intron cause the splicing efficiency of the intron to drop from greater than 95% to less than 2% (essentially nonfunctional). Other deletions however have essentially no effect (Hampson and Rottman, 1988). Finally, a variety of in vitro and in vivo experiments indicate that mutations that disrupt normal splicing lead to rapid degradation of the RNA in the nucleus. Splicing is a multistep process in the nucleus and mutations in normal splicing can lead to blockades in the process at a variety of steps. Any of these blockades can then lead to an abnormal and unstable RNA. Studies of mutants of normally processed (polyadenylation and splicing) genes are relevant to the study of heterologous genes such as B.t. B.t. genes might

-12-

5 contain functional signals that lead to the production of aberrant nonfunctional mRNAs, and these mRNAs are likely to be unstable. But the *B.t.* genes are perhaps even more likely to contain signals that are analogous to mutant signals in a natural gene. As shown above these mutant signals are very likely to cause defects in the processing pathways whose consequence is to produce unstable mRNAs.

10 It is not known with any certainty what signals RNA transcription termination in plant or animal cells. Some studies on animal genes that indicate that stretches of sequence rich in T cause termination by calf thymus RNA polymerase II in vitro. These studies  
15 have shown that the 3' ends of in vitro terminated transcripts often lie within runs of T such as T5, T6 or T7. Other identified sites have not been composed solely of T, but have had one or more other nucleotides as well. Termination has been found to  
20 occur within the sequences TATTTTTT, ATTCTC, TTCTT (Dedrick et al., 1987; Reines et al., 1987). In the case of these latter two, the context in which the sequence is found has been C+T rich as well. It is not known if this is essential. Other studies have implicated stretches of A as potential transcriptional  
25 terminators. An interesting example from SV40 illustrates the uncertainty in defining terminators based on sequence alone. One potential terminator in SV40 was identified as being A rich and having a region of dyad symmetry (potential stem-loop) 5' to  
30 the A rich stretch. However, a second terminator identified experimentally downstream in the same gene

-13-

was not A rich and included no potential secondary structure (Kessler et al., 1988). Of course, due to the A+T content of B.t. genes, they are rich in runs of A or T that could act as terminators. The importance of termination to stability of the mRNA is shown by the globin gene example described above. Absence of a normal polyA site leads to a failure in proper termination with a consequent decrease in mRNA.

There is also an effect on mRNA stability due the translation of the mRNA. Premature translational termination in human triose phosphate isomerase leads to instability of the mRNA (Daar et al., 1988). Another example is the beta-thallemic globin mRNA described above that is specifically unstable in bone marrow cells (Lim et al., 1988). The defect in this mutant gene is a single base pair deletion at codon 44 that leads to translational termination (a nonsense codon) at codon 60. Compared to properly translated normal globin mRNA, this mutant RNA is very unstable. These results indicate that an improperly translated mRNA is unstable. Other work in yeast indicates that proper but poor translation can have an effect on mRNA levels. A heterologous gene was modified to convert certain codons to more yeast preferred codons. An overall 10-fold increase in protein production was achieved, but there was also about a 3-fold increase in mRNA (Hoekema et al., 1987). This indicates that more efficient translation can lead to greater mRNA stability, and that the effect of codon usage can be at the RNA level as well as the translational level. It is not clear from codon usage studies which codons

-14-

lead to poor translation, or how this is coupled to mRNA stability.

5 Therefore, it is an object of the present invention to provide a method for preparing synthetic plant genes which express their respective proteins at relatively high levels when compared to wild-type genes. It is yet another object of the present invention to provide synthetic plant genes which  
10 express the crystal protein toxin of *Bacillus thuringiensis* at relatively high levels.

#### BRIEF DESCRIPTION OF THE DRAWINGS

15 Figure 1 illustrates the steps employed in modifying a wild-type gene to increase expression efficiency in plants.

Figure 2 illustrates a comparison of the changes in the modified *B.t.k.* HD-1 sequence of Example 1 (lower line) versus the wild-type sequence of *B.t.k.* HD-1  
20 which encodes the crystal protein toxin (upper line).

Figure 3 illustrates a comparison of the changes in the synthetic *B.t.k.* HD-1 sequence of Example 2 (lower line) versus the wild-type sequence of *B.t.k.* HD-1 which encodes the crystal protein toxin (upper line).  
25

Figure 4 illustrates a comparison of the changes in the synthetic *B.t.k.* HD-73 sequence of Example 3 (lower line) versus the wild-type sequence of *B.t.k.* HD-73 (upper line).

30 Figure 5 represents a plasmid map of intermediate plant transformation vector cassette pMON893.

-15-

Figure 6 represents a plasmid map of intermediate plant transformation vector cassette pMON900.

5 Figure 7 represents a map for the disarmed T-DNA of *A. tumefaciens* ACO.

Figure 8 illustrates a comparison of the changes in the synthetic truncated *B.t.k.* HD-73 gene (Amino acids 29-615 with an N-terminal Met-Ala) of Example 3 (lower line) versus the wild-type sequence of *B.t.k.* HD-73  
10 (upper line).

Figure 9 illustrates a comparison of the changes in the synthetic/wild-type full length *B.t.k.* HD-73 sequence of Example 3 (lower line) versus the wild-type full-length sequence of *B.t.k.* HD-73 (upper line).  
15

Figure 10 illustrates a comparison of the changes in the synthetic/modified full length *B.t.k.* HD-73 sequence of Example 3 (lower line) versus the wild-type full-length sequence of *B.t.k.* HD-73 (upper line).  
20

Figure 11 illustrates a comparison of the changes in the fully synthetic full-length *B.t.k.* HD-73 sequence of Example 3 (lower line) versus the wild-type full-length sequence of *B.t.k.* HD-73 (upper line).  
25

Figure 12 illustrates a comparison of the changes in the synthetic *B.t.t.* sequence of Example 5 (lower line) versus the wild-type sequence of *B.t.t.* which encodes the crystal protein toxin (upper line).

Figure 13 illustrates a comparison of the changes  
30 in the synthetic *B.t.* P2 sequence of Example 6 (lower

-16-

line) versus the wild-type sequence of *B.t.k.* HD-1 which encodes the P2 protein toxin (upper line).

5 Figure 14 illustrates a comparison of the changes in the synthetic *B.t. entomocidus* sequence of Example 7 (lower line) versus the wild-type sequence of *B.t. entomocidus* which encodes the Btent protein toxin (upper line).

10 Figure 15 illustrates a plasmid map for plant expression cassette vector pMON744.

Figure 16 illustrates a comparison of the changes in the synthetic potato leaf roll virus (PLRV) coat protein sequence of Example 9 (lower line) versus the wild-type coat protein sequence of PLRV (upper line).

15

#### STATEMENT OF THE INVENTION

The present invention provides a method for preparing synthetic plant genes which genes express  
20 their protein product at levels significantly higher than the wild-type genes which were commonly employed in plant transformation heretofore. In another aspect, the present invention also provides novel synthetic plant genes which encode non-plant proteins.

25 For brevity and clarity of description, the present invention will be primarily described with respect to the preparation of synthetic plant genes which encode the crystal protein toxin of *Bacillus thuringiensis* (*B.t.*). Suitable *B.t.* subspecies include, but are not limited to, *B.t. kurstaki* HD-1, *B.t. kurstaki* HD-73,  
30 *B.t. sotto*, *B.t. berliner*, *B.t. thuringiensis*, *B.t. tolworthi*, *B.t. dendrolimus*, *B.t. alesti*, *B.t.*

-17-

galleriae, *B.t. aizawai*, *B.t. subtoxicus*, *B.t. entomocidus*, *B.t. tenebrionis* and *B.t. san diego*. However, those skilled in the art will recognize and it should be understood that the present method may be used to prepare synthetic plant genes which encode non-plant proteins other than the crystal protein toxin of *B.t.* as well as plant proteins (see for instance, Example 9).

The expression of *B.t.* genes in plants is problematic. Although the expression of *B.t.* genes in plants at insecticidal levels has been reported, this accomplishment has not been straightforward. In particular, the expression of a full-length lepidopteran specific *B.t.* gene (comprising DNA from a *B.t.k.* isolate) has been reported to be unsuccessful in yielding insecticidal levels of expression in some plant species (Vaeck et al., 1987 and Barton et al., 1987).

It has been reported that expression of the full-length gene from *B.t.k.* HD-1 was detectable in tomato plants but that truncated genes led to a higher frequency of insecticidal plants with an overall higher level of expression. Truncated genes of *B.t. berliner* also led to a higher frequency of insecticidal plants in tobacco (Vaeck et al., 1987). On the other hand, insecticidal plants were provided from lettuce transformants using a full-length gene.

It has also been reported that the full length gene from *B.t.k.* HD-73 gave some insecticidal effect in tobacco (Adang et al., 1987). However, the *B.t.* mRNA detected in these plants was only 1.7 kb compared to

-18-

the expected 3.7 kb indicating improper expression of the gene. It was suggested that this truncated mRNA was too short to encode a functional truncated toxin, but there must have been a low level of longer mRNA in some plants or no insecticidal activity would have been observed. Others have reported in a publication that they observed a large amount of shorter than expected mRNA from a truncated *B.t.k.* gene, but some mRNA of the expected size was also observed. In fact, it was suggested that expression of the full length gene is toxic to tobacco callus (Barton et al., 1987). The above illustrates that lepidopteran type *B.t.* genes are poorly expressed in plants compared to other chimeric genes previously expressed from the same promoter cassettes.

The expression of *B.t.t.* in tomato and potato is at levels similar to that of *B.t.k.* (i.e., poor). *B.t.t.* and *B.t.k.* genes share only limited sequence homology, but they share many common features in terms of base composition and the presence of particular A+T rich elements.

All reports in the field have noted the lower than expected expression of *B.t.* genes in plants. In general, insecticidal efficacy has been measured using insects very sensitive to *B.t.* toxin such as tobacco hornworm. Although it has been possible to obtain plants totally protected against tobacco hornworm, it is important to note that hornworm is up to 500 fold more sensitive to *B.t.* toxin than some agronomically important insect pests such as beet armyworm. It is therefore of interest to obtain transgenic plants that



-19-

are protected against all important lepidopteran pests (or against Colorado potato beetle in the case of *B.t. tenebrionis*), and in addition to have a level of *B.t.* expression that provides an additional safety margin over and above the efficacious protection level. It is also important to devise plant genes which function reproducibly from species to species, so that insect resistant plants can be obtained in a predictable fashion.

In order to achieve these goals, it is important to understand the nature of the poorer than expected expression of *B.t.* genes in plants. The level of stable *B.t.* mRNA in plants is much lower than expected. That is, compared to other coding sequences driven by the same promoter, the level of *B.t.* mRNA measured by Northern analysis or nuclease protection experiments is much lower. For example, tomato plant 337 (Fischhoff et al., 1987) was selected as the best expressing plant with pMON9711 which contains the *B.t.k.* HD-1 KpnI fragment driven by the CaMV 35S promoter and contains the NOS-NPTII-NOS selectable marker gene. In this plant the level of *B.t.* mRNA is between 100 to 1000 fold lower than the level of NPTII mRNA, even though the 35S promoter is approximately 50-fold stronger than the NOS promoter (Sanders et al., 1987).

The level of *B.t.* toxin protein detected in plants is consistent with the low level of *B.t.* mRNA. Moreover, the insecticidal efficacy of the transgenic plants correlates with the *B.t.* protein level indicating that the toxin protein produced in plants

-20-

is biologically active. Therefore, the low level of *B.t.* toxin expression may be the result of the low levels of *B.t.* mRNA.

5 Messenger RNA levels are determined by the rate of synthesis and rate of degradation. It is the balance between these two that determines the steady state level of mRNA. The rate of synthesis has been maximized by the use of the CaMV 35S promoter, a  
10 strong constitutive plant expressible promoter. The use of other plant promoters such as nopaline synthase (NOS), mannopine synthase (MAS) and ribulose biphosphatecarboxylase small subunit (RUBISCO) have not led to dramatic changes in the levels of *B.t.*  
15 toxin protein expression indicating that the effects determining *B.t.* toxin protein levels are promoter independent. These data imply that the coding sequences of DNA genes encoding *B.t.* toxin proteins are somehow responsible for the poor expression level, and that this effect is manifested by a low level of  
20 accumulated stable mRNA.

Lower than expected levels of mRNA have been observed with four different lepidopteran specific genes (two from *B.t.k.* HD-1; *B.t. berliner* and *B.t.k.* HD-73) as well as the gene from the coleopteran  
25 specific *B.t. tenebrionis*. It appears that for lepidopteran type *B.t.* genes these effects are manifest more strongly in the full length coding sequences than in the truncated coding sequences. These effects are seen across plant species although  
30 their magnitude seems greater in some plant species such as tobacco.

-21-

The nature of the coding sequences of *B.t.* genes distinguishes them from plant genes as well as many other heterologous genes expressed in plants. In particular, *B.t.* genes are very rich (~62%) in adenine (A) and thymine (T) while plant genes and most bacterial genes which have been expressed in plants are on the order of 45-55% A+T. The A+T content of the genomes (and thus the genes) of any organism are features of that organism and reflect its evolutionary history. While within any one organism genes have similar A+T content, the A+T content can vary tremendously from organism to organism. For example, some *Bacillus* species have among the most A+T rich genomes while some *Streptomyces* species are among the least A+T rich genomes (~30 to 35% A+T).

Due to the degeneracy of the genetic code and the limited number of codon choices for any amino acid, most of the "excess" A+T of the structural coding sequences of some *Bacillus* species are found in the third position of the codons. That is, genes of some *Bacillus* species have A or T as the third nucleotide in many codons. Thus A+T content in part can determine codon usage bias. In addition, it is clear that genes evolve for maximum function in the organism in which they evolve. This means that particular nucleotide sequences found in a gene from one organism, where they may play no role except to code for a particular stretch of amino acids, have the potential to be recognized as gene control elements in another organism (such as transcriptional promoters or terminators, polyA addition sites, intron splice

-22-

5 sites, or specific mRNA degradation signals). It is perhaps surprising that such misread signals are not a more common feature of heterologous gene expression, but this can be explained in part by the relatively homogeneous A+T content (~50%) of many organisms. This A+T content plus the nature of the genetic code put clear constraints on the likelihood of occurrence of any particular oligonucleotide sequence. Thus, a 10 gene from *E. coli* with a 50% A+T content is much less likely to contain any particular A+T rich segment than a gene from *B. thuringiensis*.

15 As described above, the expression of *B.t.* toxin protein in plants has been problematic. Although the observations made in other systems described above offer the hope of a means to elevate the expression level of *B.t.* toxin proteins in plants, the success obtained by the present method is quite unexpected. Indeed, inasmuch as it has been recently reported that 20 expression of the full-length *B.t.k.* toxin protein in tobacco makes callus tissue necrotic (Barton et al., 1987); one would reasonably expect that high level expression of *B.t.* toxin protein to be unattainable due to the reported toxicity effects.

25 In its most rigorous application, the method of the present invention involves the modification of an existing structural coding sequence ("structural gene") which codes for a particular protein by removal of ATTTA sequences and putative polyadenylation signals by site directed mutagenesis of the DNA 30 comprising the structural gene. It is most preferred that substantially all the polyadenylation signals and

-23-

ATTTA sequences are removed although enhanced expression levels are observed with only partial removal of either of the above identified sequences.

5 Alternately if a synthetic gene is prepared which codes for the expression of the subject protein, codons are selected to avoid the ATTTA sequence and putative polyadenylation signals. For purposes of the present invention putative polyadenylation signals

10 include, but are not necessarily limited to, AATAAA, AATAAT, AACCAA, ATATAA, AATCAA, ATACTA, ATAAAA, ATGAAA, AAGCAT, ATTAAT, ATACAT, AAAATA, ATTAAA, AATTAA, AATACA and CATAAA. In replacing the ATTTA sequences and polyadenylation signals, codons are

15 preferably utilized which avoid the codons which are rarely found in plant genomes.

Another embodiment of the present invention, represented in the flow diagram of Figure 1, employs a method for the modification of an existing structural

20 gene or alternately the *de novo* synthesis of a structural gene which method is somewhat less rigorous than the method first described above. Referring to Figure 1, the selected DNA sequence is scanned to identify regions with greater than four consecutive adenine (A) or thymine (T) nucleotides. The A+T

25 regions are scanned for potential plant polyadenylation signals. Although the absence of five or more consecutive A or T nucleotides eliminates most plant polyadenylation signals, if there are more than one of the minor polyadenylation signals identified

30 within ten nucleotides of each other, then the nucleotide sequence of this region is preferably

-24-

altered to remove these signals while maintaining the original encoded amino acid sequence.

5       The second step is to consider the 15 to 30  
nucleotide regions surrounding the A+T rich region  
identified in step one. If the A+T content of the  
surrounding region is less than 80%, the region should  
be examined for polyadenylation signals. Alteration  
of the region based on polyadenylation signals is  
10       dependent upon (1) the number of polyadenylation  
signals present and (2) presence of a major plant  
polyadenylation signal.

      The extended region is examined for the presence of  
plant polyadenylation signals. The polyadenylation  
15       signals are removed by site-directed mutagenesis of  
the DNA sequence. The extended region is also  
examined for multiple copies of the ATTTA sequence  
which are also removed by mutagenesis.

      It is also preferred that regions comprising many  
20       consecutive A+T bases or G+C bases are disrupted since  
these regions are predicted to have a higher  
likelihood to form hairpin structure due to self-  
complementarity. Therefore, insertion of  
heterogeneous base pairs would reduce the likelihood  
of self-complementary secondary structure formation  
25       which are known to inhibit transcription and/or  
translation in some organisms. In most cases, the  
adverse effects may be minimized by using sequences  
which do not contain more than five consecutive A+T or  
G+C.

30

-25-

SYNTHETIC OLIGONUCLEOTIDES FOR MUTAGENESIS

5       The oligonucleotides used in the mutagenesis are  
designed to maintain the proper amino acid sequence  
and reading frame and preferably to not introduce  
common restriction sites such as BglII, HindIII, SacI,  
KpnI, EcoRI, NcoI, PstI and SalI into the modified  
10    gene. These restriction sites are found in multi-  
linker insertion sites of cloning vectors such as  
plasmids pUC118 and pMON7258. Of course, the  
introduction of new polyadenylation signals, ATTTA  
sequences or consecutive stretches of more than five  
A+T or G+C, should also be avoided. The preferred  
15    size for the oligonucleotides is around 40-50 bases,  
but fragments ranging from 18 to 100 bases have been  
utilized. In most cases, a minimum of 5 to 8 base  
pairs of homology to the template DNA on both ends of  
the synthesized fragment are maintained to insure  
20    proper hybridization of the primer to the template.  
The oligonucleotides should avoid sequences longer  
than five base pairs A+T or G+C. Codons used in the  
replacement of wild-type codons should preferably  
avoid the TA or CG doublet wherever possible. Codons  
25    are selected from a plant preferred codon table (such  
as Table I below) so as to avoid codons which are  
rarely found in plant genomes, and efforts should be  
made to select codons to preferably adjust the G+C  
content to about 50%.

30

-26-

Table I

Preferred Codon Usage in Plants

5

<u>Amino Acid</u>	<u>Codon</u>	<u>Percent Usage in Plants</u>
10	ARG	CGA
		7
		CGC
		11
		CGG
		5
15		CGU
		25
		AGA
		29
		AGG
		23
20	LEU	CUA
		8
		CUC
		20
		CUG
		10
25		CUU
		28
		UUA
		5
		UUG
		30
30	SER	UCA
		14
		UCC
		26
		UCG
		3
		UCU
		21
		AGC
		21
		AGU
		15
	THR	ACA
		21
		ACC
		41
		ACG
		7
		ACU
		31
	PRO	CCA
		45
		CCC
		19
		CCG
		9
		CCU
		26
	ALA	GCA
		23
		GCC
		32
		GCG
		3
		GCU
		41



-27-

Table I - continued

Preferred Codon Usage in Plants

5	<u>Amino Acid</u>	<u>Codon</u>	<u>Percent Usage in Plants</u>
	GLY	GGA	32
		GGC	20
		GGG	11
		GGU	37
10	ILE	AUA	12
		AUC	45
		AUU	43
	VAL	GUA	9
		GUC	20
		GUG	28
15		GUU	43
	LYS	AAA	36
		AAG	64
	ASN	AAC	72
		AAU	28
20	GLN	CAA	64
		CAG	36
	HIS	CAC	65
		CAU	35
	GLU	GAA	48
25		GAG	52
	ASP	GAC	48
		GAU	52
	TYR	UAC	68
		UAU	32
30	CYS	UGC	78
		UGU	22

-28-

Table I - continued

Preferred Codon Usage in Plants

5	<u>Amino Acid</u>	<u>Codon</u>	<u>Percent Usage in Plants</u>
	PHE	UUC	56
		UUU	44
	MET	AUG	100
10	TRP	UGG	100

Regions with many consecutive A+T bases or G+C bases are predicted to have a higher likelihood to form hairpin structures due to self-complementarity. Disruption of these regions by the insertion of heterogeneous base pairs is preferred and should reduce the likelihood of the formation of self-complementary secondary structures such as hairpins which are known in some organisms to inhibit transcription (transcriptional terminators) and translation (attenuators). However, it is difficult to predict the biological effect of a potential hairpin forming region.

It is evident to those skilled in the art that while the above description is directed toward the modification of the DNA sequences of wild-type genes, the present method can be used to construct a completely synthetic gene for a given amino acid sequence. Regions with five or more consecutive A+T or G+C nucleotides should be avoided. Codons should be selected avoiding the TA and CG doublets in codons whenever possible. Codon usage can be normalized

-29-

against a plant preferred codon usage table (such as Table I) and the G+C content preferably adjusted to about 50%. The resulting sequence should be examined to ensure that there are minimal putative plant polyadenylation signals and ATTTA sequences. Restriction sites found in commonly used cloning vectors are also preferably avoided. However, placement of several unique restriction sites throughout the gene is useful for analysis of gene expression or construction of gene variants.

#### Plant Gene Construction

The expression of a plant gene which exists in double-stranded DNA form involves transcription of messenger RNA (mRNA) from one strand of the DNA by RNA polymerase enzyme, and the subsequent processing of the mRNA primary transcript inside the nucleus. This processing involves a 3' non-translated region which adds polyadenylate nucleotides to the 3' end of the RNA. Transcription of DNA into mRNA is regulated by a region of DNA usually referred to as the "promoter." The promoter region contains a sequence of bases that signals RNA polymerase to associate with the DNA and to initiate the transcription of mRNA using one of the DNA strands as a template to make a corresponding strand of RNA.

A number of promoters which are active in plant cells have been described in the literature. These include the nopaline synthase (NOS) and octopine synthase (OCS) promoters (which are carried on tumor-

-30-

inducing plasmids of *Agrobacterium tumefaciens*), the Cauliflower Mosaic Virus (CaMV) 19S and 35S promoters, the light-inducible promoter from the small subunit of ribulose bis-phosphate carboxylase (ssRUBISCO, a very abundant plant polypeptide) and the mannopine synthase (MAS) promoter (Velten et al. 1984 and Velten & Schell, 1985). All of these promoters have been used to create various types of DNA constructs which have been expressed in plants (see e.g., PCT publication WO84/02913 (Rogers et al., Monsanto)).

Promoters which are known or are found to cause transcription of RNA in plant cells can be used in the present invention. Such promoters may be obtained from plants or plant viruses and include, but are not limited to, the CaMV35S promoter and promoters isolated from plant genes such as ssRUBISCO genes. As described below, it is preferred that the particular promoter selected should be capable of causing sufficient expression to result in the production of an effective amount of protein.

The promoters used in the DNA constructs (i.e. chimeric plant genes) of the present invention may be modified, if desired, to affect their control characteristics. For example, the CaMV35S promoter may be ligated to the portion of the ssRUBISCO gene that represses the expression of ssRUBISCO in the absence of light, to create a promoter which is active in leaves but not in roots. The resulting chimeric promoter may be used as described herein. For purposes of this description, the phrase "CaMV35S" promoter thus includes variations of CaMV35S promoter,

-31-

e.g., promoters derived by means of ligation with operator regions, random or controlled mutagenesis, etc. Furthermore, the promoters may be altered to contain multiple "enhancer sequences" to assist in elevating gene expression.

The RNA produced by a DNA construct of the present invention also contains a 5' non-translated leader sequence. This sequence can be derived from the promoter selected to express the gene, and can be specifically modified so as to increase translation of the mRNA. The 5' non-translated regions can also be obtained from viral RNA's, from suitable eukaryotic genes, or from a synthetic gene sequence. The present invention is not limited to constructs, as presented in the following examples. Rather, the non-translated leader sequence can be part of the 5' end of the non-translated region of the coding sequence for the virus coat protein, or part of the promoter sequence, or can be derived from an unrelated promoter or coding sequence. In any case, it is preferred that the sequence flanking the initiation site conform to the translational consensus sequence rules for enhanced translation initiation reported by Kozak (1984).

The DNA construct of the present invention also contains a modified or fully-synthetic structural coding sequence which has been changed to enhance the performance of the gene in plants. In a particular embodiment of the present invention the enhancement method has been applied to design modified and fully synthetic genes encoding the crystal toxin protein of *Bacillus thuringiensis*. The structural genes of the

-32-

present invention may optionally encode a fusion protein comprising an amino-terminal chloroplast transit peptide or secretory signal sequence (see for instance, Examples 10 and 11).

The DNA construct also contains a 3' non-translated region. The 3' non-translated region contains a polyadenylation signal which functions in plants to cause the addition of polyadenylate nucleotides to the 3' end of the viral RNA. Examples of suitable 3' regions are (1) the 3' transcribed, non-translated regions containing the polyadenylation signal of *Agrobacterium* tumor-inducing (Ti) plasmid genes, such as the nopaline synthase (NOS) gene, and (2) plant genes like the soybean storage protein (7S) genes and the small subunit of the RuBP carboxylase (E9) gene. An example of a preferred 3' region is that from the 7S gene, described in greater detail in the examples below.

## Plant Transformation

A chimeric plant gene containing a structural coding sequence of the present invention can be inserted into the genome of a plant by any suitable method. Suitable plants for use in the practice of the present invention include, but are not limited to, soybean, cotton, alfalfa, oilseed rape, flax, tomato, sugarbeet, sunflower, potato, tobacco, maize, rice and wheat. Suitable plant transformation vectors include those derived from a Ti plasmid of *Agrobacterium tumefaciens*, as well as those disclosed, e.g., by

-33-

Herrera-Estrella (1983), Bevan (1983), Klee (1985) and EPO publication 120,516 (Schilperoort et al.). In addition to plant transformation vectors derived from the Ti or root-inducing (Ri) plasmids of *Agrobacterium*, alternative methods can be used to insert the DNA constructs of this invention into plant cells. Such methods may involve, for example, the use of liposomes, electroporation, chemicals that increase free DNA uptake, free DNA delivery via microprojectile bombardment, and transformation using viruses or pollen.

A particularly useful Ti plasmid cassette vector for transformation of dicotyledonous plants is shown in Figure 5. Referring to Figure 5, the expression cassette pMON893 consists of the enhanced CaMV35S promoter (EN 35S) and the 3' end including polyadenylation signals from a soybean gene encoding the alpha-prime subunit of beta-conglycinin. Between these two elements is a multilinker containing multiple restriction sites for the insertion of genes.

The enhanced CaMV35S promoter was constructed as follows. A fragment of the CaMV35S promoter extending between position -343 and +9 was previously constructed in pUC13 by Odell et al. (1985). This segment contains a region identified by Odell et al. (1985) as being necessary for maximal expression of the CaMV35S promoter. It was excised as a *Cla*I-*Hind*III fragment, made blunt ended with DNA polymerase I (Klenow fragment) and inserted into the *Hinc*II site of pUC18. This upstream region of the 35S promoter was excised from this plasmid as a *Hind*III-

-34-

5 EcoRV fragment (extending from -343 to -90) and inserted into the same plasmid between the HindIII and PstI sites. The enhanced CaMV35S promoter thus contains a duplication of sequences between -343 and -90 (Kay et al., 1987).

10 The 3' end of the 7S gene is derived from the 7S gene contained on the clone designated 17.1 (Schuler et al., 1982). This 3' end fragment, which includes the polyadenylation signals, extends from an AvaII site located about 30 bp upstream of the termination codon for the beta-conglycinin gene in clone 17.1 to an EcoRI site located about 450 bp downstream of this termination codon.

15 The remainder of pMON893 contains a segment of pBR322 which provides an origin of replication in *E. coli* and a region for homologous recombination with the disarmed T-DNA in *Agrobacterium* strain ACO (described below); the oriV region from the broad host range plasmid RK1; the streptomycin/spectinomycin resistance gene from Tn7; and a chimeric NPTII gene, containing the CaMV35S promoter and the nopaline synthase (NOS) 3' end, which provides kanamycin resistance in transformed plant cells.

25 Referring to Figure 6, transformation vector plasmid pMON900 is a derivative of pMON893. The enhanced CaMV35S promoter of pMON893 has been replaced with the 1.5kb mannopine synthase (MAS) promoter (Velten et al. 1984). The other segments are the same as plasmid pMON893. After incorporation of a DNA construct into plasmid vector pMON893 or pMON900, the intermediate vector is introduced into *A. tumefaciens*



-35-

strain ACO which contains a disarmed Ti plasmid. Cointegrate Ti plasmid vectors are selected and used to transform dicotyledonous plants.

5 Referring to Figure 7, *A. tumefaciens* ACO is a disarmed strain similar to pTiB6SE described by Fraley et al. (1985). For construction of ACO the starting *Agrobacterium* strain was the strain A208 which contains a nopaline-type Ti plasmid. The Ti plasmid  
10 was disarmed in a manner similar to that described by Fraley et al. (1985) so that essentially all of the native T-DNA was removed except for the left border and a few hundred base pairs of T-DNA inside the left border. The remainder of the T-DNA extending to a  
15 point just beyond the right border was replaced with a novel piece of DNA including (from left to right) a segment of pBR322, the oriV region from plasmid RK2, and the kanamycin resistance gene from Tn601. The pBR322 and oriV segments are similar to the segments  
20 in pMON893 and provide a region of homology for cointegrate formation.

The following examples are provided to better elucidate the practice of the present invention and should not be interpreted in any way to limit the scope of the present invention. Those skilled in the  
25 art will recognize that various modifications, truncations etc. can be made to the methods and genes described herein while not departing from the spirit and scope of the present invention.

30

-36-

Example 1 -- Modified B.t.k. HD-1 Gene

5 Referring to Figure 2, the wild-type B.t.k. HD-1  
 gene is known to be expressed poorly in plants as a  
 full length gene or as a truncated gene. The G+C  
 content of the B.t.k. gene is low (37%) containing  
 many A+T rich regions, potential polyadenylation sites  
 (18 sites; see Table II for the list of sequences)  
 10 and numerous ATTTA sequences.

Table II

15 List of Sequences of the Potential  
Polyadenylation Signals

	AATAAA*	AAGCAT
	AATAAT*	ATTAAT
	AACCAA	ATACAT
20	ATATAA	AAAATA
	AATCAA	ATTAAA**
	ATACTA	AATTAA**
	ATAAAA	AATACA**
	ATGAAA	CATAAA**

25 \* indicates a potential major plant polyadenylation  
 site.

\*\* indicates a potential minor animal polyadenylation  
 site.

All others are potential minor plant polyadenylation sites.

30

-37-

Table III lists the synthetic oligonucleotides designed and synthesized for the site-directed mutagenesis of the *B.t.k.* HD-1 gene.

5

Table III

Mutagenesis Primers for *B.t.k.* HD-1 Gene

10	<u>Primer</u>	<u>Length (bp)</u>	<u>Sequence</u>
	BTK185	18	TCCCCAGATA ATATCAAC
	BTK240	48	GGCTTGATTC CTAGCGAACT CTTCGATTCT CTGGTTGATG AGCTGTTC
15			
	BTK462	54	CAAACTGAG AGGTGGAGGT TGGCAGCTTG AACGTACACG GAGAGGAGAGGAAC
20			
	BTK669	48	AGTTAGTGTA AGCTCTCTTC TGAAGTGGTT GTACCTGATC CAATCTCT
25	BTK930	39	AGCCATGATC TGGTGACCGG ACCAGTAGTA TTCTCCTCT
	BTK1110	32	AGTTGTTGGT TGTTGATCCC GATGTTAAAA GG
30			

-38-

Table III - continued

Mutagenesis Primers for B.t.k. HD-1 Gene

5

<u>Primer</u>	<u>Length (bp)</u>	<u>Sequence</u>
BTK1380A	37	GTGATGAAGG GATGATGTTG TTGAACTCAG CACTACG
BTK1380T	100	CAGAAGTTCC AGAGCCAAGA TTAGTAGACT TGGTGAGTGG GATTTGGGTG ATTTGTGATG AAGGGATGAT GTTGTGAAC TCAGCACTAC GATGTATCCA
BTK1600	27	TGATGTGTGG AACTGAAGGT TTGTGGT

10

15

20

The B.t.k. HD-1 gene (BglII fragment from pMON9921 encoding amino acids 29-607 with a Met-Ala at the N-terminus) was cloned into pMON7258 (pUC118 derivative which contains a BglII site in the multilinker cloning region) at the BglII site resulting in pMON5342. The orientation of the B.t.k. gene was chosen so that the opposite strand (negative strand) was synthesized in filamentous phage particles for the mutagenesis. The procedure of Kunkle (1985) was used for the mutagenesis using plasmid pMON5342 as starting material.

25

30

The regions for mutagenesis were selected in the following manner. All regions of the DNA sequence of

-39-

the *B.t.k.* gene were identified which contained five or more consecutive base pairs which were A or T. These were ranked in terms of length and highest percentage of A+T in the surrounding sequence over a 20-30 base pair region. The DNA was then analysed for regions which might contain polyadenylation sites (see Table II above) or ATTTA sequences. Oligonucleotides were designed which maximized the elimination of A+T consecutive regions which contained one or more polyadenylation sites or ATTTA sequences. Two potential plant polyadenylation sites were rated more critical (see Table II) based on published reports. Codons were selected which increased G+C content, did not generate restriction sites for enzymes useful for cloning and assembly of the modified gene (BamHI, BglII, SacI, NcoI, EcoRV) and did not contain the doublets TA or GC which have been reported to be infrequently found in codons in plants. The oligonucleotides were at least 18 bp long ranging up to 100 base pairs and contained at least 5-8 base pairs of direct homology to native sequences at the ends of the fragments for efficient hybridization and priming in site-directed mutagenesis reactions. Figure 2 compares the wild-type *B.t.k.* HD-1 gene sequence with the sequence which resulted from the modifications by site-directed mutagenesis.

The end result of these changes was to increase the G+C content of *B.t.k.* gene from 37% to 41% while also decreasing the potential plant polyadenylation sites from 18 to 7 and decreasing the ATTTA regions from 13 to 7. Specifically, the mutagenesis changes from

-40-

amino (5') terminus to the carboxy (3') terminus are as follows:

5 BTK185 is an 18-mer used to eliminate a plant polyadenylation site in the midst of a nine base pair region of A+T.

10 BTK240 is a 48-mer. Seven base pairs were changed by this oligonucleotide to eliminate three potential polyadenylation sites (2 AACCAA, 1 AATTAA). Another region close to the region altered by BTK240, starting at bp 312, had a high A+T content (13 of 15 base pairs) and an ATTTA region. However, it did not contain a potential polyadenylation site and its longest string of uninterrupted A+T was seven base  
15 pairs.

20 BTK462 is a 54-mer introducing 13 base pair changes. The first six changes were to reduce the A+T richness of the gene by replacing wild-type codons with codons containing G and C while avoiding the CG doublet. The next seven changes made by BTK462 were used to eliminate an A+T rich region (13 of 14 base pairs were A or T) containing two ATTTA regions.

25 BTK669 is a 48-mer making nine individual base pair changes eliminating three possible polyadenylation sites (ATATAA, AATCAA, and AATTAA) and a single ATTTA site.

30 BTK930 is a 39-mer designed to increase the G+C content and to eliminate a potential polyadenylation site (AATAAT - a major site). This region did contain a nine base pair region of consecutive A+T sequence. One of the base pair changes was a G to A because a G at this position would have created a G+C rich region

-41-

(CCGG(G)C). Since sequencing reactions indicate that there can be difficulties generating sequence through G+C consecutive bases, it was thought to be prudent to avoid generating potentially problematic regions even if they were problematic only in vitro.

BTK1110 is a 32-mer designed to introduce five changes in the wild-type gene. One potential site (AATAAT - a major site) was eliminated in the midst of an A+T rich region (19 of 22 base pairs).

BTK1380A and BTK1380T are responsible for 14 individual base pair changes. The first region (1380A) has 17 consecutive A+T base pairs. In this region is an ATTTA and a potential polyadenylation site (AATAAT). The 100-mer (1380T) contains all the changes dictated by 1380A. The large size of this primer was in part an experiment to determine if it was feasible to utilize large oligonucleotides for mutagenesis (over 60 bases in length). A second consideration was that the 100-mer was used to mutagenize a template which had previously been mutagenized by 1380A. The original primer ordered to mutagenize the region downstream and adjacent to 1380A did not anneal efficiently to the desired site as indicated by an inability to obtain clean sequence utilizing the primer. The large region of homology of 1380T did assure proper annealing. The extended size of 1380T was more of a convenience rather than a necessity. The second region adjacent to 1380A covered by 1380T has a high A+T content (22 of 29 bases are A or T).

-42-

5 BTK1600 is a 27-mer responsible for five individual base pair changes. An ATTTA region and a plant polyadenylation site were identified and the appropriate changes engineered.

10 A total of 62 bases were changed by site-directed mutagenesis. The G+C content increased by 55 base pairs, the potential polyadenylation sites were reduced from 18 to seven and the ATTTA sequences decreased from 13 to seven. The changes in the DNA sequence resulted in changes in 55 of the 579 codons in the truncated *B.t.k.* gene in pMON5342 (approximately 9.5%).

15 Referring to Table IV modified *B.t.k.* HD-1 genes were constructed that contained all of the above modifications (pMON5370) or various subsets of individual modifications. These genes were inserted into pMON893 for plant transformation and tobacco plants containing these genes were analyzed. The analysis of tobacco plants with the individual modifications was undertaken for several reasons. Expression of the wild type truncated gene in tobacco is very poor, resulting in infrequent identification of plants toxic to THW. Toxicity is defined by leaf feeding assays as at least 60% mortality of tobacco hornworm neonate larvae with a damage rating of 1 or less (scale is 0 to 4; 0 is equivalent to total protection, 4 total damage). The modified HD-1 gene (pMON5370) shows a large increase in expression (estimated to be approximately 100-fold; see Table 25 VIII) in tobacco. Therefore, increases in expression 30 of the wild-type gene due to individual modifications



-43-

would be apparently a large increase in the frequency of toxic tobacco plants and the presence of detectable B.t.k. protein. Results are shown in the following table:

Table IV

Relative effects of Regional Modifications  
within the B.t.k. Gene

	<u>Construct</u>	<u>Position Modified</u>	<u># of Plants</u>	<u># of Toxic Plants</u>
15	pMON5370	185,240,669,930, 1110,1380a+b,1600	38	22
	pMON10707	185,240,462,669	48	19
	pMON10706	930,1110,1380a+b,1600	43	1
20	pMON10539	185	55	2
	pMON10537	240	57	17
25	pMON10540	185,240	88	23
	pMON10705	462	47	1

The effects of each individual oligonucleotides' changes on expression did reveal some overall trends.

Six different constructs were generated which were

-44-

designed to identify the key regions. The nine different oligonucleotides were divided in half by their position on the gene. Changes in the N-terminal half were incorporated into pMON10707 (185,240,462,669). C-terminal half changes were incorporated into pMON10706 (930,1110,1380a+b,1600). The results of analysis of plants with these two constructs indicate that pMON10707 produces a substantial number of toxic plants (19 of 48). Protein from these plants is detectable by ELISA analysis. pMON10706 plants were rarely identified as insecticidal (1 of 43) and the levels of B.t.k. were barely detectable by immunological analysis. Investigation of the N-terminal changes in greater detail was done with 4 pMON constructs; 10539 (185 alone), 10537 (240 alone), 10540 (185 and 240) and 10705 (462 alone). The results indicate that the presence of the changes in 240 were required to generate a substantial number of toxic plants (pMON10540; 23 of 88, pMON10537; 17 of 57). The absence of the 240 changes resulted in a low frequency of toxic plants with low B.t.k. protein levels, identical to results with the wild type gene. These results indicate that the changes in 240 are responsible for a substantial increase in B.t.k. expression levels over an analogous wild-type construct in tobacco. Changes in additional regions (185,462,669) in conjunction with 240 may result in increases in B.t.k. expression (>2 fold). However, changes at the 240 region of the N-terminal portion of the gene do result in dramatic increases in expression.

-45-

Despite the importance of the alteration of the 240 region in expression of modified genes, increased expression can be achieved by alteration of other regions. Hybrid genes, part wild-type, part synthetic, were generated to determine the effects of synthetic gene segments on the levels of *B.t.k.* expression. A hybrid gene was generated with a synthetic N-terminal third (base pair 1 to 590 of Figure 2: to the *Xba*I site) with the C-terminal wild type *B.t.k.* HD-1 (pMON5378). Plants transformed with this vector were as toxic as plants transformed with the modified HD-1 gene (pMON5370). This is consistent with the alteration of the 240 region. However, pMON10538, a hybrid with a wild-type N-terminal third (wild type gene for the first 600 base pairs, to the second *Xba*I site) and a synthetic C-terminal last two-thirds (base pair 590 to 1845 of Figure 3 was used to transform tobacco and resulted in a dramatic increase in expression. The levels of expression do not appear to be as high as those seen with the synthetic gene, but are comparable to the modified gene levels. These results indicate that modification of the 240 segment is not essential to increased expression since pMON10538 has an intact 240 region. A fully synthetic gene is, in most cases, superior for expression levels of *B.t.k.* (See Example 2.)

#### Example 2 -- Fully Synthetic *B.t.k.* HD-1 Gene

A synthetic *B.t.k.* HD-1 gene was designed using the preferred plant codons listed in Table V below.

-46-

5 Table V lists the codons and frequency of use in plant  
genes of dicotyledonous plants compared to the  
frequency of their use in the wild type *B.t.k.* HD-1  
gene (amino acids 1-615) and the synthetic gene of  
this example. The total number of each amino acid in  
this segment of the gene is listed in the parenthesis  
under the amino acid designated.

10

15

20

25

30

5

30

-48-

Table V - continued

5      Codon in Usage Synthetic B.t.k. HD-1 Gene

<u>Amino Acid</u>		<u>Codon</u>	<u>Percent Usage in</u> <u>Plants/Wt B.t.k./Syn</u>		
10	THR (42)	ACA	21	31	14
		ACC	41	19	53
		ACG	7	14	0
		ACU	31	36	33
15	PRO (34)	CCA	45	35	53
		CCC	19	6	12
		CCG	9	21	3
		CCU	26	38	32
20	ALA (31)	GCA	23	38	26
		GCC	32	9	29
		GCG	3	3	0
		GCU	41	50	45
25	GLY (46)	GGA	32	52	45
		GGC	20	17	15
		GGG	11	15	6
		GGU	37	15	34
30	ILE (46)	AUA	12	39	2
		AUC	45	11	67
		AUU	43	50	30

-49-

Table V - continued

5      Codon in Usage Synthetic B.t.k. HD-1 Gene

	<u>Amino Acid</u>	<u>Codon</u>	<u>Percent Usage in Plants/Wt B.t.k./Syn</u>		
10	VAL	GUA	9	45	3
	(38)	GUC	20	5	16
		GUG	28	11	37
		GUU	43	39	45
15	LYS	AAA	36	100	33
	(3)	AAG	64	0	67
	ASN	AAC	72	27	80
	(44)	AAU	28	73	20
20	GLN	CAA	64	77	61
	(31)	CAG	36	23	39
	HIS	CAC	65	0	80
	(10)	CAU	35	100	20
25	GLU	GAA	48	87	50
	(30)	GAG	52	13	50
	ASP	GAC	48	17	65
	(23)	GAU	52	83	35
30					

-50-

Table V - continued

Codon in Usage Synthetic B.t.k. HD-1 Gene

	<u>Amino Acid</u>	<u>Codon</u>	<u>Percent Usage in</u>		
			<u>Plants/Wt B.t.k./Syn</u>		
10					
	TYR	UAC	68	20	72
	(25)	UAU	32	80	28
	CYS	UGC	78	50	100
15	(2)	UGU	22	50	0
	PHE	UUC	56	17	83
	(36)	UUU	44	83	17
20	MET	AUG	100	100	100
	(9)				
	TRP	UGG	100	100	100
	(9)				

25

The resulting synthetic gene lacks ATTTA sequences, contains only one potential polyadenylation site and has a G+C content of 48.5%. Figure 3 is a comparison of the wild-type HD-1 sequence to the synthetic gene sequence for amino acids 1-615. There is approximately 77% DNA homology between the synthetic

30



-51-

gene and the wild-type gene and 356 of the 615 codons have been changed (approximately 60%).

5       Example 3 -- Synthetic B.t.k. HD-73 Gene

10       The crystal protein toxin from B.t.k. HD-73 exhibits a higher unit activity against some important agricultural pests. The toxin protein of HD-1 and HD-73 exhibit substantial homology (~90%) in the N-terminal 450 amino acids, but differ substantially in the amino acid region 451-615. Fusion proteins comprising amino acids 1-450 of HD-1 and 451-615 of HD-73 exhibit the insecticidal properties of the wild-type HD-73. The strategy employed was to use the 5'-two thirds of the synthetic HD-1 gene (first 1350 bases, up to the SacI site) and to dramatically modify the final 590 bases (through amino acid 645) of the HD-73 in a manner consistent with the algorithm used to design the synthetic HD-1 gene. Table VI below lists the oligonucleotides used to modify the HD-73 gene in the order used in the gene from 5' to 3' end. Nine oligonucleotides were used in a 590 base pair region, each nucleotide ranging in size from 33 to 60 bases. The only regions left unchanged were areas where there were no long consecutive strings of A or T bases (longer than six). All polyadenylation sites and ATTTA sites were eliminated.

30

-52-

Table VI

<u>Mutagenesis Primers for B.t.k. HD-73</u>			
5	<u>Primer</u>	<u>Length (bp)</u>	<u>Sequence</u>
10	73K1363	51	AATACTATCG GATGCGATGA TGTTGTTGAA CTCAGCACTA CGGTGTATCC A
	73K1437	33	TCCTGAAATG ACAGAACCGT TGAAGAGAAA GTT
15	73K1471	48	ATTTCCACTG CTGTTGAGTC TAACGAGGTC TCCACCAGTG AATCCTGG
20	73K1561	60	GTGAATAGGG GTCACAGAAG CATACTCAC ACGAACTCTA TATCTGGTAG ATGTTGGATGG
	73K1642	33	TGTAGCTGGA ACTGTATTGG AGAAGATGGA TGA
25	73K1675	48	TTCAAAGTAA CCGAAATCGC TGGATTGGAG ATTATCCAAG GAGGTAGC
30	73K1741	39	ACTAAAGTTT CTAACACCCA CGATGTTACC GAGTGAAGA

-53-

Table VI - continued

Mutagenesis Primers for B.t.k. HD-73

5

<u>Primer</u>	<u>Length (bp)</u>	<u>Sequence</u>
73K1797	36	AACTGGAATG AACTCGAATC TGTCGATAAT CACTCC
73KTERM	54	GGACACTAGA TCTTAGTGAT AATCGGTCAC ATTTGTCTTG AGTCCAAGCT GGT

10

15 The resulting gene has two potential polyadenylation sites (compared to 18 in the WT) and no ATTTA sequence (12 in the WT). The G+C content has increased from 37% to 48%. A total of 59 individual base pair changes were made using the primers in

20 Table VI. Overall, there is 90% DNA homology between the region of the HD-73 gene modified by site directed mutagenesis and the wild-type sequence of the analogous region of HD-73. The synthetic HD-73 is a hybrid of the first 1360 bases from the synthetic HD-1 and the next 590 bases or so modified HD-73 sequence.

25 Figure 4 is a comparison of the above-described synthetic B.t.k. HD-73 and the wild-type B.t.k. HD-73 encoding amino acids 1-645. In the modified region of the HD-73 gene 44 of the 170 codons (25%) were changed as a result of the site-directed mutagenesis changes

30 resulting from the oligonucleotides found in Table VI. Overall, approximately 50% of the codons in the

-54-

synthetic *B.t.k.* HD-73 differ from the analogous segment of the wild-type and HD-73 gene.

5 A one base pair deletion in the synthetic HD-73 gene was detected in the course of sequencing the 3' end at base pair 1890. This results in a frame-shift mutation at amino acid 625 with a premature stop codon at amino acid 640 (pMON5379). Table VII below compares the codon usage of the wild-type gene of *B.t.k.* HD-73  
10 versus the synthetic gene of this example for amino acids 451-645 and codon usage of naturally occurring genes of dicotyledonous plants. The total number of each amino acid encoded in this segment of the gene is found in the parentheses under the amino acid  
15 designation.

Table VII  
Codon Usage in Synthetic *B.t.k.* HD-73 Gene

20	<u>Amino Acid</u>	<u>Codon</u>	<u>Percent Usage in</u>		
			<u>Plants/Wt HD-73/Syn</u>		
	ARG	CGA	7	10	0
	(10)	CGC	11	0	8
		CGG	5	10	0
		CGU	25	20	23
25		AGA	29	60	62
		AGG	23	0	8

30

-55-

Table VII - continued  
Codon Usage in Synthetic B.t.k. HD-73 Gene

5	<u>Amino Acid</u>	<u>Codon</u>	<u>Percent Usage in</u> <u>Plants/Wt HD-73/Syn</u>		
10	LEU (12)	CUA	8	25	8
		CUC	20	17	58
		CUG	10	17	8
		CUU	28	8	0
		UUA	5	33	8
		UUG	30	0	17
15	SER (21)	UCA	14	24	18
		UCC	26	10	27
		UCG	3	10	0
		UCU	21	24	18
		AGC	21	0	14
		AGU	15	33	23
20	THR (15)	ACA	21	47	38
		ACC	41	13	31
		ACG	7	13	0
		ACU	31	27	31
25	PRO (7)	CCA	45	71	71
		CCC	19	0	0
		CCG	9	14	0
		CCU	26	14	29

30

-56-

Table VII - continued  
Codon Usage in Synthetic B.t.k. HD-73 Gene

5	<u>Amino Acid</u>	<u>Codon</u>	<u>Percent Usage in</u>		
			<u>Plants/Wt HD-73/Syn</u>		
10	ALA	GCA	23	29	31
	(14)	GCC	32	7	8
		GCG	3	21	15
		GCU	41	43	46
15	GLY	GGA	32	33	43
	(15)	GGC	20	0	0
		GGG	11	27	14
		GGU	37	40	43
20	ILE	AUA	12	33	7
	(15)	AUC	45	7	40
		AUU	43	60	53
	VAL	GUA	9	40	7
25	(15)	GUC	20	0	7
		GUG	28	20	36
		GUU	43	40	50
	LYS	AAA	36	67	100
30	(3)	AAG	64	33	0
	ASN	AAC	72	20	53
	(20)	AAU	28	80	47

-57-

Table VII - continued  
Codon Usage in Synthetic B.t.k. HD-73 Gene

5	<u>Amino Acid</u>	<u>Codon</u>	<u>Percent Usage in</u>		
			<u>Plants/Wt HD-73/Syn</u>		
	GLN	CAA	64	60	67
	(5)	CAG	36	40	33
10	HIS	CAC	65	67	100
	(3)	CAU	35	33	0
	GLU	GAA	48	86	57
	(7)	GAG	52	14	43
15	ASP	GAC	48	40	50
	(5)	GAU	52	60	50
	TYR	UAC	68	0	20
20	(5)	UAU	32	100	80
	CYS	UGC	78	0	0
	(0)	UGU	22	0	0
	PHE	UUC	56	8	67
25	(13)	UUU	44	92	33
	MET	AUG	100	100	100
	(2)				
30	TRP	UGG	100	100	100
	(2)				

-58-

Another truncated synthetic HD-73 gene was constructed. The sequence of this synthetic HD-73 gene is identical to that of the above synthetic HD-73 gene in the region in which they overlap (amino acids 29-615), and it also encodes Met-Ala at the N-terminus. Figure 8 shows a comparison of this truncated synthetic HD-73 gene with the N-terminal Met-Ala versus the wild-type HD-73 gene.

While the previous examples have been directed at the preparation of synthetic and modified genes encoding truncated *B.t.k.* proteins, synthetic or modified genes can also be prepared which encode full length toxin proteins.

One full length *B.t.k.* gene consists of the synthetic HD-73 sequence of Figure 4 from nucleotide 1-1845 plus wild-type HD-73 sequence encoding amino acids 616 to the C-terminus of the native protein. Figure 9 shows a comparison of this synthetic/wild-type full length HD-73 gene versus the wild-type full length HD-73 gene.

Another full length *B.t.k.* gene consists of the synthetic HD-73 sequence of Figure 4 from nucleotide 1-1845 plus a modified HD-73 sequence ending amino acids 616 to the C-terminus of the native protein. The C-terminal portion has been modified by site-directed mutagenesis to remove putative polyadenylation signals and ATTTA sequences according to the algorithm of Figure 1. Figure 10 shows a comparison of this synthetic/modified full length HD-73 gene versus the wild-type full length HD-73 gene.



-59-

Another full length *B.t.k.* gene consists of a fully synthetic HD-73 sequence which incorporates the synthetic HD-73 sequence of Figure 4 from nucleotide 1-1845 plus a synthetic sequence encoding amino acids 616 to the C-terminus of the native protein. The C-terminal synthetic portion has been designed to eliminate putative polyadenylation signals and ATTTA sequences and to include plant preferred codons. Figure 11 shows a comparison of this fully synthetic full length HD-73 gene versus the wild-type full length HD-73 gene.

Alternatively, another full length *B.t.k.* gene consists of a fully synthetic sequence comprising base pairs 1-1830 of *B.t.k.* HD-1 (Figure 3) and base pairs 1834-3534 of *B.t.k.* HD-73 (Figure 11).

Example 4 -- Expression of Modified and Synthetic *B.t.k.* HD-1 and Synthetic HD-73

A number of plant transformation vectors for the expression of *B.t.k.* genes were constructed by incorporating the structural coding sequences of the previously described genes into plant transformation cassette vector pMON893. The respective intermediate transformation vector is inserted into a suitable disarmed *Agrobacterium* vector such as *A. tumefaciens* ACO, supra. Tissue explants are cocultured with the disarmed *Agrobacterium* vector and plants regenerated under selection for kanamycin resistance using known protocols: tobacco (Horsch et al., 1985); tomato

-60-

(McCormick et al., 1986) and cotton (Trolinder et al., 1987).

5 a) Tobacco.

The level of *B.t.k.* HD-1 protein in transgenic tobacco plants containing pMON9921 (wild type truncated), pMON5370 (modified HD-1, Example 1, Figure 2) and pMON5377 (synthetic HD-1, Example 2, Figure 3) were analyzed by Western analysis. Leaf tissue was frozen in liquid nitrogen, ground to a fine powder and then ground in a 1:2 (wt:volume) of SDS-PAGE sample buffer. Samples were frozen on dry ice, then incubated for 10 minutes in a boiling water bath and microfuged for 10 minutes. The protein concentration of the supernatant was determined by the method of Bradford (Anal. Biochem. 72:248-254). Fifty ug of protein was run per lane on 9% SDS-PAGE gels, the protein transferred to nitrocellulose and the *B.t.k.* HD-1 protein visualized using antibodies produced against *B.t.k.* HD-1 protein as the primary antibody and alkaline phosphatase conjugated second antibody as described by the manufacturer (Promega, Madison, WI). Purified HD-1 tryptic fragment was used as the control. Whereas the *B.t.k.* protein from tobacco plants containing pMON9921 was below the level of detection, the *B.t.k.* protein from plants containing the modified (pMON5370) and synthetic (pMON5377) genes was easily detected. The *B.t.k.* protein from plants containing pMON9921 remained undetectable, even with 10 fold longer incubation times. The relative levels of *B.t.k.* HD-1 protein in these plants is estimated in

-61-

Table VIII. Because the protein from plants containing pMON9921 was not observed, the level of protein in these plants was estimated from the relative mRNA levels (see below). Plants containing the modified gene (pMON5370) expressed approximately 100 fold more *B.t.k.* protein than plants containing the wild-type gene (pMON9921). Plants containing the fully synthetic *B.t.k.* HD-1 gene (pMON5377) expressed approximately five fold more protein than plants containing the modified gene. The modified gene contributes the majority of the increase in *B.t.k.* expression observed. The plants used to generate the above data are the best representatives from each construct based either on a tobacco hornworm bioassay or on data derived from previous Western analysis.

Table VIII

Expression of *B.t.k.* HD-1 Protein  
in Transgenic Tobacco

Gene Description	Vector	<i>B.t.k.</i> Protein* Concentration	Fold Increase in <i>B.t.k.</i> Expression
Wild type	pMON9921	10	1
Modified	pMON5370	1000	100
Synthetic	pMON5377	5000	500

\* *B.t.k.* protein concentrations are expressed in ng/mg of total soluble protein. The level of *B.t.k.*

-62-

protein for plants containing the wild type gene are estimated from mRNA levels.

5       Plants containing these genes were tested for bioactivity to determine whether the increased quantities of protein observed by Western analysis result in a corresponding increase in bioactivity. Leaves from the same plants used for the Western data  
10 in Table 1 were tested for bioactivity against two insects. A detached leaf bioassay was first done using tobacco hornworm, an extremely sensitive lepidopteran insect. Leaves from all three transgenic tobacco plants were totally protected and 100%  
15 mortality of tobacco hornworm observed (see Table IX below). A much less sensitive insect, beet armyworm, was then used in another detached leaf bioassay. Beet armyworm is approximately 500 fold less sensitive to *B.t.k.* HD-1 protein than tobacco hornworm. The  
20 difference in sensitivity of these two insects was determined using purified HD-1 protein in a diet incorporation assay (see below). Plants containing the wild-type gene (pMON9921) showed only minimal protection against beet armyworm, whereas plants containing the modified gene showed almost complete  
25 protection and plants containing the fully synthetic gene were totally protected against beet armyworm damage. The results of these bioassays confirm the levels of *B.t.k.* HD-1 expression observed in the Western analysis and demonstrates that the increased  
30 levels of *B.t.k.* HD-1 protein correlates with increased insecticidal activity.

-63-

Table IX

5

Protection of Tobacco Plants from  
Tobacco Hornworm and Beet Armyworm

Gene		Tobacco Hornworm	Beet Armyworm
<u>Description</u>	<u>Vector</u>	<u>Damage*</u>	<u>Damage*</u>
None	None	NL	NL
Wild type	pMON9921	0	3
Modified	pMON5370	0	1
Synthetic	pMON5377	0	0

15

\* Extent of insect damage was rated: 0, no damage; 1, slight; 2, moderate; 3, severe; or NL, no leaf left.

20

25

30

The bioactivity of the *B.t.k.* HD-1 protein produced by these transgenic plants was further investigated to more accurately quantitate the relative activities. Leaf tissue from tobacco plants containing the wild-type, modified and synthetic genes were ground in 100 mM sodium carbonate buffer, pH 10 at a 1:2 (wt:vol) ratio. Particulate material was removed by centrifugation. The supernatant was incorporated into a synthetic diet similar to that described by Marrone et al. (1985). The diet medium was prepared the day of the test with the plant extract solutions incorporated in place of the 20% water component. One ml of the diet was aliquoted into 96 well plates.

-64-

After the diet dried, one neonate tobacco budworm larva was added to each well. Sixteen insects were tested with each plant sample. The plants were incubated at 27°C. After seven days, the larvae from each treatment were combined and weighed on an analytical balance. The average weight per insect was calculated and compared to a standard curve relating *B.t.k.* protein concentrations to average larval weight. Insect weight was inversely proportional (in a logarithmic manner) to the relative increase in *B.t.k.* protein concentration. The amount of *B.t.k.* HD-1 protein, based on the extent of larval growth inhibition was determined for two different plants containing each of the three genes. The specific activity (ng of *B.t.k.* HD-1 per mg of plant protein) was determined for each plant. Plants containing the modified HD-1 gene (pMON5370) averaged approximately 1400 ng (1200 and 1600 ng) of *B.t.k.* HD-1 per mg of plant extract protein. This value compares closely with the 1000 ng of *B.t.k.* HD-1 protein per mg of plant extract protein as determined by Western analysis (Table I). *B.t.k.* HD-1 concentrations for the plants containing the synthetic HD-1 gene averaged approximately 8200 ng (7200 and 9200 ng) of *B.t.k.* HD-1 protein per mg of plant extract protein. This number compares well to the 5000 ng of HD-1 protein per mg of plant extract protein estimated by Western analysis. Likewise, plants containing the synthetic gene showed approximately a six-fold higher specific activity than the corresponding plants containing the modified gene for these bioassays. In the Western

-65-

analysis the ratio was approximately 10 fold, again both are in good agreement. The level of *B.t.k.* protein in plants containing the wild-type HD-1 gene (pMON9921) was too low to give a significant decrease in larval weight and hence was below a level that could be quantitated in this assay. In conclusion, the levels of *B.t.k.* HD-1 protein determined by both the bioassays and the Western analysis for these plants containing the modified and synthetic genes agree, which demonstrates that the *B.t.k.* HD-1 protein produced by these plants is biologically active.

The levels of mRNA were determined in the plants containing the wild-type *B.t.k.* HD-1 gene (pMON9921) and the modified gene (pMON5370) to establish whether the increased levels of protein production result from increased transcription or translation. mRNA from plants containing the synthetic gene could not be analyzed directly with the same DNA probe as used for the wild-type and modified genes because of the numerous changes made in the coding sequence. mRNA was isolated and hybridized with a single-stranded DNA probe homologous to approximately the 5' 90 bp of the wild-type or modified gene coding sequences. The hybrids were digested with S1 nuclease and the protected probe fragments analyzed by gel electrophoresis. Because the procedure used a large excess of probe and long hybridization time, the amount of protected probe is proportional to the amount of *B.t.k.* mRNA present in the sample. Two plants expressing the modified gene (pMON5370) were

-66-

found to produce up to ten-fold more RNA than a plant expressing the wild-type gene (pMON9921).

5       The increased mRNA level from the modified gene is  
consistent with the result expected from the  
modifications introduced into this gene. However,  
this 10 fold increase in mRNA with the modified gene  
10 compared to the wild-type gene is in contrast to the  
100 fold increase in *B.t.k.* protein from these genes  
in tobacco plants. If the two mRNAs were equally well  
translated then a 10 fold increase in stable mRNA  
would be expected to yield a 10 fold increase in  
protein. The higher increase in protein indicates  
15 that the modified gene mRNA is translated at about a  
10 fold higher efficiency than wild-type. Thus, about  
half of the total effect on gene expression can be  
explained by changes in mRNA levels and about half to  
changes in translational efficiency. This increase in  
translational efficiency is striking in that only  
20 about 9.5% of the codons have been changed in the  
modified gene; that is, this effect is clearly not due  
to wholesale codon usage changes. The increased  
translational efficiency could be due to changes in  
mRNA secondary structure that affect translation or to  
the removal of specific translational blockades due to  
25 specific codons that were changed.

The increased expression seen with the synthetic HD-  
1 gene was also seen with a synthetic HD-73 gene in  
tobacco. *B.t.k.* HD-73 was undetected in extracts of  
tobacco plants containing the wild-type truncated HD-  
30 73 gene (pMON5367), whereas *B.t.k.* HD-73 protein was  
easily detected in extracts from tobacco plants



-67-

containing the synthetic HD-73 gene of Figure 4 (pMON5383). Approximately 1000 ng of *B.t.k.* HD-73 protein was detected per mg of total soluble plant protein.

As described in Example 3 above, the *B.t.k.* HD-73 protein encoded in pMON5383 contains a small C-terminal extension of amino acids not encoded in the wild-type HD-73 protein. These extra amino acids had no effect on insect toxicity or on increased plant expression. A second synthetic HD-73 gene was constructed as described in Example 3 (Figure 8) and used to transform tobacco (pMON5390). Analysis of plants containing pMON5390 showed that this gene was expressed at levels comparable to that of pMON5383 and that these plants had similar insecticidal efficacy.

In tobacco plants the synthetic HD-1 gene was expressed at approximately a 5-fold higher level than the synthetic HD-73 gene. However, this synthetic HD-73 gene still was expressed at least 100-fold better than the wild-type HD-73 gene. The HD-73 protein is approximately 5-fold more toxic to many insect pests than the HD-1 protein, so both synthetic HD-1 and HD-73 genes provide approximately comparable insecticidal efficacy in tobacco.

The full length *B.t.k.* HD-73 genes described in Example 3 were also incorporated into the plant transformation vector pMON893 so that they were expressed from the *En* 35S promoter. The synthetic/wild-type full length HD-73 gene of Figure 9 was incorporated into pMON893 to create pMON10505. The synthetic/modified full length HD-73 gene of

-68-

Figure 10 was incorporated into pMON893 to create pMON10526. The fully synthetic HD-73 gene of Figure 11 was incorporated into pMON893 to create pMON10518. These vectors were used to obtain transformed tobacco plants, and the plants were analyzed for insecticidal efficacy and for *B.t.k.* HD-73 protein levels by Western blot or ELISA immunoassay.

Tobacco plants containing all three of these full length *B.t.k.* genes produced detectable *B.t.k.* protein and showed 100% mortality of tobacco hornworm. This result is surprising in light of previous reported attempts to express the full length *B.t.k.* genes in transgenic plants. Vaeck et al. (1987) reported that a full length *B.t.k. berliner* gene similar to our HD-1 gene could not be detectably expressed in tobacco. Barton et al. (1987) reported a similar result for another full length gene from *B.t.k.* HD-1 (the so called 4.5 kb gene), and further indicated that tobacco callus containing this gene became necrotic, indicating that the full length gene product was toxic to plant cells. Fischhoff et al. (1987) reported that the full length *B.t.k.* HD-1 gene in tomato was poorly expressed compared to a truncated gene, and no plants that were fully toxic to tobacco hornworm could be recovered. All three of the above reports indicated much higher expression levels and recovery of toxic plants if the respective *B.t.k.* genes were truncated. Adang et al. reported that the full length HD-73 gene yielded a few tobacco plants with some biological activity (none were highly toxic) against hornworm and barely detectable *B.t.k.* protein. It was also noted

-69-

by them that the major *B.t.k.* mRNA in these plants was a truncated 1.7 kb species that would not encode a functional toxin. This indicated improper expression of the gene in tobacco. In contrast to all of these reports, the three full length *B.t.k.* HD-73 genes described above all lead to relatively high levels of protein and high levels of insect toxicity.

*B.t.k.* protein and mRNA levels in tobacco plants are shown in Table X for these three vectors. As can be seen from the table, the synthetic/wild-type gene (pMON10506) produces *B.t.k.* protein as about 0.01% of total soluble protein; the synthetic/modified gene produces *B.t.k.* as about 0.02% of total soluble protein; and the fully synthetic gene produces *B.t.k.* as about 0.2% of total soluble protein. *B.t.k.* mRNA was analyzed in these plants by Northern blot analysis using the common 5' synthetic half of the genes as a probe. As shown in Table X, the increased protein levels can largely be attributed to increased mRNA levels. Compared to the truncated modified and synthetic genes, this could indicate that the major contributors to increased translational efficiency are in the 5' half of the gene while the 3' half of the gene contains mostly determinants of mRNA stability. The increased protein levels also indicate that increasing the amount of the full length gene that is synthetic or modified increases *B.t.k.* protein levels. Compared to the truncated synthetic *B.t.k.* HD-73 genes (pMON5383 or pMON5390), the fully synthetic gene (pMON10518) produces as much or slightly more *B.t.k.* protein demonstrating that the full length genes are

-70-

capable of being expressed at high levels in plants. These tobacco plants with high levels of full length HD-73 protein show no evidence of abnormality and are  
5 fully fertile. The *B.t.k.* protein levels in these plants also produce the expected levels of insect toxicity based on feeding studies with beet armyworm or diet incorporation assays of plant extracts with tobacco budworm. The *B.t.k.* protein detected by  
10 Western blot analysis in these tobacco plants often contains a varying amount of protein of about 80 kDa which is apparently a proteolytic fragment of the full length protein. The C-terminal half of the full length protein is known to be proteolytically  
15 sensitive, and similar proteolytic fragments are seen from the full length gene in *E. coli* and *B.t.* itself. These fragments are fully insecticidal. The Northern analysis indicated that essentially all of the mRNA from these full length genes was of the expected full  
20 length size. There is no evidence of truncated mRNAs that could give rise to the 80 kDa protein fragment. In addition, it is possible that the fragment is not present in intact plant cells and is merely due to proteolysis during extraction for immunoassay.

25

30

-71-

Table X

Full Length B.t.k. HD-73 Protein and  
mRNA Levels in Transgenic Tobacco Plants

5

Gene	Vector	B.t.k. protein concentration	Relative B.t.k. mRNA level
description			
Synthetic/wild type	pMON10506	>100	0.5
10 Synthetic/modified	pMON10526	400	1
Fully synthetic	pMON10518	>2000	40

15

Thus, there is no serious impediment to producing high levels of *B.t.k.* HD-73 protein in plants from synthetic genes, and this is expected to be true of other full length lepidopteran active genes such as *B.t.k.* HD-1 or *B.t. entomocidus*. The fully synthetic *B.t.k.* HD-1 gene of Example 3 has been assembled in plant transformation vectors such as pMON893.

20

25

30

The fully synthetic gene in pMON10518 was also utilized in another plant vector and analyzed in tobacco plants. Although the CaMV35S promoter is generally a high level constitutive promoter in most plant tissues, the expression level of genes driven the CaMV35S promoter is low in floral tissue relative to the levels seen in leaf tissue. Because the economically important targets damaged by some insects are the floral parts or derived from floral parts (e.g., cotton squares and bolls, tobacco buds, tomato buds and fruit), it may be advantageous to increase the expression of *B.t.* protein in these tissues over that obtained with the CaMV35S promoter.

-72-

The 35S promoter of Figwort Mosaic Virus (FMV) is analogous to the CaMV35S promoter. This promoter has been isolated and engineered into a plant transformation vector analogous to pMON893. Relative to the CaMV promoter, the FMV 35S promoter is highly expressed in the floral tissue, while still providing similar high levels of gene expression in other tissues such as leaf. A plant transformation vector, pMON10517, was constructed in which the full length synthetic *B.t.k.* HD-73 gene of Figure 11 was driven by the FMV 35S promoter. This vector is identical to pMON10518 of Example 3 except that the FMV promoter is substituted for the CaMV promoter. Tobacco plants transformed with pMON10517 and pMON10518 were obtained and compared for expression of the *B.t.k.* protein by Western blot or ELISA immunoassay in leaf and floral tissue. This analysis showed that pMON10517 containing the FMV promoter expressed the full length HD-73 protein at higher levels in floral tissue than pMON10518 containing the CaMV promoter. Expression of the full length *B.t.k.* HD-73 protein from pMON10517 in leaf tissue is comparable to that seen with the most highly expressing plants containing pMON10518. However, when floral tissue was analyzed, tobacco plants containing pMON10518 that had high levels of *B.t.k.* protein in leaf tissue did not have detectable *B.t.k.* protein in the flowers. On the other hand, flowers of tobacco plants containing pMON10517 had levels of *B.t.k.* protein nearly as high as the levels in leaves at approximately 0.05% of total soluble protein. This analysis showed that the FMV promoter

-73-

could be used to produce relatively high levels of  
B.t.k. protein in floral tissue compared to the CaMV  
5 promoter.

b) Tomato.

The wild-type, modified and synthetic B.t.k. HD-1  
genes tested in tobacco were introduced into other  
plants to demonstrate the broad utility of this  
10 invention. Transgenic tomatoes were produced which  
contain these three genes. Data show that the  
increased expression observed with the modified and  
synthetic gene in tobacco also extends to tomato.  
Whereas the B.t.k. HD-1 protein is only barely  
15 detectable in plants containing the wild type HD-1  
gene (pMON9921), B.t.k. HD-1 was readily detected and  
the levels determined for plants containing the  
modified (pMON5370) or synthetic (pMON5377) genes.  
Expression levels for the plants containing the wild-  
20 type, modified and synthetic HD-1 genes were  
approximately 10, 100 and 500 ng per mg of total plant  
extract see Table XI below). The increase in B.t.k.  
HD-1 protein for the modified gene accounted for the  
majority of increase observed; 10 fold higher than the  
plants containing the wild-type gene, compared to only  
25 an additional five-fold increase for plants containing  
the synthetic gene. Again the site-directed changes  
made in the modified gene are the major contributors  
to the increased expression of B.t.k. HD-1.

30

-74-

Table XI

B.t.k. HD-1 Expression in  
Transgenic Tomato Plants

5

Gene	Vector	B.t.k. Protein* Concentration	Fold Increase in B.t.k. Expression
<u>Description</u>	<u>Vector</u>	<u>Concentration</u>	<u>Expression</u>
Wild type	pMON9921	10	1
Modified	pMON5370	100	10
Synthetic	pMON5377	500	50

10

15 \* B.t.k. HD-1 protein concentrations are expressed in ng/mg of total soluble plant protein. Data for plants containing the wild-type gene are estimates from mRNA levels and protein levels determined by ELISA.

20 These differences in B.t.k. HD-1 expression were confirmed with bioassays against tobacco hornworm and beet armyworm. Leaves from tomato plants containing each of these genes controlled tobacco hornworm damage and produced 100% mortality. With beet armyworm, leaves from plants containing the wild-type HD-1 gene (pMON9921) showed significant damage, leaves from plants containing the modified gene (pMON5370) showed less damage and leaves from plants containing the synthetic gene (pMON5377) were completely protected (see Table XII below).

25

30



-75-

Table XII

5                    Protection of Tomato Plants from  
                     Tobacco Hornworm and Beet Armyworm

	Gene Description	Vector	Tobacco Hornworm	Beet Armyworm
			Damage*	Damage*
10	None	None	NL	NL
	Wild type	pMON9921	0	3
	Modified	pMON5370	0	1
	Synthetic	pMON5377	0	0

15    \* Damage was rated as shown in Table IX.

The generality of the synthetic gene approach was extended in tomato with a synthetic *B.t.k.* HD-73 gene.

20    In tomato, extracts from plants containing the wild-type truncated HD-73 gene (pMON5367) showed no detectable HD-73 protein. Extracts from plants containing the synthetic HD-73 gene (pMON5383) showed high levels of *B.t.k.* HD-73 protein, approximately 2000 ng per mg of plant extract protein. These data clearly demonstrate that the changes made in the synthetic HD-73 gene lead to dramatic increases in the expression of the HD-73 protein in tomato as well as in tobacco

25    In contrast to tobacco, the synthetic HD-73 gene in tomato is expressed at approximately 4-fold to 5-fold higher levels than the synthetic HD-1 gene. Because  
 30    the HD-73 protein is about 5-fold more active than the

-76-

5 HD-1 protein against many insect pests including  
Heliothis species, the increased expression of  
synthetic HD-73 compared to synthetic HD-1 corresponds  
to about a 25-fold increased insecticidal efficacy in  
tomato.

10 In order to determine the mechanisms involved in  
the increased expression of modified and synthetic  
B.t.k. HD-1 genes in tomato, S1 nuclease analysis of  
mRNA levels from transformed tomato plants was  
performed. As indicated above, a similar analysis had  
been performed with tobacco plants, and this analysis  
showed that the modified gene produced up to 10-fold  
more mRNA than the wild-type gene. The analysis in  
15 tomato utilized a different DNA probe that allowed the  
analysis of wild-type (pMON9921), modified (pMON5370)  
and synthetic (pMON5377) HD-1 genes with the same  
probe. This probe was derived from the 5'  
untranslated region of the CaMV35S promoter in pMON893  
20 that was common to all three of these vectors  
(pMON9921, pMON5370 and pMON5377). This S1 analysis  
indicated that B.t.k. mRNA levels from the modified  
gene were 3 to 5 fold higher than for the wild-type  
gene, and that mRNA levels for the synthetic gene were  
about 2 to 3 fold higher than for the modified gene.  
25 Three independent transformants were analyzed for each  
gene. Compared to the fold increases in B.t.k. HD-1  
protein from these genes in tomato shown in Table XI,  
these mRNA increases can explain about half of the  
total protein increase as was seen in tobacco for the  
30 wild-type and modified genes. For tomato the total  
mRNA increase from wild-type to synthetic is about 6

-77-

to 15 fold compared to a protein increase of about 50 fold. This result is similar to that seen for tobacco in comparing the wild-type and modified genes, and it  
5 extends to the synthetic gene as well. That is, about half of the total fold increase in *B.t.k.* protein from wild-type to modified genes can be explained by mRNA increases and about half to enhanced translational efficiency. The same is also true in comparing the  
10 modified gene to the synthetic gene. Although there is an additional increase in RNA levels, this mRNA increase can explain only about half of the total protein increase.

The full length *B.t.k.* genes described above were  
15 also used to transform tomato plants and these plants were analyzed for *B.t.k.* protein and insecticidal efficacy. The results of this analysis are shown in Table XIII. Plants containing the synthetic/wild-type gene (pMON10506) produce the *B.t.k.* HD-73 protein at  
20 levels of about 0.01% of their total soluble protein. Plants containing the synthetic/modified gene (pMON10526) produce about 0.04% *B.t.k.* protein, and plants containing the fully synthetic gene (pMON10518) produce about 0.2% *B.t.k.* protein. These results are  
25 very similar to the tobacco plant results for the same genes. mRNA levels estimated by Northern blot analysis in tomato also increase in parallel with the protein level increase. As for tobacco with these three genes, most of the protein increase can be  
30 attributed to increased mRNA with a small component of translational efficiency increase indicated for the fully synthetic gene. The highest levels of full

-78-

length *B.t.k.* protein (from pMON10518) are comparable to or just slightly lower than the highest levels observed for the truncated HD-73 genes (pMON5383 and pMON5390). Tomato plants expressing these full length genes have the insecticidal activity expected for the observed protein levels as determined by feeding assays with beet armyworm or by diet incorporation of plant extracts with tobacco hornworm.

Table XIII

Full Length *B.t.k.* HD-73 Protein and  
mRNA Levels in Transgenic Tomato Plants

Gene description	Vector	<i>B.t.k.</i> protein concentration	Relative <i>B.t.k.</i> mRNA level
Synthetic/wild type	pMON10506	100	1
Synthetic/modified	pMON10526	400	2-4
Fully synthetic	pMON10518	2000	10

c) Cotton.

The generality of the increased expression of *B.t.k.* HD-1 and *B.t.k.* HD-73 by use of the modified and synthetic genes was extended to cotton. Transgenic calli were produced which contain the wild type (pMON9921) and the synthetic HD-1 (pMON5377) genes. Here again the *B.t.k.* HD-1 protein produced from calli containing the wild-type gene was not detected, whereas calli containing the synthetic HD-1

-79-

gene expressed the HD-1 protein at easily detectable levels. The HD-1 protein was produced at approximately 1000 ng/mg of plant calli extract protein. Again, to ensure that the protein produced by the transgenic cotton calli was biologically active and that the increased expression observed with the synthetic gene translated to increased biological activity, extracts of cotton calli were made in similar manner as described for tobacco plants, except that the calli was first dried between Whatman filter paper to remove as much of the water as possible. The dried calli were then ground in liquid nitrogen and ground in 100 mM sodium carbonate buffer, pH 10. Approximately 0.5 ml aliquotes of this material was applied to tomato leaves with a paint brush. After the leaf dried, five tobacco hornworm larvae were applied to each of two leaf samples. Leaves painted with extract from control calli were completely destroyed. Leaves painted with extract from calli containing the wild-type HD-1 gene (pMON9921) showed severe damage. Leaves painted with extract from calli containing the synthetic HD-1 gene (pMON5377) showed no damage (see Table XIV below).

25

30

-80-

Table XIV

5     Protection against Tobacco Hornworm by Tomato Leaves  
       Painted with Extracts Prepared from Cotton Calli  
       Containing a Control, the Wild-Type B.t.k. HD-1 Gene,  
       Synthetic HD-1 Gene or Synthetic HD-73 Gene

10	Gene	Tobacco Hornworm	
	<u>Description</u>	<u>Vector</u>	<u>Damage*</u>
	Control	Control	NL
	Wild type HD-1	pMON9921	3
	Synthetic HD-1	pMON5377	0
15	Synthetic HD-73	pMON5383	0

\* Damage was rated as shown in Table VIII.

20     Cotton calli were also produced containing another  
       synthetic gene, a gene encoding B.t.k. HD-73. The  
       preparation of this gene is described in Example 3.  
       Calli containing the synthetic HD-73 gene produced the  
       corresponding HD-73 protein at even higher levels than  
       the calli which contained the synthetic HD-1 gene.  
       Extracts made from calli containing the HD-73  
       25     synthetic gene (pMON5383) showed complete control of  
       tobacco hornworm when painted onto tomato leaves as  
       described above for extracts containing the HD-1  
       protein. (See Table XIV).

30     Transgenic cotton plants containing the synthetic  
       B.t.k. HD-1 gene (pMON5377) or the synthetic B.t.k. HD-  
       73 gene (pMON5383) have also been examined. These

-81-

5 plants produce the HD-1 or HD-73 proteins at levels comparable to that seen in cotton callus with the same genes and comparable to tomato and tobacco plants with these genes. For either synthetic truncated HD-1 or HD-73 genes, cotton plants expressing *B.t.k.* protein at 1000 to 2000 ng/mg total protein (0.1% to 0.2%) were recovered at a high frequency. Insect feeding assays were performed with leaves from cotton plants  
10 expressing the synthetic HD-1 or HD-73 genes. These leaves showed no damage (rating of 0) when challenged with larvae of cabbage looper (*Trichoplusia ni*), and only slight damage when challenged with larvae of beet armyworm (*Spodoptera exigua*). Damage ratings are as defined in Table VIII above. This demonstrated that  
15 cotton plants as well as calli expressed the synthetic HD-1 or HD-73 genes at high levels and that those plants were protected from damage by Lepidopteran insect larvae.

20 Transgenic cotton plants containing either the synthetic truncated HD-1 gene (pMON5377) or the synthetic truncated HD-73 gene (pMON5383) were also assessed for protection against cotton bollworm at the whole plant level in the greenhouse. This is a more realistic test of the ability of these plants to  
25 produce an agriculturally acceptable level of control. The cotton bollworm (*Heliothis zea*) is a major pest of cotton that produces economic damage by destroying terminals, squares and bolls, and protection of these fruiting bodies as well as the leaf tissue will be  
30 important for effective insect control and adequate crop protection. To test the protection afforded to

-82-

whole plants, R1 progeny of cotton plants expressing high levels of either *B.t.k.* HD-1 (pMON5377) or *B.t.k.* HD-73 (pMON5383) were assayed by applying 10-15 eggs of cotton bollworm per boll or square to the 20 uppermost squares or bolls on each plant. At least 12 plants were analyzed per treatment. The hatch rate of the eggs was approximately 70%. This corresponds to very high insect pressure compared to numbers of larvae per plant seen under typical field conditions. Under these conditions 100% of the bolls on control cotton plants were destroyed by insect damage. For the transgenics, significant boll protection was observed. Plants containing pMON5377 (HD-1) had 70-75% of the bolls survive the intense pressure of this assay. Plants containing pMON5383 (HD-73) had 80% to 90% boll protection. This is likely to be a consequence of the higher activity of HD-73 protein against cotton bollworm compared to HD-1 protein. In cases where the transgenic plants were damaged by the insects, the surviving larvae were delayed in their development by at least one instar.

Therefore, the increased expression obtained with the modified and synthetic genes is not limited to any one crop; tobacco, tomato and cotton calli and cotton plants all showed drastic increases in *B.t.k.* expression when the plants/calli were produced containing the modified or synthetic genes. Likewise, the utility of changes made to produce the modified and synthetic *B.t.k.* HD-1 gene is not limited to the HD-1 gene. The synthetic HD-73 gene in all three species also showed drastic increases in expression.



-83-

In summary, it has been demonstrated that: (1) the genetic changes made in the HD-1 modified gene lead to very significant increases in *B.t.k.* HD-1 expression;

5  
10 further five-fold increase in *B.t.k.* HD-1 expression; (3) the changes incorporated into the modified HD-1 gene accounted for the majority of the increased *B.t.k.* expression observed with the synthetic gene; (4) the increased expression was demonstrated in three different plants -- tobacco plants, tomato plants and cotton calli and cotton plants; (5) the increased expression as observed by Western analysis also correlated with similar increases in bioactivity, showing that the *B.t.k.* HD-1 proteins produced were comparably active; (6) when the method of the present invention used to design the synthetic HD-1 gene was employed to design a synthetic HD-73 gene it also was expressed at much higher levels in tobacco, tomato and cotton than the wild-type equivalent gene with consequent increases in bioactivity; (7) a fully synthetic full length *B.t.k.* gene was expressed at levels comparable to synthetic truncated genes.

25 Example 5 -- Synthetic *B.t. tenebrionis* Gene in Tobacco, Tomato and Potato

Referring to Figure 12, a synthetic gene encoding a Coleopteran active toxin is prepared by making the indicated changes in the wild-type gene of *B.t. tenebrionis* or de novo synthesis of the synthetic

30

-84-

an intermediate plant transformation vector such as pMON893: Plasmid pMON893 containing the synthetic B.t.t. gene is then inserted into a suitable disarmed Agrobacterium strain such as *A. tumefaciens* ACO.

#### Transformation and Regeneration of Potato

Sterile shoot cultures of Russet Burbank are maintained in vials containing 10 ml of PM medium (Murashige and Skoog (MS) inorganic salts, 30 g/l sucrose, 0.17 g/l  $\text{NaH}_2\text{PO}_4\cdot\text{H}_2\text{O}$ , 0.4 mg/l thiamine-HCl, and 100 mg/l myo-inositol, solidified with 1 g/l Gelrite at pH 6.0). When shoots reached approximately 5 cm in length, stem internode segments of 7-10 mm are excised and smeared at the cut ends with a disarmed *Agrobacterium tumefaciens* vector containing the synthetic B.t.t. gene from a four day old plate culture. The stem explants are co-cultured for three days at 23°C on a sterile filter paper placed over 1.5 ml of a tobacco cell feeder layer overlaid on 1/10 P medium (1/10 strength MS inorganic salts and organic addenda without casein as in Jarret et al. (1980), 30 g/l sucrose and 8.0 g/l agar). Following co-culture the explants are transferred to full strength P-1 medium for callus induction, composed of MS inorganic salts, organic additions as in Jarret et al. (1980) with the exception of casein, 3.0 mg/l benzyladenine (BA), and 0.01 mg/l naphthaleneacetic acid (NAA) (Jarret, et al., 1980). Carbenicillin (500 mg/l) is included to inhibit bacterial growth, and 100 mg/l

-85-

kanamycin is added to select for transformed cells. After four weeks the explants are transferred to medium of the same composition but with 0.3 mg/l gibberellic acid (GA3) replacing the BA and NAA (Jarret et al., 1981) to promote shoot formation. Shoots begin to develop approximately two weeks after transfer to shoot induction medium; these are excised and transferred to vials of PM medium for rooting. Shoots are tested for kanamycin resistance conferred by the enzyme neomycin phosphotransferase II, by placing a section of the stem onto callus induction medium containing MS organic and inorganic salts, 30 g/l sucrose, 2.25 mg/l BA, 0.186 mg/l NAA, 10 mg/l GA3 (Webb, et al., 1983) and 200 mg/l kanamycin to select for transformed cells.

The synthetic *B.t.t.* gene described in figure 12, was placed into a plant expression vector as described in example 5. The plasmid has the following characteristics; a synthetic *Bgl*II fragment having approximately 1800 base pairs was inserted into pMON893 in such a manner that the enhanced 35S promoter would express the *B.t.t.* gene. This construct, pMON1982, was used to transform both tobacco and tomato. Tobacco plants, selected as kanamycin resistant plants were screened with rabbit anti-*B.t.t.* antibody. Cross-reactive material was detected at levels predicted to be suitable to cause mortality to CPB. These target insects will not feed on tobacco, but the transgenic tobacco plants do demonstrate that the synthetic gene does improve expression of this protein to detectable levels.

-86-

Tomato plants with the pMON1982 construct were determined to produce *B.t.t.* protein at levels insecticidal to CPB. In initial studies, the leaves of four plants (5190, 5225, 5328 and 5133) showed little or no damage when exposed to CPB larvae (damage rating of 0-1 on a scale of 0 to 4 with 4 as no leaf remaining). Under these conditions the control leaves were completely eaten. Immunological analysis of these plants confirmed the presence of material cross-reactive with anti-*B.t.t.* antibody. Levels of protein expression in these plants were estimated at approximately 1 to 5 ng of *B.t.t.* protein in 50 ug of total extractable protein. A total of 17 tomato plants (17 of 65 tested) have been identified which demonstrate protection of leaf tissue from CPB (rating of 0 or 1) and show good insect mortality.

Results similar to those seen in tobacco and tomato with pMON1982 were seen with pMON1984 in the same plant species. pMON1984 is identical to pMON1982 except that the synthetic protease inhibitor (CMTI) is fused upstream of the native proteolytic cleavage site. Levels of expression in tobacco were estimated to be similar to pMON1982, between 10-15 ng per 50ug of total soluble protein.

Tomato plants expressing pMON1984 have been identified which protect the leaves from ingestion by CPB. The damage rating was 0 with 100% insect mortality.

Potato was transformed as described in example 5 with a vector similar to pMON1982 containing the enhanced CaMV35S/synthetic *B.t.t.* gene. Leaves of

-87-

potato plants transformed with this vector, were screened by CPB insect bioassay. Of the 35 plants tested, leaves from 4 plants, 16a, 13c, 13d, and 23a were totally protected when challenged. Insect bioassays with leaves from three other plants, 13e, 1a, and 13b, recorded damage levels of 1 on a scale of 0 to 4 with 4 being total devastation of the leaf material. Immunological analysis confirmed the presence of *B.t.t.* cross-reactive material in the leaf tissue. The level of *B.t.t.* protein in leaf tissue of plant 16a (damage rating of 0) was estimated at 20-50 ng of *B.t.t.* protein/50 ug of total soluble protein. The levels of *B.t.t.* protein seen in 16a tissue was consistent with its biological activity. Immunological analysis of 13e and 13b (tissue which scored 1 in damage rating) reveal less protein (5-10 ng/50 ug of total soluble protein) than in plant 16a. Cuttings of plant 16a were challenged with 50 to 200 eggs of CPB in a whole plant assay. Under these conditions 16a showed no damage and 100% mortality of insects while control potato plants were heavily damaged.

#### Example 6 -- Synthetic *B.t.k.* P2 Protein Gene

The P2 protein is a distinct insecticidal protein produced by some strains of *B.t.* including *B.t.k.* HD-1. It is characterized by its activity against both lepidopteran and dipteran insects (Yamamoto and Iizuka, 1983). Genes encoding the P2 protein have been isolated and characterized (Donovan et al.,

-88-

1988). The P2 proteins encoded by these genes are approximately 600 amino acids in length. These proteins share only limited homology with the  
5 lepidopteran specific P1 type proteins, such as the *B.t.k.* HD-1 and HD-73 proteins described in previous examples.

The P2 proteins have substantial activity against a variety of lepidopteran larvae including cabbage  
10 looper, tobacco hornworm and tobacco budworm. Because they are active against agronomically important insect pests, the P2 proteins are a desirable candidate in the production of insect tolerant transgenic plants either alone or in combination with the other *B.t.*  
15 toxins described in the above examples. In some plants, expression of the P2 protein alone might be sufficient to provide protection against damaging insects. In addition, the P2 proteins might provide protection against agronomically important dipteran  
20 pests. In other cases, expression of P2 together with the *B.t.k.* HD-1 or HD-73 protein might be preferred. The P2 proteins should provide at least an additive level of insecticidal activity when combined with the crystal protein toxin of *B.t.k.* HD-1 or HD-73, and the combination may even provide a synergistic activity.  
25 Although the mode of action of the P2 protein is unknown, its distinct amino acid sequence suggests that it functions differently from the *B.t.k.* HD-1 and HD-73 type of proteins. Production of two insect tolerance proteins with different modes of action in  
30 the same plant would minimize the potential for development of insect resistance to *B.t.* proteins in

-89-

plants. The lack of substantial DNA homology between P2 genes and the HD-1 and HD-73 genes minimizes the potential for recombination between multiple insect tolerance genes in the plant chromosome.

The genes encoding the P2 protein although distinct in sequence from the *B.t.k.* HD-1 and HD-73 genes share many common features with these genes. In particular, the P2 protein genes have a high A+T content (65%), multiple potential polyadenylation signal sequences (26) and numerous ATTTA sequences (10). Because of its overall similarity to the poorly expressed wild-type *B.t.k.* HD-1 and HD-73 genes, the same problems are expected in expression of the wild-type P2 gene as were encountered with the previous examples. Based on the above-described method for designing the synthetic *B.t.* genes, a synthetic P2 gene has been designed which gene should be expressed at adequate levels for protection in plants. A comparison of the wild-type and synthetic P2 genes is shown in Figure 13.

#### Example 7 -- Synthetic *B.t. Entomocidus* Gene

The *B.t. entomocidus* ("Btent") protein is a distinct insecticidal protein produced by some strains of *B.t.* bacteria. It is characterized by its high level of activity against some lepidopterans that are relatively insensitive to *B.t.k.* HD-1 and HD-73 such as *Spodoptera* species including beet armyworm (Visser et al., 1988). Genes encoding the Btent protein have been isolated and characterized (Honee et al, 1988). The Btent proteins encoded by these genes are

-90-

5 approximately the same length as *B.t.k.* HD-1 and HD-73. These proteins share only 68% amino acid homology with the *B.t.k.* HD-1 and HD-73 proteins. It is likely that only the N-terminal half of the Btent protein is required for insecticidal activity as is the case for HD-1 and HD-73. Over the first 625 amino acids, Btent shares only 38% amino acid homology with HD-1 and HD-73.

10 Because of their higher activity against *Spodoptera* species that are relatively insensitive to HD-1 and HD-73, the Btent proteins are a desirable candidate for the production of insect tolerant transgenic plants either alone or in combination with the other *B.t.*  
15 toxins described in the above examples. In some plants production of Btent alone might be sufficient to control the agronomically important pests. In other plants, the production of two distinct insect tolerance proteins would provide protection against a  
20 wider array of insects. Against those insects where both proteins are active, the combination of the *B.t.k.* HD-1 or HD-73 type protein plus the Btent protein should provide at least additive insecticidal efficacy, and may even provide a synergistic activity. In addition, because of its distinct amino acid  
25 sequence, the Btent protein may have a different mode of action than HD-1 or HD-73. Production of two insecticidal proteins in the same plant with different modes of action would minimize the potential for development of insect resistance to *B.t.* proteins in  
30 plants. The relative lack of DNA sequence homology with the *B.t.k.* type genes minimizes the potential for



-91-

recombination between multiple insect tolerance genes

---

5       The genes encoding the Btent protein although  
distinct in sequence from the *B.t.k.* HD-1 and HD-73  
genes share many common features with these genes. In  
particular, the Btent protein genes have a high A+T  
content (62%), multiple potential polyadenylation  
10       signal sequences (39 in the full length coding  
sequence and 27 in the first 1875 nucleotides that is  
likely to encode the active toxic fragment) and  
numerous ATTTA sequences (16 in the full length coding  
sequence and 12 in the first 1875 nucleotides).  
Because of its overall similarity to the poorly  
15       expressed wild type *B.t.k.* HD-1 and HD-73 genes, the  
wild-type Btent genes are expected to exhibit similar  
problems in expression as were encountered with the  
wild-type HD-1 and HD-73 genes. Based on the above-  
described method used for designing the other  
20       synthetic *B.t.* genes, a synthetic Btent gene has been  
designed which gene should be expressed at adequate  
levels for protection in plants. A comparison of the  
wild type and synthetic Btent genes is shown in  
Figure 14.

25       Example 8 -- Synthetic *B.t.k.* Genes for Expression  
in Corn

High level expression of heterologous genes in corn  
cells has been shown to be enhanced by the presence of  
30       a corn gene intron (Callis et al., 1987). Typically  
these introns have been located in the 5' untranslated

-92-

5 region of the chimeric gene. It has been shown that the CaMV35S promoter and the NOS 3' end function efficiently in the expression of heterologous genes in corn cells (Fromm et al., 1986).

Referring to Figure 15, a plant expression cassette vector (pMON744) was constructed that contains these sequences. Specifically the expression cassette contains the enhanced CaMV 35S promoter followed by  
10 intron 1 of the corn Adh1 gene (Callis et al., 1987). This is followed by a multilinker cloning site for insertion of coding sequences; this multilinker contains a BglII site among others. Following the multilinker is the NOS 3' end. pMON744 also contains  
15 the selectable marker gene 35S/NPTII/NOS 3' for kanamycin selection of transgenic corn cells. In addition, pMON744 has an *E. coli* origin of replication and an ampicillin resistance gene for selection of the plasmid in *E. coli*.

20 Five *B.t.k.* coding sequences described in the previous examples were inserted into the BglII site of pMON744 for corn cell expression of *B.t.k.* The coding sequences inserted and resulting vectors were:

25 1. Wild type *B.t.k.* HD-1 from pMON9921 to make pMON8652.

2. Modified *B.t.k.* HD-1 from pMON5370 to make pMON8642.

3. Synthetic *B.t.k.* HD-1 from pMON5377 to make pMON8643.

30 4. Synthetic *B.t.k.* HD-73 from pMON5390 to make pMON8644.

-93-

5. Synthetic full length *B.t.k.* HD-73 from pMON10518 to make pMON10902.

5 pMON8652 (wild-type *B.t.k.* HD-1) was used to transform corn cell protoplasts and stably transformed kanamycin resistant callus was isolated. *B.t.k.* mRNA in the corn cells was analyzed by nuclease S1 protection and found to be present at a level  
10 comparable to that seen with the same wild-type coding sequence (pMON9921) in transgenic tomato plants.

pMON8652 and pMON8642 (modified HD-1) were used to transform corn cell protoplasts in a transient expression system. The level of *B.t.k.* mRNA was  
15 analyzed by nuclease S1 protection. The modified HD-1 gave rise to a several fold increase in *B.t.k.* mRNA compared to the wild-type coding sequence in the transiently transformed corn cells. This indicated that the modifications introduced into the *B.t.k.* HD-1  
20 gene are capable of enhancing *B.t.k.* expression in monocot cells as was demonstrated for dicot plants and cells.

pMON8642 (modified HD-1) and pMON8643 (synthetic HD-1) were used to transform Black Mexican Sweet (BMS) corn cell protoplasts by PEG-mediated DNA uptake, and  
25 stably transformed corn callus was selected by growth on kanamycin containing plant growth medium. Individual callus colonies that were derived from single transformed cells were isolated and propagated separately on kanamycin containing medium.

30 To assess the expression of the *B.t.k.* genes in these cells, callus samples were tested for insect

-94-

toxicity by bioassay against tobacco hornworm larvae. For each vector, 96 callus lines were tested by bioassay. Portions of each callus were placed on sterile water agar plates, and five neonate tobacco hornworm larvae were added and allowed to feed for 4 days. For pMON8643, 100% of the larvae died after feeding on 15 of the 96 calli and these calli showed little feeding damage. For pMON8642, only 1 of the 96 calli was toxic to the larvae. This showed that the B.t.k. gene was being expressed in these samples at insecticidal levels. The observation that significantly more calli containing pMON8643 were toxic than for pMON8642 showed that significantly higher levels of expression were obtained when the synthetic HD-1 coding sequence was contained in corn cells than when the modified HD-1 coding sequence was used, similar to the previous examples with dicot plants. A semiquantitative immunoassay showed that the pMON8643 toxic samples had significantly higher B.t.k. protein levels than the pMON8642 toxic sample.

The 16 callus samples that were toxic to tobacco hornworm were also tested for activity against European corn borer. European corn borer is approximately 40-fold less sensitive to the HD-1 gene product than is tobacco hornworm. Larvae of European corn borer were applied to the callus samples and allowed to feed for 4 days. Two of the 16 calli tested, both of which contained pMON8643 (synthetic HD-1), were toxic to European corn borer larvae.

-95-

5 To assess the expression of the *B.t.k.* genes in differentiated corn tissue, another method of DNA delivery was used. Young leaves were excised from corn plants, and DNA samples were delivered into the leaf tissue by microprojectile bombardment. In this system, the DNA on the microprojectiles is transiently expressed in the leaf cells after bombardment. Three DNA samples were used, and each DNA was tested in triplicate.

1. pMON744, the corn expression vector with no



15 3. pMON752, a corn expression vector for the GUS gene, no *B.t.k.* gene.

20 The leaves were incubated at room temperature for 24 hours. The pMON752 samples were stained with a substrate that allows visual detection of the GUS gene product. This analysis showed that over one hundred spots in each sample were expressing the GUS product and the the triplicate samples showed very similar levels of GUS expression. For the pMON744 and pMON8643 samples 5 larvae of tobacco hornworm were added to each leaf and allowed to feed for 48 hours. All three samples bombarded with pMON744 showed extensive feeding damage and no larval mortality. All three samples bombarded with pMON8643 showed no evidence of feeding damage and 100% larval mortality.

25 30 The samples were also assayed for the presence of *B.t.k.* protein by a qualitative immunoassay. All of

-96-

the pMON8643 samples had detectable B.t.k. protein. These results demonstrated that the the synthetic B.t.k. gene was expressed in differentiated corn plant tissue at insecticidal levels.

Example 9 -- Synthetic Potato Leaf Roll Virus Coat Protein Gene

Expression in plants of the coat protein genes from a variety of plant viruses has proven to be an effective method of engineering resistance to these viruses. In order to achieve virus resistance, it is important to express the viral coat protein at an effective level. For many plant virus coat protein genes, this has not proved to be a problem. However, for the coat protein gene from potato leaf roll virus (PLRV), expression of the coat protein has been observed to be low relative to other coat protein genes, and this lower level of protein has not led to optimal resistance to PLRV.

The gene for PLRV coat protein is shown in Figure 16. Referring to Figure 16, the upper line of sequence shows the gene as it was originally engineered for plant expression in vector pMON893. The gene was contained on a 749 nucleotide BglII-EcoRI fragment with the coding sequence contained between nucleotides 20 and 643. This fragment also contained 19 nucleotides of 5' noncoding sequence and 104 nucleotides of 3' noncoding sequence. This PLRV coat protein gene was relatively poorly expressed in plants compared to other viral coat protein genes.

-97-

5 A synthetic gene was designed to improve plant  
expression of the PLRV coat protein. Referring again  
to Figure 16, the changes made in the synthetic PLRV  
gene are shown in the lower line. This gene was  
designed to encode exactly the same protein as the  
naturally occurring gene. Note that the beginning of  
the synthetic gene is at nucleotide 14 and the end of  
the sequence is at nucleotide 654. The coding  
10 sequence for the synthetic gene is from nucleotide 20  
to 643 of the figure. The changes indicated just  
upstream and downstream of these endpoints serve only  
to introduce convenient restriction sites just outside  
the coding sequence. Thus the size of the synthetic  
15 gene is 641 nucleotides which is smaller than the  
naturally occurring gene. The synthetic gene is  
smaller because substantially all of the noncoding  
sequence at both the 5' and 3' ends, except for  
segments encoding the BglII and EcoRI restriction  
20 sites has been removed.

The synthetic gene differs from the naturally  
occurring gene in two main respects. First, 41  
individual codons within the coding sequence have been  
changed to remove nearly all codons for a given amino  
acid that constitute less than about 15% of the codons  
25 for that amino acid in a survey of dicot plant genes.  
Second, the 5' and 3' noncoding sequences of the  
original gene have been removed. Although not  
strictly conforming to the algorithm described in  
Figure 1, a few of the codon changes and especially  
30 the removal of the long 3' noncoding region is  
consistent with this algorithm.

-98-

The original PLRV sequence contains two potential plant polyadenylation signals (AACCAA and AAGCAT) and both of these occur in the 3' noncoding sequence that has been removed in the synthetic gene. The original PLRV gene also contains an ATTTA sequence. This is also contained in the 3' noncoding sequence, and is in the midst of the longest stretch of uninterrupted A+T in the gene (a stretch of 7 A+T nucleotides). This sequence was removed in the synthetic gene. Thus, sequences that the algorithm of Figure 1 targets for change have been changed in the synthetic PLRV coat protein gene by removal of the 3' noncoding segment. Within the coding sequence, codon changes were also made to remove three other regions of sequence described above. In particular, two regions of 5 consecutive A+T and one region of 5 consecutive G+C within the coding sequence have been removed in the synthetic gene.

The synthetic PLRV coat protein gene is cloned in a plant transformation vector such as pMON893 and used to transform potato plants as described above. These plants express the PLRV coat protein at higher levels than achieved with the naturally occurring gene, and these plants exhibit increased resistance to infection by PLRV.

30



-99-

Example 10 -- Expression of Synthetic B.t. Genes  
with RUBISCO Small Subunit Promoters and  
Chloroplast Transit Peptides

5

The genes in plants encoding the small subunit of RUBISCO (SSU) are often highly expressed, light regulated and sometimes show tissue specificity. These expression properties are largely due to the promoter sequences of these genes. It has been possible to use SSU promoters to express heterologous genes in transformed plants. Typically a plant will contain multiple SSU genes, and the expression levels and tissue specificity of different SSU genes will be different. The SSU proteins are encoded in the nucleus and synthesized in the cytoplasm as precursors that contain an N-terminal extension known as the chloroplast transit peptide (CTP). The CTP directs the precursor to the chloroplast and promotes the uptake of the SSU protein into the chloroplast. In this process, the CTP is cleaved from the SSU protein. These CTP sequences have been used to direct heterologous proteins into chloroplasts of transformed plants.

25

The SSU promoters might have several advantages for expression of B.t.k. genes in plants. Some SSU promoters are very highly expressed and could give rise to expression levels as high or higher than those observed with the CaMV35S promoter. The tissue distribution of expression from SSU promoters is different from that of the CaMV35S promoter, so for control of some insect pests, it may be advantageous

30

-100-

to direct the expression of *B.t.k.* to those cells in which SSU is most highly expressed. For example, although relatively constitutive, in the leaf the  
5 CaMV35S promoter is more highly expressed in vascular tissue than in some other parts of the leaf, while most SSU promoters are most highly expressed in the mesophyll cells of the leaf. Some SSU promoters also are more highly tissue specific, so it could be  
10 possible to utilize a specific SSU promoter to express *B.t.k.* in only a subset of plant tissues, if for example *B.t.* expression in certain cells was found to be deleterious to those cells. For example, for control of Colorado potato beetle in potato, it may be  
15 advantageous to use SSU promoters to direct *B.t.t.* expression to the leaves but not to the edible tubers.

Utilizing SSU CTP sequences to localize *B.t.* proteins to the chloroplast might also be  
20 advantageous. Localization of the *B.t.* to the chloroplast could protect the protein from proteases found in the cytoplasm. This could stabilize the *B.t.* protein and lead to higher levels of accumulation of active protein. *B.t.* genes containing the CTP could  
25 be used in combination with the SSU promoter or with other promoters such as CaMV35S.

A variety of plant transformation vectors were constructed for the expression of *B.t.k.* genes utilizing SSU promoters and SSU CTPs. The promoters and CTPs utilized were from the petunia SSU11a gene  
30 described by Tumer et al. (1986) and from the *Arabidopsis atslA* gene (an SSU gene) described by

-101-

Krebbers et al. (1988) and by Elionor et al. (1989). The petunia SSU11a promoter was contained on a DNA fragment that extended approximately 800 bp upstream of the SSU coding sequence. The *Arabidopsis* atslA promoter was contained on a DNA fragment that extended approximately 1.8 kb upstream of the SSU coding sequence. At the upstream end convenient sites from the multilinker of pUC18 were used to move these promoters into plant transformation vectors such as pMON893. These promoter fragments extended to the start of the SSU coding sequence at which point an NcoI restriction site was engineered to allow insertion of the *B.t.* coding sequence, replacing the SSU coding sequence.

When SSU promoters were used in combination with their CTP, the DNA fragments extended through the coding sequence of the CTP and a small portion of the mature SSU coding sequence at which point an NcoI restriction site was engineered by standard techniques to allow the in frame fusion of *B.t.* coding sequences with the CTP. In particular, for the petunia SSU11a CTP, *B.t.* coding sequences were fused to the SSU sequence after amino acid 8 of the mature SSU sequence at which point the NcoI site was placed. The 8 amino acids of mature SSU sequence were included because preliminary in vitro chloroplast uptake experiments indicated that uptake was of *B.t.k.* was observed only if this segment of mature SSU was included. For the *Arabidopsis* atslA CTP, the complete CTP was included plus 24 amino acids of mature SSU sequence plus the sequence gly-gly-arg-val-asn-cys-met-gln-ala-met,

-102-

terminating in an NcoI site for *B.t.* fusion. This short sequence reiterates the native SSU CTP cleavage site (between the cys and met) plus a short segment surrounding the cleavage site. This sequence was included in order to insure proper uptake into chloroplasts. *B.t.* coding sequences were fused to this *ats1A* CTP after the met codon. In vitro uptake experiments with this CTP construction and other (non-*B.t.*) coding sequences showed that this CTP did target proteins to the chloroplast.

When CTPs were used in combination with the CaMV 35S promoter, the same CTP segments were used. They were excised just upstream of the ATG start sites of the CTP by engineering of BglII sites, and placed downstream of the CaMV35S promoter in pMON893, as BglII to NcoI fragments. *B.t.* coding sequences were fused as described above.

The wild type *B.t.k.* HD-1 coding sequence of pMON9921 (see Figure 1) was fused to the *ats1A* promoter to make pMON1925 or the *ats1A* promoter plus CTP to make pMON1921. These vectors were used to transform tobacco plants, and the plants were screened for activity against tobacco hornworm. No toxic plants were recovered. This is surprising in light of the fact that toxic plants could be recovered, albeit at a low frequency, after transformation with pMON9921 in which the *B.t.k.* coding sequence was expressed from the enhanced CaMV35S, promoter in pMON893, and in light of the fact that Elionor et al. (1989) report that the *ats1A* promoter itself is comparable in strength to the CaMV35S promoter and approximately 10-

-103-

fold stronger when the CTP sequence is included. At  
least for the wild-type *B.t.k.* HD-1 coding sequence,  
this does not appear to be the case.

5

A variety of plant transformation vectors were  
constructed utilizing either the truncated synthetic .  
HD-73 coding sequence of Figure 4 or the full length  
*B.t.k.* HD-73 coding sequence of Figure 11. These are  
listed in the table below.

10

15

20

25

30

-104-

Table XV

5                                    Gene Constructs with CTPs

<u>Vector</u>	<u>Promoter</u>	<u>CTP</u>	<u>B.t.k. HD-73</u> <u>Coding Sequence</u>
pMON10806	En 35S	ats1A	truncated
pMON10814	En35S	SSU11a	full length
10 pMON10811	SSU11a	SSU11a	truncated
pMON10819	SSU11a	none	truncated
pMON10815	ats1A	none	truncated
pMON10817	ats1A	ats1A	truncated
pMON10821	En 35S	ats1A	truncated
pMON10822	En 35S	ats1A	full length
15 pMON10838	SSU11a	SSU11a	full length
pMON10839	ats1A	ats1A	full length

All of the above vectors were used to transform tobacco plants. For all of the vectors containing truncated *B.t.k.* genes, leaf tissue from these plants has been analyzed for toxicity to insects and *B.t.k.* protein levels by immunoassay. pMON10806, 10811, 10819 and 10821 produce levels of *B.t.k.* protein comparable to pMON5383 and pMON5390 which contain synthetic *B.t.k.* HD-73 coding sequences driven by the En 35S promoter itself with no CTP. These plants also have the insecticidal activity expected for the *B.t.k.* protein levels detected. For pMON10815 and pMON10817 (containing the *ats1A* promoter), the level of *B.t.k.* protein is about 5-fold higher than that found in plants containing pMON5383 or 5390. These plants also

-105-

5 have higher insecticidal activity. Plants containing 10815 and 10817 contain up to 1% of their total soluble leaf protein as *B.t.k.* HD-73. This is the highest level of *B.t.k.* protein yet obtained with any of the synthetic genes.

10 This result is surprising in two respects. First, as noted above, the wild type coding sequences fused to the *ats1A* promoter and CTP did not show any evidence of higher levels of expression than for En 35S, and in fact had lower expression based on the absence of any insecticidal plants. Second, Elionor et al. (1989) show that for two other genes, the *ats1A* CTP can increase expression from the *ats1A* promoter by 15 about 10-fold. For the synthetic *B.t.k.* HD-73 gene, there is no consistent increase seen by including the CTP over and above that seen for the *ats1A* promoter alone.

20 Tobacco plants containing the full length synthetic HD-73 fused to the SSU11A CTP and driven by the En 35S promoter produced levels of *B.t.k.* protein and insecticidal activity comparable to pMON1518 which contains does not include the CTP. In addition, for pMON10518 the *B.t.k.* protein extracted from plants was 25 observed by gel electrophoresis to contain multiple forms less than full length, apparently due the cleavage of the C-terminal portion (not required for toxicity) in the cytoplasm. For pMON10814, the majority of the protein appeared to be intact full length indicating that the protein has been stabilized 30 from proteolysis by targeting to the chloroplast.

-106-

Example 11 -- Targeting of B.t. Proteins to the Extracellular Space or Vacuole through the Use of Signal Peptides

5

The B.t. proteins produced from the synthetic genes described here are localized to the cytoplasm of the plant cell, and this cytoplasmic localization results in plants that are insecticidally effective. It may be advantageous for some purposes to direct the B.t. proteins to other compartments of the plant cell. Localizing B.t. proteins in compartments other than the cytoplasm may result in less exposure of the B.t. proteins to cytoplasmic proteases leading to greater accumulation of the protein yielding enhanced insecticidal activity. Extracellular localization could lead to more efficient exposure of certain insects to the B.t. proteins leading to greater efficacy. If a B.t. protein were found to be deleterious to plant cell function, then localization to a noncytoplasmic compartment could protect these cells from the . protein.

In plants as well as other eucaryotes, proteins that are destined to be localized either extracellularly or in several specific compartments are typically synthesized with an N-terminal amino acid extension known as the signal peptide. This signal peptide directs the protein to enter the compartmentalization pathway, and it is typically cleaved from the mature protein as an early step in compartmentalization. For an extracellular protein, the secretory pathway typically involves



-107-

cotranslational insertion into the endoplasmic reticulum with cleavage of the signal peptide occurring at this stage. The mature protein then passes thru the Golgi body into vesicles that fuse with the plasma membrane thus releasing the protein into the extracellular space. Proteins destined for other compartments follow a similar pathway. For example, proteins that are destined for the endoplasmic reticulum or the Golgi body follow this scheme, but they are specifically retained in the appropriate compartment. In plants, some proteins are also targeted to the vacuole, another membrane bound compartment in the cytoplasm of many plant cells. Vacuole targeted proteins diverge from the above pathway at the Golgi body where they enter vesicles that fuse with the vacuole.

A common feature of this protein targeting is the signal peptide that initiates the compartmentalization process. Fusing a signal peptide to a protein will in many cases lead to the targeting of that protein to the endoplasmic reticulum. The efficiency of this step may depend on the sequence of the mature protein itself as well. The signals that direct a protein to a specific compartment rather than to the extracellular space are not as clearly defined. It appears that many of the signals that direct the protein to specific compartments are contained within the amino acid sequence of the mature protein. This has been shown for some vacuole targeted proteins, but it is not yet possible to define these sequences precisely. It appears that secretion into the

-108-

extracellular space is the "default" pathway for a protein that contains a signal sequence but no other compartmentalization signals. Thus, a strategy to  
5 direct *B.t.* proteins out of the cytoplasm is to fuse the genes for synthetic *B.t.* genes to DNA sequences encoding known plant signal peptides. These fusion genes will give rise to *B.t.* proteins that enter the secretory pathway, and lead to extracellular  
10 secretion or targeting to the vacuole or other compartments.

Signal sequences for several plant genes have been described. One such sequence is for the tobacco pathogenesis related protein PR1b described by  
15 Cornelissen et al. The PR1b protein is normally localized to the extracellular space. Another type of signal peptide is contained on seed storage proteins of legumes. These proteins are localized to the protein body of seeds, which is a vacuole like  
20 compartment found in seeds. A signal peptide DNA sequence for the beta subunit of the 7S storage protein of common bean (*Phaseolus vulgaris*), PvuB has been described by Doyle et al. Based on the published these published sequences, genes were synthesized by  
25 chemical synthesis of oligonucleotides that encoded the signal peptides for PR1b and PvuB. The synthetic genes for these signal peptides corresponded exactly to the reported DNA sequences. Just upstream of the translational initiation codon of each signal peptide a  
30 BamHI and BglII site were inserted with the BamHI site at the 5' end. This allowed the insertion of the signal peptide encoding segments into the BglII site

-109-

of pMON893 for expression from the En 35S promoter. In some cases to achieve secretion or compartmentalization of heterologous proteins, it has  
5 proved necessary to include some amino acid sequence beyond the normal cleavage site of the signal peptide. This may be necessary to insure proper cleavage of the signal peptide. For PR1b the synthetic DNA sequence also included the first 10 amino acids of mature PR1b.  
10 For PvuB the synthetic DNA sequence included the first 13 amino acids of mature PvuB. Both synthetic signal peptide encoding segments ended with NcoI sites to allow fusion in frame to the methionine initiation codon of the synthetic *B.t.* genes.

15 Four vectors encoding synthetic *B.t.k.* HD-73 genes were constructed containing these signal peptides. The synthetic truncated HD-73 gene from pMON5383 was fused with the signal peptide sequence of PvuB and incorporated into pMON893 to create pMON10827. The  
20 synthetic truncated HD-73 gene from pMON5383 was also fused with the signal peptide sequence of PR1b to create pMON10824. The full length synthetic HD-73 gene from pMON10518 was fused with the signal peptide sequence of PvuB and incorporated into pMON893 to create pMON10828. The full length synthetic HD-73  
25 gene from pMON10518 was also fused with the signal peptide sequence of PR1b and incorporated into pMON893 to create pMON10825.

These vectors were used to transform tobacco plants and the plants were assayed for expression of the  
30 *B.t.k.* protein by Western blot analysis and for insecticidal efficacy. pMON10824 and pMON10827

-110-

produced amounts of *B.t.k.* protein in leaf comparable  
to the truncated HD-73 vectors, pMON5383 and pMON5390.  
5 pMON10825 and pMON10828 produced full length *B.t.k.*  
protein in amounts comparable to pMON10518. In all  
cases, the plants were insecticidally active against  
tobacco hornworm.

10

15

20

25

30

-111-

BIBLIOGRAPHY

- 5 Adami, G. and Nevins, J. (1988) RNA Processing, Cold Spring Harbor Laboratory, p. 26.
- Adang, et al., Molecular Strategies for Crop Protection (1987) pp. 345-353, Alan R. Liss, Inc.
- Barton, K. A. et al., Plant Physiol. (1987), 85,1103-1109.
- 10 Bevan, M. et al., Nature (1983) 304:184.
- Brady, H. and Wold, W. (1988), RNA Processing, Cold Spring Harbor Laboratory, p. 224.
- Brown, John W., Nucleic Acids Research (1986) Vol. 14, No. 24, p. 9549.
- 15 Callis, J. Fromm, M. and Walbot, V., Genes and Develop. (1987), 1:1183-1200.
- Conway, L. and Wickens, M. (1988), RNA Processing, Cold Spring Harbor Laboratory, p. 40.
- Cornellssen, B.J.C., et al., EMBO J. (1986) Vol. 5, No. 1, 37-40.
- 20 Daar, I. O. et al. (1988), RNA Processing, Cold Spring Harbor Laboratory, p. 45.
- Dean, C. et al., Nucleic Acids Research (1986), Vol. 14, No. 5, p. 2229.
- 25 Dedrick, R., et al., The Journal of Biological Chemistry (1987), Vol. 262, No. 19, pp. 9098-1106.
- Donovan, W. P. et al., The J. of Biol. Chem. (1988), Vol. 263, No. 1, pp. 561-567.
- Doyle, J.J., et al., J. Biol. Chem. (1986), Vol. 261, No. 20, 9228-9236.
- 30 Elionor, R.P., et al., Mol. Gen. Genet. (1989), 218:78-86.

-112-

- 5 Fischhoff, D. A. et al., Bio/Technology (1987), Vol. 5, p. 807.
- Fraley, R. T. et al., Bio/Technology (1985) 3:629-635.
- Fromm, M., Taylor, L. P. and Walbot, V., Nature (1986), 319:791-793.
- 10 Gallego, M. E. and Nadal-Ginard B. (1988), RNA Processing, Cold Spring Harbor Laboratory, p. 61.
- Genovese, C. and Milcarek, C. (1988), RNA Processing, Cold Spring Harbor Laboratory, p. 62.
- Gil, A. and Proudfoot, N. J., Nature (1984), Vol. 312, p. 473.
- 15 Goodall, G. et al. (1988), RNA Processing, Cold Spring Harbor Laboratory, p. 63.
- Gross, et al. (1988), RNA Processing, Cold Spring Harbor Laboratory, p. 128.
- Hampson, R. K. and Rottman, F. M. (1988), RNA Processing, Cold Spring Harbor Laboratory, p. 68.
- 20 Hanley, Brian A and Schuler, Mary A., Nucleic Acids Research (1988), Vol. 16, No. 14, p. 7159.
- Helfman, D. M. and Ricci, W. M. (1988), RNA Processing, Cold Spring Harbor Laboratory, p. 219.
- Herrera-Estrella, L. et al., Nature (1983), 303:209.
- 25 Hoekema, A. et al., Molecular and Cellular Biology (1987), Vol. 7, pp. 2914-2924.
- Honee, G. et al., Nucleic Acids Research (1988), Vol. 16, No. 13.
- Horsch, R. B. et al., Science (1985), 227:1229.
- 30 Jarret, R. L. et al., Physiol. Plant (1980), 49:177.

-113-

- Jarret, R. L. et al., In Vitro (1981), 17:825.
- 5 Kay, R. et al., Science (1987), 236:1299-1302.
- Kessler, M. et al. (1988), RNA Processing, Cold Spring Harbor Laboratory, p. 85.
- Klee, H. J. et al., Bio/Technology (1985), 3:637-642.
- Kozak, M., Nature (1984), 308:241-246.
- 10 Krebbers, E., et al., Plant Molecular Biology (1988), 11:745-759.
- Kunkel, T. A., Proc. Natl. Acad. Sci. USA (1985), Vol. 82, pp. 488-492.
- Marrone et al., J. Econ. Entomol. (1985), 78-290-293.
- 15 Marzluff, W. and Pandey, N. (1988), RNA Processing, Cold Spring Harbor Laboratory, p. 244.
- McCormick, S. et al., Plant Cell Reports (1986), 5:81-84.
- McDevitt, M. A. et al., Cell (1984), Vol. 37, pp. 993-999.
- 20 Murashige, T. and Skoog, F., Physiol. Plant (1962), 15:473.
- Odell, J. et al., Nature (1985), 313:810.
- Pandey, N. B. and Marzluff, W. F. (1987), RNA Processing, Cold Spring Harbor Laboratory, p. 133.
- 25 Proudfoot, N. J. et al. (1987), RNA Processing, Cold Spring Harbor Laboratory, p. 17.
- Reines, D., et al., J. Mol. Biol. (1987) 196:299-312.
- Sadofsky, M. and Alwine, J. C., Molecular and Cellular Biology (1984), Vol. 4, No. 8, pp. 1460-1468.
- 30

-114-

- Sanders, P. R. et al., Nucleic Acids Research (1987), Vol. 15, No. 4, p. 1543.
- 5 Schuler, M. A. et al., Nucleic Acids Research (1982), Vol. 10, No. 24, pp. 8225-8244.
- Shaw, G. & Kamen, R., Cell (1986), 46:659-667.
- Shaw, G. and Kamen, R. (1987), RNA Processing, Cold Spring Harbor Laboratory, p. 220.
- 10 Trolinder, N. L. and Goodin, J. R., Plant Cell Reports (1987), 6:231-234.
- Tsurushita, N. and Korn, L. J. (1987), RNA Processing, Cold Spring Harbor Laboratory, p. 215.
- Tumer, N.E., et al., Nucleic Acids Res. (1986), Vol. 14:8, 3325.
- 15 Vaeck, M. et al., Nature (1987), Vol. 328, p. 33.
- Velten et al., EMBO J. (1984), 3:2723-2730.
- Velten & Schell, Nucleic Acids Research (1985), 13:6981-6998.
- 20 Visser, B. et al., Mol. Gen. Genet. (1988), 212:219-224.
- Webb, K. J. et al., Plant Sci. Letters (1983), 30:1.
- Wickens, M. and Stephenson, P., Science (1984), Vol. 226, p. 1045.
- 25 Wickens, M. et al. (1987), RNA Processing, Cold Spring Harbor Laboratory, p. 9.
- Wiebauer, K. et al., Molecular and Cellular Biology (1988), Vol. 8, No. 5, pp. 2042-2051.
- 30 Yamamoto, T. and Iizuka, T., Archives of Biochemistry and Biophysics (1983), Vol. 227, No. 1, pp. 233-241.



-115-

## Claims:

- 5        1. In a method for improving the expression of a heterologous gene in plants by modifying the structural coding sequence of said gene, the improvement which comprises reducing the occurrence of polyadenylation signals selected from the group consisting of AATAAA, AATAAT, AACCAA, ATATAA, AATCAA, 10 ATACTA, ATAAAA, ATGAAA, AAGCAT, ATTAAT, ATACAT, AAAATA, ATTAAA, AATTAA, AATACA and CATAAA.
2. The method of Claim 1 further comprising the improvement of reducing the occurrence of ATTTA sequences within the structural coding sequence.
- 15       3. A method for modifying a wild-type structural gene sequence which encodes an insecticidal protein of *Bacillus thuringiensis* to enhance the expression of said protein in plants which comprises:
- 20       a) removing polyadenylation signals contained in said wild-type gene while retaining a sequence which encodes said protein; and
- b) removing ATTTA sequences contained in said wild-type gene while retaining a sequence which encodes said protein.
- 25       4. A method of Claim 3 further comprising the removal of self-complementary sequences and replacement of such sequences with nonself-complementary DNA comprising plant preferred codons while retaining a structural gene sequence encoding said protein.
- 30       5. A method of Claim 4 further comprising the use of plant preferred sequences in the removal of the polyadenylation signals and ATTTA sequences.

-116-

5 6. A method of Claim 3 in which the polyadenylation signals are selected from the group consisting of AATAAA, AATAAT, AACCAA, ATATAA, AATCAA, ATACTA, ATAAAA, ATGAAA, AAGCAT, ATTAAT, ATACAT, AAAATA, ATTAAA, AATTAA, AATACA and CATAAA.

10 7. A method of Claim 4 in which the polyadenylation signals are selected from the group consisting of AATAAA, AATAAT, AACCAA, ATATAA, AATCAA, ATACTA, ATAAAA, ATGAAA, AAGCAT, ATTAAT, ATACAT, AAAATA, ATTAAA, AATTAA, AATACA and CATAAA.

15 8. A method of Claim 5 in which the polyadenylation signals are selected from the group consisting of AATAAA, AATAAT, AACCAA, ATATAA, AATCAA, ATACTA, ATAAAA, ATGAAA, AAGCAT, ATTAAT, ATACAT, AAAATA, ATTAAA, AATTAA, AATACA and CATAAA.

20 9. A method for modifying a wild-type structural gene sequence which encodes an insecticidal protein of *Bacillus thuringiensis* to enhance the expression of said protein in plants which comprises:

- a) identifying regions within said sequence with greater than four consecutive adenine or thymine nucleotides;
- 25 b) modifying the regions of step (a) which have two or more polyadenylation signals within a ten base sequence to remove said signals while maintaining a gene sequence which encodes said protein; and
- 30 c) modifying the 15-30 base regions surrounding the regions of step (a) to remove major plant polyadenylation signals, consecutive sequences containing more than one minor polyadenylation

-117-

signal and consecutive sequences containing more than one ATTTA sequence while maintaining a gene sequence which encodes said protein.

5

10. A method of Claim 9 in which the major plant polyadenylation signals are selected from the group consisting of AATAAA and AATAAT.

10

11. A method of Claim 10 in which the polyadenylation signals are selected from the group consisting of AATAAA, AATAAT, AACCAA, ATATAA, AATCAA, ATACTA, ATAAAA, ATGAAA, AAGCAT, ATTAAT, ATACAT, AAAATA, ATTAAA, AATTAA, AATACA and CATAAA.

15

12. A method of Claim 11 further comprising the use of plant preferred sequences in the removal of polyadenylation signals and ATTTA sequences.

20

13. A structural gene which encodes an insecticidal protein of *Bacillus thuringiensis*, said gene being substantially devoid of polyadenylation signals and ATTTA sequences.

25

14. A structural gene of Claim 13 which is substantially devoid of polyadenylation signals selected from the group consisting of AATAAA, AATAAT, AACCAA, ATATAA, AATCAA, ATACTA, ATAAAA, ATGAAA, AAGCAT, ATTAAT, ATACAT, AAAATA, ATTAAA, AATTAA, AATACA and CATAAA.

30

-118-

15. A structural gene of Claim 13 which encodes an insecticidal protein of *B.t.k.* HD-1 having the sequence:

5

1	ATGGCTATAGAACTGGTTACACCCCAATCGATATTCCT	40
41	TGTCGCTAACGCAATTTCTTTTGAGTGAATTTGTTCCCGG	80
10	81 TGCTGGATTTGTGTTAGGACTAGTTGATATTATCTGGGGA	120
121	ATTTTGGTCCCTCTCAATGGGACGCATTCTTGTACAAA	160
161	TTGAACAGCTCATCAACCAGAGAATCGAAGAGTTCGCTAG	200
15	201 GAATCAAGCCATTTCTAGATTAGAAGGACTAAGCAATCTT	240
241	TATCAAATTTACGCAGAATCTTTAGAGAGTGGGAAGCAG	280
281	ATCCTACTAATCCAGCATTAAAGAGAAGAGATGCGTATTCA	320
321	ATTCAATGACATGAACAGTGCCCTTACAACCGCTATTCCT	360
361	CTTTTTGCAGTTCAAATTATCAAGTTCCTCTCCTCTCCG	400
25	401 TGTACGTTCAAGCTGCCAACCTCCACCTCTCAGTTTTGAG	440
441	AGATGTTTCAGTGTTTGGACAAAGGTGGGGATTGATGCC	480
481	GCGACTATCAATAGTCGTTATAATGATTAACTAGGCTTA	520

30

-119-

521 TTGGCAACTATACAGATCATGCTGTACGCTGGTACAATAC 560  
5 561 GGGATTAGAGCGTGTATGGGGACCGGATTCTAGAGATTGG 600  
601 ATCAGGTACAACCAGTTCAGAAGAGAGCTTACACTAACTG 640  
641 TATTAGATATCGTTTCTCTATTTCCGAACCTATGATAGTAG 680  
10 681 AACGTATCCAATTCGAACAGTTTCCCAATTAACAAGAGAA 720  
721 ATTTATACAAACCCAGTATTAGAAAATTTTGATGGTAGTT 760  
15 761 TTCGAGGCTCGGCTCAGGGCATAGAAGGAAGTATTAGGAG 800  
801 TCCACATTTGATGGATATACTTAATAGTATAACCATCTAT 840  
841 ACGGATGCTCATAGAGGAGAATACTACTGGTCCGGTCACC 880  
20 881 AGATCATGGCTTCTCCTGTAGGGTTTTTCGGGGCCAGAATT 920  
921 CACTTTTCCGCTATATGGAACCTATGGGAAATGCAGCTCCA 960  
961 CAACAACGTATTGTTGCTCAACTAGGTCAGGGCGTGTATA 1000  
25 1001 GAACATTATCGTCCACCTTATATAGAAGACCTTTTAACAT 1040  
1041 CGGGATCAACAACCAACAACCTATCTGTTCTTGACGGGACA 1080  
30 1081 GAATTTGCTTATGGAACCTCCTCAAATTTGCCATCCGCTG 1120

-120-

	1121	TATACAGAAAAAGCGGAACGGTAGATTTCGCTGGATGAAAT	1160
5	1161	ACCGCCACAGAATAACAACGTGCCACCTAGGCAAGGATTT	1200
	1201	AGTCATCGATTAAGCCATGTTTCAATGTTTCGTTTCAGGCT	1240
10	1241	TTAGTAATAGTAGTGTAAGTATAATAAGAGCTCCTATGTT	1280
	1281	CTCTTGGATACATCGTAGTGCTGAGTTCAACAACATCATC	1320
	1321	CCTTCATCACAAATCACCCAAATCCCACTCACCAAGTCTA	1360
15	1361	CTAATCTTGGCTCTGGAACCTTCTGTCGTTAAAGGACCAGG	1400
	1401	ATTTACAGGAGGAGATATTCTTCGAAGAACTTCACCTGGC	1440
	1441	CAGATTTCAACCTTAAGAGTAAATATTACTGCACCATTAT	1480
20	1481	CACAAAGATATCGGGTAAGAATTCGCTACGCTTCTACCAC	1520
	1521	AAACCTTCAGTTCCACACATCAATTGACGGAAGACCTATT	1560
25	1561	AATCAGGGGAATTTTTCAGCAACTATGAGTAGTGGGAGTA	1600
	1601	ATTTACAGTCCGGAAGCTTTAGGACTGTAGGTTTTACTAC	1640
	1641	TCCGTTTAACTTTTCAAATGGATCAAGTGTATTTACGTTA	1680
30			

-121-

1681 AGTGCTCATGTCTTCAATTCAGGCAATGAAGTTTATATAG 1720  
5 1721 ATCGAATTGAATTTGTTCCGGCA 1743.

16. A structural gene of Claim 13 which encodes an insecticidal protein of *B.t.k.* HD-73 having the sequence:

10  
1 ATGGCCATTGAAACCGGTTACACTCCCATCGACATCTCCT 40  
41 TGTCTTGACACAGTTTCTGCTCAGCGAGTTCGTGCCAGG 80  
15 81 TGCTGGGTTTCGTTCTCGGACTAGTTGACATCATCTGGGGT 120  
121 ATCTTTGGTCCATCTCAATGGGATGCATTCCTGGTGCAA 160  
161 TTGAGCAGTTGATCAACCAGAGGATCGAAGAGTTCGCCAG 200  
20 201 GAACCAGGCCATCTCTAGGTTGGAAGGATTGAGCAATCTC 240  
241 TACCAAATCTATGCAGAGAGCTTCAGAGAGTGGGAAGCCG 280  
25 281 ATCCTACTAACCCAGCTCTCCGCGAGGAAATGCGTATTCA 320  
321 ATTCAACGACATGAACAGCGCCTTGACCACAGCTATCCCA 360  
361 TTGTTGCGCAGTCCAGAACTACCAAGTTCCTCTCTTGTCG 400  
30 401 TGTACGTTCAAGCAGCTAATCTTCACCTCAGCGTGCTTCG 440

-122-

	441	AGACGTTAGCGTGTTTGGGCAAAGGTGGGGATTTCGATGCT	480
5	481	GCAACCATCAATAGCCGTTACAACGACCTTACTAGGCTGA	520
	521	TTGGAAACTACACCGACCACGCTGTTTCGTTGGTACAACAC	560
	561	TGGCTTGGAGCGTGCTCTGGGGTCCTGATTCTAGAGATTGG	600
10	601	ATTAGATACAACCAGTTCAGGAGAGAATTGACCCTCACAG	640
	641	TTTTGGACATTGTGTCTCTCTTCCCGAACTATGACTCCAG	680
15	681	AACCTACCCTATCCGTACAGTGTCCTCAACTTACCAGAGAA	720
	721	ATCTATACTAACCCAGTTCTTGAGAACTTCGACGGTAGCT	760
	761	TCCGTGGTTCTGCCCCAAGGTATCGAAGGCTCCATCAGGAG	800
20	801	CCCACACTTGATGGACATCTTGAACAGCATAACTATCTAC	840
	841	ACCGATGCTCACAGAGGAGAGTATTACTGGTCTGGACACC	880
25	881	AGATCATGGCCTCTCCAGTTGGATTTCAGCGGGCCCGAGTT	920
	921	TACCTTTCCTCTCTATGGAACTATGGGAAACGCCGCTCCA	960
	961	CAACAACGTATCGTTGCTCAACTAGGTCAGGGTGTCTACA	1000
30	1001	GAACCTTGTCTTCCACCTTGTACAGAAGACCCTTCAATAT	1040



-123-

	1041	CGGTATCAACAACCAGCAACTTTCCGTTCTTGACGGAACA	1080
5	1081	GAGTTCGCCTATGGAACCTCTTCTAACTTGCCATCCGCTG	1120
	1121	TTTACAGAAAGAGCGGAACCGTTGATTCCTTGGACGAAAT	1160
	1161	CCCACCACAGAACAACAATGTGCCACCCAGGCAAGGATTC	1200
10	1201	TCCCACAGGTTGAGCCACGTGTCCATGTTCCGTTCCGGAT	1240
	1241	TCAGCAACAGTTCGCTGAGCATCATCAGAGCTCCTATGTT	1280
15	1281	CTCTTGGATACACCGTAGTGCTGAGTTCAACAACATCATC	1320
	1321	GCATCCGATAGTATTACTCAAATCCCTGCAGTGAAGGGAA	1360
	1361	ACTTTCTCTTCAACGGTTCTGTCAATTCAGGACCAGGATT	1400
20	1401	CACTGGTGGAGACCTCGTTAGACTCAACAGCAGTGGAAAT	1440
	1441	AACATTCAGAATAGAGGGTATATTGAAGTTCCAATTCACT	1480
	1481	TCCCATCCACATCTACCAGATATAGAGTTCGTGTGAGGTA	1520
25	1521	TGCTTCTGTGACCCCTATTCACCTCAACGTTAATTGGGGT	1560
	1561	AATTCATCCATCTTCTCCAATACAGTTCCAGCTACAGCTA	1600
30	1601	CCTCCTTGGATAATCTCCAATCCAGCGATTTCCGGTTACTT	1640

-124-

1641 TGAAAGTGCCAATGCTTTTACATCTTCACTCGGTAACATC 1680  
5 1681 GTGGGTGTTAGAACTTTAGTGGGACTGCAGGAGTGATTA 1720  
1721 TCGACAGATTCGAGTTCATTCCAGTTACTGCAACACTCGA 1760  
1761 GGCTGAG 1767.  
10

17. A structural gene of Claim 13 encoding a insecticidal protein of *B.t.k.* HD-1 having the sequence:

15 1 ATGGACAACAACCCAAACATCAACGAATGCATTCCATACA 40  
41 ACTGCTTGAGTAACCCAGAAGTTGAAGTACTTGGTGGAGA 80  
81 ACGCATTGAAACCGGTTACACTCCCATCGACATCTCCTTG 120  
20 121 TCCTTGACACAGTTTCTGCTCAGCGAGTTCGTGCCAGGTG 160  
161 CTGGGTTCGTTCTCGGACTAGTTGACATCATCTGGGGTAT 200  
25 201 CTTTGGTCCATCTCAATGGGATGCATTCTGGTGCAAATT 240  
241 GAGCAGTTGATCAACCAGAGGATCGAAGAGTTCGCCAGGA 280  
281 ACCAGGCCATCTCTAGGTTGGAAGGATTGAGCAATCTCTA 320  
30 321 CCAAATCTATGCAGAGAGCTTCAGAGAGTGGGAAGCCGAT 360

-125-

361 CCTACTAACCCAGCTCTCCGCGAGGAAATGCGTATTCAAT 400

5 401 TCAACGACATGAACAGCGCCTTGACCACAGCTATCCCATT 440

441 GTTCGCAGTCCAGAACTACCAAGTTCCTCTCTTGTCGGTG 480

481 TACGTTCAAGCAGCTAATCTTCACCTCAGCGTGCTTCGAG 520

10 521 ACGTTAGCGTGTTTGGGCAAAGGTGGGGATTTCGATGCTGC 560

561 AACCATCAATAGCCGTTACAACGACCTTACTAGGCTGATT 600

15 601 GGAAACTACACCGACCACGCTGTTTCGTTGGTACAACACTG 640

641 GCTTGGAGCGTGTCTGGGGTCTGATTCTAGAGATTGGAT 680

681 TAGATAACAACAGTTCAGGAGAGAATTGACCCTCACAGTT 720

20 721 TTGGACATTGTGTCTCTCTTCCCGAACTATGACTCCAGAA 760

761 CCTACCCTATCCGTACAGTGTCCCAACTTACCAGAGAAAT 800

801 CTATACTAACCCAGTTCTTGAGAACTTCGACGGTAGCTTC 840

25 841 CGTGGTTCTGCCCCAAGGTATCGAAGGCTCCATCAGGAGCC 880

881 CACACTTGATGGACATCTTGAACAGCATAACTATCTACAC 920

30 921 CGATGCTCACAGAGGAGAGTATTACTGGTCTGGACACCAG 960

-126-

	961	ATCATGGCCTCTCCAGTTGGATTTCAGCGGGCCCGAGTTTA	1000
5	1001	CCTTTCCTCTCTATGGAACCTATGGGAAACGCCGCTCCACA	1040
	1041	ACAACGTATCGTTGCTCAACTAGGTCAGGGTGTCTACAGA	1080
10	1081	ACCTTGTCTTCCACCTTGTACAGAAGACCCTTCAATATCG	1120
	1121	GTATCAACAACCAGCAACTTTCGGTTCTTGACGGAACAGA	1160
	1161	GTCGCCTATGGAACCTCTTCTAACTTGCCATCCGCTGTT	1200
15	1201	TACAGAAAGAGCGGAACCGTTGATTCCCTGGACGAAATCC	1240
	1241	CACCACAGAACAACAATGTGCCACCCAGGCAAGGATTCTC	1280
	1281	CCACAGGTTGAGCCACGTGTCCATGTTCCGTTCCGGATTC	1320
20	1321	AGCAACAGTTCCGTGAGCATCATCAGAGCTCCTATGTTCT	1360
	1361	CATGGATTCATCGTAGTGCTGAGTTCAACAATATCATTCC	1400
25	1401	TTCCTCTCAAATCACCCAAATCCCATTGACCAAGTCTACT	1440
	1441	AACCTTGGATCTGGAACCTTCTGTCGTGAAAGGACCAGGCT	1480
	1481	TCACAGGAGGTGATATTCTTAGAAGAACTTCTCCTGGCCA	1520
30	1521	GATTAGCACCCCTCAGAGTTAACATCACTGCACCACTTTCT	1560

-127-

```

      .       .       .       .
1561 CAAAGATATCGTGT CAGGATTCGTTACGCATCTACCACTA 1600
      .       .       .       .
5  1601 ACTTGCAATTCCACACCTCCATCGACGGAAGGCCTATCAA 1640
      .       .       .       .
1641 TCAGGGTAACTTCTCCGCAACCATGTCAAGCGGCAGCAAC 1680
      .       .       .       .
1681 TTGCAATCCGGCAGCTTCAGAACCGTCGGTTTCACTACTC 1720
10      .       .       .       .
1721 CTTTCAACTTCTCTAACGGATCAAGCGTTTTTACCCTTAG 1760
      .       .       .       .
1761 CGCTCATGTGTTCAATTCTGGCAATGAAGTGACATTGAC 1800
      .       .       .       .
15 1801 CGTATTGAGTTTGTGCCTGCCGAAGTTACCTTCGAGGCTG 1840
      .       .       .       .
1841 AGTAC 1845.

```

18. A structural gene of Claim 13 encoding an insecticidal protein derived from *B.t.k.* HD-73 having the sequence:

```

      .       .       .       .
      1 ATGGACAACAACCCAAACATCAACGAATGCATTCCATACA 40
      .       .       .       .
25 41 ACTGCTTGAGTAACCCAGAAGTTGAAGTACTTGGTGGAGA 80
      .       .       .       .
      81 ACGCATTGAAACCGGTTACTCCCATCGACATCTCCTTG 120
      .       .       .       .
121 TCCTTGACACAGTTTCTGCTCAGCGAGTTCGTGCCAGGTG 160
      .       .       .       .
30 161 CTGGGTTTCGTTCTCGGACTAGTTGACATCATCTGGGGTAT 200

```

-128-

	201	CTTTGGTCCATCTCAATGGGATGCATTCCTGGTGCAAATT	240
5	241	GAGCAGTTGATCAACCAGAGGATCGAAGAGTTCGCCAGGA	280
	281	ACCAGGCCATCTCTAGGTTGGAAGGATTGAGCAATCTCTA	320
10	321	CCAAATCTATGCAGAGAGCTTCAGAGAGTGGGAAGCCGAT	360
	361	CCTACTAACCCAGCTCTCCGCGAGGAAATGCGTATTCAAT	400
	401	TCAACGACATGAACAGCGCCTTGACCACAGCTATCCCATT	440
15	441	GTTCGCAGTCCAGAACTACCAAGTTCCTCTCTTGTCCGTG	480
	481	TACGTTCAAGCAGCTAATCTTCACCTCAGCGTGCTTCGAG	520
	521	ACGTTAGCGTGTTTGGGCAAAGGTGGGGATTTCGATGCTGC	560
20	561	AACCATCAATAGCCGTTACAACGACCTTACTAGGCTGATT	600
	601	GGAAACTACACCGACCACGCTGTTCGTTGGTACAACACTG	640
25	641	GCTTGGAGCGTGTCTGGGGTCCTGATTCTAGAGATTGGAT	680
	681	TAGATACAACCAGTTCAGGAGAGAATTGACCCTCACAGTT	720
	721	TTGGACATTGTGTCTCTTCCCGAACTATGACTCCAGAA	760
30	761	CCTACCCTATCCGTACAGTGTCCCAACTTACCAGAGAAAT	800

-129-

	801	CTATACTAACCCAGTTCTTGAGAACTTCGACGGTAGCTTC	840
5	841	CGTGGTTCTGCCCAAGGTATCGAAGGCTCCATCAGGAGCC	880
	881	CACACTTGATGGACATCTTGAACAGCATAACTATCTACAC	920
	921	CGATGCTCACAGAGGAGAGTATTACTGGTCTGGACACCAG	960
10	961	ATCATGGCCTCTCCAGTTGGATTGAGCGGGCCCGAGTTTA	1000
	1001	CCTTTCCTCTCTATGGAACATGGAACGCGCTCCACA	1040
15	1041	ACAACGTATCGTTGCTCAACTAGGTCAGGGTGTCTACAGA	1080
	1081	ACCTTGTCTTCCACCTTGTACAGAAGACCCTTCAATATCG	1120
	1121	GTATCAACAACCAGCAACTTCCGTTCTTGACGGAACAGA	1160
20	1161	GTTGCGCTATGGAACCTCTTCTAACTTGCCATCCGCTGTT	1200
	1201	TACAGAAAGAGCGGAACCGTTGATTCCCTGGACGAAATCC	1240
	1241	CACCACAGAACAACAATGTGCCACCCAGGCAAGGATTCTC	1280
25	1281	CCACAGGTTGAGCCACGTGTCCATGTTCCGTTCCGGATTTC	1320
	1321	AGCAACAGTTCCGTGAGCATCATCAGAGCTCCTATGTTCT	1360
30	1361	CTTGGATACACCGTAGTGCTGAGTTCAACAACATCATCGC	1400

-130-

	1401	ATCCGATAGTATTACTCAAATCCCTGCAGTGAAGGGAAAC	1440
5	1441	TTTCTCTTCAACGGTTCTGTCATTTTCAGGACCAGGATTCA	1480
	1481	CTGGTGGAGACCTCGTTAGACTCAACAGCAGTGGAAATAA	1520
10	1521	CATTCAGAATAGAGGGTATATTGAAGTTCCAATTCCTTC	1560
	1561	CCATCCACATCTACCAGATATAGAGTTCGTGTGAGGTATG	1600
	1601	CTTCTGTGACCCCTATTACCTCAACGTTAATTGGGGTAA	1640
15	1641	TTCATCCATCTTCTCCAATACAGTTCCAGCTACAGCTACC	1680
	1681	TCCTTGGATAATCTCCAATCCAGCGATTTTCGGTTACTTTG	1720
	1721	AAAGTGCCAATGCTTTTACATCTTCACTCGGTAACATCGT	1760
20	1761	GGGTGTTAGAACTTTAGTGGGACTGCAGGAGTGATTATC	1800
	1801	GACAGATTCGAGTTCATTCCAGTTACTGCAACACTCGAGG	1840
25	1841	CTGAATATAATCTGGAAAGAGCGCAGAAGGCGGTAATGCG	1880
	1881	CTGTTTACGTCTACAAACCAGCTTGGACTCAAGACAAATG	1920.

19. A structural gene of Claim 13 encoding the full-length insecticidal protein of *B.t.k.* HD-73 having the sequence:



-131-

	1	ATGGACAACAACCCAAACATCAACGAATGCATTCCATACA	40
5	41	ACTGCTTGAGTAACCCAGAAGTTGAAGTACTTGGTGGAGA	80
	81	ACGCATTGAAACCGGTTACACTCCCATCGACATCTCCTTG	120
10	121	TCCTTGACACAGTTTCTGCTCAGCGAGTTCGTGCCAGGTG	160
	161	CTGGGTTCGTTCTCGGACTAGTTGACATCATCTGGGGTAT	200
	201	CTTTGGTCCATCTCAATGGGATGCATTCCTGGTGCAAATT	240
15	241	GAGCAGTTGATCAACCAGAGGATCGAAGAGTTCGCCAGGA	280
	281	ACCAGGCCATCTCTAGGTTGGAAGGATTGAGCAATCTCTA	320
	321	CCAAATCTATGCAGAGAGCTTCAGAGAGTGGGAAGCCGAT	360
20	361	CCTACTAACCCAGCTCTCCGCGAGGAAATGCGTATTCAAT	400
	401	TCAACGACATGAACAGCGCCTTGACCACAGCTATCCCATT	440
25	441	GTTTCGCAGTCCAGAACTACCAAGTTCCTCTCTTGTCGGTG	480
	481	TACGTTCAAGCAGCTAATCTTCACCTCAGCGTGCTTCGAG	520
	521	ACGTTAGCGTGTTTGGGCAAAGGTGGGGATTTCGATGCTGC	560
30	561	AACCATCAATAGCCGTTACAACGACCTTACTAGGCTGATT	600

-132-

	601	GGAAACTACACCGACCACGCTGTTTCGTTGGTACAACACTG	640
5	641	GCTTGGAGCGTGTCTGGGGTCCTGATTCTAGAGATTGGAT	680
	681	TAGATACAACCAGTTCAGGAGAGAATTGACCCTCACAGTT	720
10	721	TTGGACATTGTGTCTCTCTTCCCGAACTATGACTCCAGAA	760
	761	CCTACCCTATCCGTACAGTGTCCCAACTTACCAGAGAAAT	800
	801	CTATACTAACCCAGTTCTTGAGAACTTCGACGGTAGCTTC	840
15	841	CGTGGTTCTGCCCAAGGTATCGAAGGCTCCATCAGGAGCC	880
	881	CACACTTGATGGACATCTTGAACAGCATAACTATCTACAC	920
	921	CGATGCTCACAGAGGAGAGTATTACTGGTCTGGACACCAG	960
20	961	ATCATGGCCTCTCCAGTTGGATTTCAGCGGGCCCGAGTTTA	1000
	1001	CCTTTCCTCTCTATGGA ACTATGGGAAACGCCGCTCCACA	1040
25	1041	ACAACGTATCGTTGCTCAACTAGGTCAGGGTGTCTACAGA	1080
	1081	ACCTTGTCTTCCACCTTGTACAGAAGACCCTTCAATATCG	1120
	1121	GTATCAACAACCAGCAACTTTCGTTCTTGACGGAACAGA	1160
30	1161	GTTTCGCCTATGGAACCTCTTCTAACTTGCCATCCGCTGTT	1200

-133-

	1201	TACAGAAAGAGCGGAACCGTTGATTCCTTGGACGAAATCC	1240
5	1241	CACCACAGAACAACAATGTGCCACCCAGGCAAGGATTCTC	1280
	1281	CCACAGGTTGAGCCACGTGTCCATGTTCCGTTCCGGATTTC	1320
	1321	AGCAACAGTTCCGTGAGCATCATCAGAGCTCCTATGTTCT	1360
10	1361	CTTGGATACACCGTAGTGCTGAGTTCAACAACATCATCGC	1400
	1401	ATCCGATAGTATTACTCAAATCCCTGCAGTGAAGGGAAAC	1440
15	1441	TTTCTCTTCAACGGTTCTGTCAATTCAGGACCAGGATTCA	1480
	1481	CTGGTGGAGACCTCGTTAGACTCAACAGCAGTGGAAATAA	1520
	1521	CATTCAGAATAGAGGGTATATTGAAGTTCCAATTCAC TTC	1560
20	1561	CCATCCACATCTACCAGATATAGAGTTCGTGTGAGGTATG	1600
	1601	CTTCTGTGACCCCTATTCACCTCAACGTTAATTGGGGTAA	1640
	1641	TTCATCCATCTTCTCCAATACAGTTCCAGCTACAGCTACC	1680
25	1681	TCCTTGGATAATCTCCAATCCAGCGATTTCGGTTACTTTG	1720
	1721	AAAGTGCCAATGCTTTTACATCTTCACTCGGTAACATCGT	1760
30	1761	GGGTGTTAGAACTTTAGTGGGACTGCAGGAGTGATTATC	1800

-134-

	1801	GACAGATTCGAGTTCATTCCAGTTACTGCAACACTCGAGG	1840
5	1841	CTGAATATAATCTGGAAAGAGCGCAGAAGGCGGTGAATGC	1880
	1881	GCTGTTTACGTCTACAAACCAGCTCGGCCTCAAGACCAAT	1920
10	1921	GTGACGGATTATCATATTGATCAAGTGTCCAACCTTGGTGA	1960
	1961	CCTACCTCAGCGATGAGTTCTGTCTGGATGAAAAGCGAGA	2000
	2001	ATTGTCCGAGAAAGTCAAACATGCGAAGCGACTCAGTGAT	2040
15	2041	GAACGCAATTTACTCCAAGATTCAAATTTCAAAGACATTA	2080
	2081	ATAGGCAACCAGAACGTGGGTGGGGCGGAAGTACAGGGAT	2120
	2121	TACCATCCAGGGAGGTGACGACGTGTTCAAGGAGAACTAC	2160
20	2161	GTCACACTATCAGGTACCTTTGATGAGTGCTATCCAACAT	2200
	2201	ACCTCTACCAGAAGATCGACGAGTCCAAGTTGAAAGCCTT	2240
25	2241	TACCCGTTATCAATTAAGAGGGTATATCGAAGATAGTCAA	2280
	2281	GACCTCGAGATCTACCTCATCCGCTACAATGCAAAACATG	2320
	2321	AAACAGTAAATGTGCCAGGTACGGGTTCTTATGGCCGCT	2360
30	2361	TTCAGCCCAAAGTCCAATCGGAAAGTGTGGAGAGCCGAAT	2400

-135-

	2401	CGATGCGCGCCACACCTTGAATGGAATCCTGACTTAGATT	2440
5	2441	GTTTCGTGTAGGGATGGAGAAAAGTGTGCCCATCATTCGCA	2480
	2481	TCATTTCTCCTTAGACATTGATGTAGGATGTACAGACTTA	2520
10	2521	AATGAGGACCTAGGTGTATGGGTGATCTTTAAGATTAAGA	2560
	2561	CGCAAGATGGGCACGCAAGACTAGGGAATCTAGAGTTTCT	2600
	2601	CGAAGAGAAACCATTAGTAGGAGAAGCGCTAGCTCGTGTG	2640
15	2641	AAAAGAGCGGAGAAAAAATGGAGAGACAAACGTGAGAAGT	2680
	2681	TGGAATGGGAGACCAACATCGTCTACAAAGAGGCAAAGA	2720
	2721	ATCTGTAGATGCTTTATTTGTAACTCTCAATATGATCAA	2760
20	2761	TTACAAGCGGATACGAATATTGCCATGATTCATGCGGCAG	2800
	2801	ATAAACGTGTTTCATAGCATTCGAGAAGCTTATCTGCCTGA	2840
25	2841	GCTGTCTGTGATTCCGGGTGTCAATGCGGCTATTTTGTAA	2880
	2881	GAATTAGAAGGGCGTATTTTCACTGCATTCTCCCTCTACG	2920
	2921	ATGCCAGAAACGTCATCAAGAACGGTGACTTCAACAATGG	2960
30	2961	CTTATCCTGCTGGAACGTGAAAGGGCATGTAGATGTAGAA	3000

-136-

5 3001 GAACAAAACAACCAACGTTCCGGTCCTTGTTGTTCCGGAAT 3040  
3041 GGGGAAGCAGAAGTGTCACAAGAAGTTCGTGTCTGTCCGGG 3080  
3081 TCGTGGCTATATCCTTCGTGTACAGCGTACAAGGAGGGA 3120  
10 3121 TATGGAGAAGGTTGCGTAACCATTCATGAGATCGAGAACA 3160  
3161 ATACAGACGAACTGAAGTTTAGCAACTGCGTAGAAGAGGA 3200  
3201 AATCTATCCAAATAACACGGTAACGTGTAATGATTATACT 3240  
15 3241 GTAAATCAAGAAGAATACGGAGGTGCGTACACTTCTCGTA 3280  
3281 ATCGAGGATATAACGAAGCTCCTTCCGTACCAGCTGATTA 3320  
3321 TGCGTCAGTCTATGAAGAAAAATCGTATACAGATGGACGA 3360  
20 3361 AGAGAGAATCCTTGTGAATTTAACAGAGGGTATAGGGATT 3400  
3401 ACACGCCACTACCAGTTGGTTATGTGACAAAAGAATTAGA 3440  
25 3441 ATACTTCCCAGAAACCGATAAGGTATGGATTGAGATTGGA 3480  
3481 GAAACGGAAGGAACATTTATCGTGGACAGCGTGGAATTAC 3520  
3521 TCCTTATGGAGGAA 3534.

30

-137-

20. A structural gene of Claim 13 encoding a full-length insecticidal protein of *B.t.k.* HD-73 having the sequence:

5

```
      .       .       .       .
1  ATGGACAACAACCCAAACATCAACGAATGCATTCCATACA  40
      .       .       .       .
41 ACTGCTTGAGTAACCCAGAAGTTGAAGTACTTGGTGGAGA  80
      .       .       .       .
10 81 ACGCATTGAAACCGGTTACTCTCCATCGACATCTCCTTG  120
      .       .       .       .
121 TCCTTGACACAGTTTCTGCTCAGCGAGTTCGTGCCAGGTG  160
      .       .       .       .
161 CTGGGTTCGTTCTCGGACTAGTTGACATCATCTGGGGTAT  200
15
201 CTTTGGTCCATCTCAATGGGATGCATTCCTGGTGCAAATT  240
      .       .       .       .
241 GAGCAGTTGATCAACCAGAGGATCGAAGAGTTCGCCAGGA  280
      .       .       .       .
20 281 ACCAGGCCATCTCTAGGTTGGAAGGATTGAGCAATCTCTA  320
      .       .       .       .
321 CCAAATCTATGCAGAGAGCTTCAGAGAGTGGGAAGCCGAT  360
      .       .       .       .
361 CCTACTAACCCAGCTCTCCGCGAGGAAATGCGTATTCAAT  400
25
401 TCAACGACATGAACAGCGCCTTGACCACAGCTATCCCATT  440
      .       .       .       .
441 GTTCGCAGTCCAGAACTACCAAGTTCCTCTCTTGTCCGTG  480
      .       .       .       .
481 TACGTTCAAGCAGCTAATCTTCACCTCAGCGTGCTTCGAG  520
30
```

-138-

	521	ACGTTAGCGTGTTTGGGCAAAGGTGGGGATTGATGCTGC	560
5	561	AACCATCAATAGCCGTTACAACGACCTTACTAGGCTGATT	600
	601	GGAAACTACACCGACCACGCTGTTCGTTGGTACAACACTG	640
10	641	GCTTGGAGCGTGCTCTGGGGTCCTGATTCTAGAGATTGGAT	680
	681	TAGATACAACCAGTTCAGGAGAGAATTGACCCTCACAGTT	720
	721	TTGGACATTGTGTCTCTCTTCCCGAACTATGACTCCAGAA	760
15	761	CCTACCCTATCCGTACAGTGTCCCAACTTACCAGAGAAAT	800
	801	CTATACTAACCCAGTTCTTGAGAACTTCGACGGTAGCTTC	840
	841	CGTGGTTCTGCCCCAAGGTATCGAAGGCTCCATCAGGAGCC	880
20	881	CACACTTGATGGACATCTTGAACAGCATAACTATCTACAC	920
	921	CGATGCTCACAGAGGAGAGTATTACTGGTCTGGACACCAG	960
25	961	ATCATGGCCTCTCCAGTTGGATTGAGCGGGCCCGAGTTTA	1000
	1001	CCTTTCCTCTCTATGGAACCTATGGGAAACGCCGCTCCACA	1040
	1041	ACAACGTATCGTTGCTCAACTAGGTCAGGGTGTCTACAGA	1080
30	1081	ACCTTGTCTTCCACCTTGTACAGAAGACCCTTCAATATCG	1120



-139-

	1121	GTATCAACAACCAGCAACTTTCCGTTCTTGACGGAACAGA	1160
5	1161	GTTTCGCCTATGGAACCTCTTCTAACTTGCCATCCGCTGTT	1200
	1201	TACAGAAAGAGCGGAACCGTTGATTCCTTGGACGAAATCC	1240
	1241	CACCACAGAACAACAATGTGCCACCCAGGCAAGGATTCTC	1280
10	1281	CCACAGGTTGAGCCACGTGTCCATGTTCCGTTCCGGATTC	1320
	1321	AGCAACAGTTCGCTGAGCATCATCAGAGCTCCTATGTTCT	1360
15	1361	CTTGGATACACCGTAGTGCTGAGTTCAACAACATCATCGC	1400
	1401	ATCCGATAGTATTACTCAAATCCCTGCAGTGAAGGGAAAC	1440
	1441	TTTCTCTTCAACGGTTCTGTCAATTCAGGACCAGGATTCA	1480
20	1481	CTGGTGGAGACCTCGTTAGACTCAACAGCAGTGGAAATAA	1520
	1521	CATTCAGAATAGAGGGTATATTGAAGTTCCAATTCACTTC	1560
	1561	CCATCCACATCTACCAGATATAGAGTTCGTGTGAGGTATG	1600
25	1601	CTTCTGTGACCCCTATTCACCTCAACGTTAATTGGGGTAA	1640
	1641	TTCATCCATCTTCTCCAATACAGTTCCAGCTACAGCTACC	1680
30	1681	TCCTTGGATAATCTCCAATCCAGCGATTTTCGGTTACTTTG	1720

-140-

	1721	AAAGTGCCAATGCTTTTACATCTTCACTCGGTAACATCGT	1760
5	1761	GGGTGTTAGAACTTTAGTGGGACTGCAGGAGTGATTATC	1800
	1801	GACAGATTCGAGTTCATTCCAGTTACTGCAACACTCGAGG	1840
10	1841	CTGAATATAATCTGGAAAGAGCGCAGAAGGCGGTGAATGC	1880
	1881	GCTGTTTACGTCTACAAACCAACTAGGGCTAAAAACAAAT	1920
	1921	GTAACGGATTATCATATTGATCAAGTGTCCAATTTAGTTA	1960
15	1961	CGTATTTATCGGATGAATTTTGTCTGGATGAAAAGCGAGA	2000
	2001	ATTGTCCGAGAAAGTCAAACATGCGAAGCGACTCAGTGAT	2040
	2041	GAACGCAATTTACTCCAAGATTCAAATTTCAAAGACATTA	2080
20	2081	ATAGGCAACCAGAACGTGGGTGGGGCGGAAGTACAGGGAT	2120
	2121	TACCATCCAAGGAGGGGATGACGTATTTAAAGAAAATTAC	2160
25	2161	GTCACACTATCAGGTACCTTTGATGAGTGCTATCCAACAT	2200
	2201	ATTTGTATCAAAAAATCGATGAATCAAATTTAAAAGCCTT	2240
	2241	TACCCGTTATCAATTAAGAGGGTATATCGAAGATAGTCAA	2280
30	2281	GACTTAGAAATCTATTTAATTCGCTACAATGCAAAACATG	2320

-141-

	2321	AAACAGTAAATGTGCCAGGTACGGGTTCCCTTATGGCCGCT	2360
5	2361	TTCAGCCCCAAAGTCCAATCGGAAAGTGTGGAGAGCCGAAT	2400
	2401	CGATGCGCGCCACACCTTGAATGGAATCCTGACTTAGATT	2440
	2441	GTTCGTGTAGGGATGGAGAAAAGTGTGCCCATCATTGCGCA	2480
10	2481	TCATTTCTCCTTAGACATTGATGTAGGATGTACAGACTTA	2520
	2521	AATGAGGACCTAGGTGTATGGGTGATCTTTAAGATTAAGA	2560
15	2561	CGCAAGATGGGCACGCAAGACTAGGGAATCTAGAGTTTCT	2600
	2601	CGAAGAGAAACCATTAGTAGGAGAAGCGCTAGCTCGTGTG	2640
	2641	AAAAGAGCGGAGAAAAAATGGAGAGACAAACGTGAAAAAT	2680
20	2681	TGGAATGGGAAACAAATATCGTTTATAAAGAGGCAAAAGA	2720
	2721	ATCTGTAGATGCTTTATTTGTAACTCTCAATATGATCAA	2760
	2761	TTACAAGCGGATACGAATATTGCCATGATTCATGCGGCAG	2800
25	2801	ATAAACGTGTTTCATAGCATTGCGAGAAGCTTATCTGCCTGA	2840
	2841	GCTGTCTGTGATTCCGGGTGTCAATGCGGCTATTTTGTAA	2880
30	2881	GAATTAGAAGGGCGTATTTTCACTGCATTCTCCCTATATG	2920

-142-

2921 ATGCGAGAAATGTCATTAAAAATGGTGATTTTAATAATGG 2960  
5 2961 CTTATCCTGCTGGAACGTGAAAGGGCATGTAGATGTAGAA 3000  
3001 GAACAAAACAACCAACGTTCGGTCCTTGTTGTTCCGGAAT 3040  
3041 GGGAAAGCAGAAGTGTACACAAGAAGTTCGTGTCTGTCCGGG 3080  
10 3081 TCGTGGCTATATCCTTCGTGTACAGCGTACAAGGAGGGA 3120  
3121 TATGGAGAAGGTTGCGTAACCATTTCATGAGATCGAGAACA 3160  
15 3161 ATACAGACGAACTGAAGTTTAGCAACTGCGTAGAAGAGGA 3200  
3201 AATCTATCCAAATAACACGGTAACGTGTAATGATTATACT 3240  
3241 GTAAATCAAGAAGAATACGGAGGTGCGTACACTTCTCGTA 3280  
20 3281 ATCGAGGATATAACGAAGCTCCTTCCGTACCAGCTGATTA 3320  
3321 TCGGTCAGTCTATGAAGAAAAATCGTATACAGATGGACGA 3360  
25 3361 AGAGAGAATCCTTGTGAATTTAACAGAGGGTATAGGGATT 3400  
3401 ACACGCCACTACCAGTTGGTTATGTGACAAAAGAATTAGA 3440  
3441 ATACTTCCCAGAAACCGATAAGGTATGGATTGAGATTGGA 3480  
30

-143-

3481 GAAACGGAAGGAACATTTATCGTGGACAGCGTGGAATTAC 3520  
5 3521 TCCTTATGGAGGAA 3534.

21. A structural gene of Claim 13 encoding a full-length insecticidal protein of *B.t.k.* HD-73 having the sequence:

10  
1 ATGGACAACAACCCAAACATCAACGAATGCATTCCATACA 40  
41 ACTGCTTGAGTAACCCAGAAGTTGAAGTACTTGGTGGAGA 80  
15 81 ACGCATTGAAACCGGTTACACTCCCATCGACATCTCCTTG 120  
121 TCCTTGACACAGTTTCTGCTCAGCGAGTTCGTGCCAGGTG 160  
161 CTGGGTTCGTTCTCGGACTAGTTGACATCATCTGGGGTAT 200  
20 201 CTTTGGTCCATCTCAATGGGATGCATTCCTGGTGCAAATT 240  
241 GAGCAGTTGATCAACCAGAGGATCGAAGAGTTCGCCAGGA 280  
25 281 ACCAGGCCATCTCTAGGTTGGAAGGATTGAGCAATCTCTA 320  
321 CCAAATCTATGCAGAGAGCTTCAGAGAGTGGGAAGCCGAT 360  
361 CCTACTAACCCAGCTCTCCGCGAGGAAATGCGTATTCAAT 400  
30 401 TCAACGACATGAACAGCGCCTTGACCACAGCTATCCCATT 440

-144-

441 GTTCGCAGTCCAGAACTACCAAGTTCCTCTCTTGTCCGTG 480  
5 481 TACGTTCAAGCAGCTAATCTTCACCTCAGCGTGCTTCGAG 520  
521 ACGTTAGCGTGTTTGGGCAAAGGTGGGGATTTCGATGCTGC 560  
561 AACCATCAATAGCCGTTACAACGACCTTACTAGGCTGATT 600  
10 601 GGAAACTACACCGACCACGCTGTTTCGTTGGTACAACACTG 640  
641 GCTTGGAGCGTGTCTGGGGTCCTGATTCTAGAGATTGGAT 680  
681 TAGATAACAACAGTTCAGGAGAGAATTGACCCTCACAGTT 720  
15 721 TTGGACATTGTGTCTCTCTTCCCGAACTATGACTCCAGAA 760  
761 CCTACCCTATCCGTACAGTGTCCTCAACTTACCAGAGAAAT 800  
20 801 CTATACTAACCCAGTTCTTGAGAACTTCGACGGTAGCTTC 840  
841 CGTGGTTCTGCCCCAAGGTATCGAAGGCTCCATCAGGAGCC 880  
881 CACACTTGATGGACATCTTGAACAGCATAACTATCTACAC 920  
25 921 CGATGCTCACAGAGGAGAGTATTACTGGTCTGGACACCAG 960  
961 ATCATGGCCTCTCCAGTTGGATTTCAGCGGGCCCGAGTTTA 1000  
30 1001 CCTTTCCTCTCTATGGAAGTATGGGAAACGCCGCTCCACA 1040

-145-

	1041	ACAACGTATCGTTGCTCAACTAGGTCAGGGTGTCTACAGA	1080
5	1081	ACCTTGTCTTCCACCTTGTACAGAAGACCCTTCAATATCG	1120
	1121	GTATCAACAACCAGCAACTTTCGGTTCTTGACGGAACAGA	1160
	1161	GTTGCGCTATGGAACCTCTTCTAACTTGCCATCCGCTGTT	1200
10	1201	TACAGAAAGAGCGGAACCGTTGATTCTTGGACGAAATCC	1240
	1241	CACCACAGAACAACAATGTGCCACCCAGGCAAGGATTCTC	1280
15	1281	CCACAGGTTGAGCCACGTGTCCATGTTCCGTTCCGGATTC	1320
	1321	AGCAACAGTTCCGTGAGCATCATCAGAGCTCCTATGTTCT	1360
	1361	CTTGGATACACCGTAGTGCTGAGTTCAACAACATCATCGC	1400
20	1401	ATCCGATAGTATTACTCAAATCCCTGCAGTGAAGGGAAAC	1440
	1441	TTTCTCTTCAACGGTCTGTTCATTTAGGACCAGGATTCA	1480
	1481	CTGGTGGAGACCTCGTTAGACTCAACAGCAGTGGAAATAA	1520
25	1521	CATTCAGAATAGAGGGTATATTGAAGTTCCAATTCCTTC	1560
	1561	CCATCCACATCTACCAGATATAGAGTTCGTGTGAGGTATG	1600
30	1601	CTTCTGTGACCCCTATTACCTCAACGTTAATTGGGGTAA	1640

-146-

	1641	TTCATCCATCTTCTCCAATACAGTTCCAGCTACAGCTACC	1680
5	1681	TCCTTGGATAATCTCCAATCCAGCGATTTCGGTTACTTTG	1720
	1721	AAAGTGCCAATGCTTTTACATCTTCACTCGGTAACATCGT	1760
	1761	GGGTGTTAGAACTTTAGTGGGACTGCAGGAGTGATTATC	1800
10	1801	GACAGATTCGAGTTCATTCCAGTTACTGCAACACTCGAGG	1840
	1841	CTGAGTACAACCTTGAGAGAGCCCAGAAGGCTGTGAACGC	1880
15	1881	CCTCTTTACCTCCACCAATCAGCTTGGCTTGAAAATAAC	1920
	1921	GTTACTGACTATCACATTGACCAAGTGTCCAAGTTGGTCA	1960
	1961	CCTACCTTAGCGATGAGTTCTGCCTCGACGAGAAGCGTGA	2000
20	2001	ACTCTCCGAGAAAGTTAAACACGCCAAGCGTCTCAGCGAC	2040
	2041	GAGAGGAATCTCTTGCAAGACTCCAAGTTCAAAGACATCA	2080
25	2081	ACAGGCAGCCAGAACGTGGTTGGGGTGGAAGCACCGGGAT	2120
	2121	CACCATCCAAGGAGGCGACGATGTGTTCAAGGAGAACTAC	2160
	2161	GTCACCCTCTCCGGAAGTTTCGACGAGTGCTACCCTACCT	2200
30	2201	ACTTGTACCAGAAGATCGATGAGTCCAAAGTCAAAGCCTT	2240



-147-

	2241	CACCAGGTATCAACTTAGAGGCTACATCGAAGACAGCCAA	2280
5	2281	GACCTTGAAATCTACTCGATCAGGTACAATGCCAAGCACG	2320
	2321	AGACCGTGAATGTCCCAGGTACTGGTTCCTCTGGCCACT	2360
10	2361	TTCTGCCCAATCTCCCATTGGGAAGTGTGGAGAGCCTAAC	2400
	2401	AGATGCGCTCCACACCTTGAGTGGAATCCTGACTTGGACT	2440
	2441	GCTCCTGCAGGGATGGCGAGAAGTGTGCCCACCATTCTCA	2480
15	2481	TCACTTCTCCTTGGACATCGATGTGGGATGTACTGACCTG	2520
	2521	AATGAGGACCTCGGAGTCTGGGTCATCTTCAAGATCAAGA	2560
	2561	CCCAAGACGGACACGCAAGACTTGGCAACCTTGAGTTTCT	2600
20	2601	CGAAGAGAAACCATTGGTCGGTGAAGCTCTCGCTCGTGTG	2640
	2641	AAGAGAGCAGAGAAGAAGTGGAGGGACAAACGTGAGAAAC	2680
25	2681	TCGAATGGGAAACTAACATCGTTTACAAGGAGGCCAAAGA	2720
	2721	GTCCGTGGATGCTTTGTTTCGTGAACTCCCAATATGATCAG	2760
	2761	TTGCAAGCCGACACCAACATCGCCATGATCCACGCCGCAG	2800
30	2801	ACAAACGTGTGCACAGCATTCGTGAGGCTTACTTGCCTGA	2840

-148-

	2841	GTTGTCCGTGATCCCTGGTGTGAACGCTGCCATCTTCGAG	2880
5	2881	GAAGTTGAGGGACGTATCTTTACCGCATTCTCCTTGTACG	2920
	2921	ATGCCAGAAACGTCATCAAGAACGGTGACTTCAACAATGG	2960
10	2961	CCTCAGCTGCTGGAATGTGAAAGGTCATGTGGACGTGGAG	3000
	3001	GAACAGAACAATCAGCGTTCCGTCCTGGTTGTGCCTGAGT	3040
	3041	GGGAAGCTGAAGTGTCCCAAGAGGTTAGAGTCTGTCCAGG	3080
15	3081	TAGAGGCTACATTCTCCGTGTGACCGCTTACAAGGAGGGA	3120
	3121	TACGGTGAGGGTTGCGTGACCATCCACGAGATCGAGAACA	3160
	3161	ACACCGACGAGCTTAAGTTCTCCAAGTTCGAGGAAGA	3200
20	3201	AATCTATCCCAACAACACCGTTACTTGCAACGACTACACT	3240
	3241	GTGAATCAGGAAGAGTACGGAGGTGCCTACACTAGCCGTA	3280
25	3281	ACAGAGGTTACAACGAAGCTCCTTCCGTTCCCTGCTGACTA	3320
	3321	TGCCTCCGTGTACGAGGAGAAATCCTACACAGATGGCAGA	3360
	3361	CGTGAGAACCCTTGCGAGTTCAACAGAGGTTACAGGGACT	3400
30	3401	ACACACCACTTCCAGTTGGCTATGTTACCAAGGAGCTTGA	3440

-149-

3441 GTACTTTCCTGAGACCGACAAAGTGTGGATCGAGATCGGT 3480  
5 3481 GAAACCGAGGGAACTTCATCGTGGACAGCGTGGAGCTTC 3520  
3521 TCTTGATGGAGGAA 3534.

22. A structural gene of Claim 13 which encodes an  
10 insecticidal protein of *B.t.t.* having the sequence:

1 ATGACTGCAGACAACAACACCGAAGCCCTCGACAGTTCTA 40  
41 CCACTAAGGATGTTATCCAGAAGGGTATCTCCGTTGTGGG 80  
15 81 AGACCTCTTGGGCGTGGTTGGATTTCCTTCGGTGGAGCC 120  
121 CTCGTGAGCTTCTATACAACTTTCTCAACACCATTGTC 160  
161 CAAGCGAGGACCCTTGGAAAGCATTCATGGAGCAAGTTGA 200  
20 201 AGCTCTTATGGATCAGAAGATTGCAGATTATGCCAAGAAC 240  
241 AAGGCTTTGGCAGAACTCCAGGGCCTTCAGAACAAATGTGG 280  
25 281 AGGACTACGTGAGTGCATTGTCCAGCTGGCAGAAGAACCC 320  
321 TGTTAGCTCCAGAAATCCTCACAGCCAAGGTAGGATCAGA 360  
361 GAGTTGTTCTCTCAAGCCGAATCCCACTTCAGAAATTCCA 400  
30

-150-

	401	TGCCTAGCTTTGCTATCTCCGGTTACGAGGTTCTTTTCCT	440
5	441	CACTACCTATGCTCAAGCTGCCAACACCCACTTGTTTCTC	480
	481	CTTAAGGACGCTCAAATCTATGGAGAAGAGTGGGGATACG	520
10	521	AGAAAGAGGACATTGCTGAGTTCTACAAGCGTCAACTTAA	560
	561	GCTCACCCAAGAGTACACTGACCATTGCGTGAAATGGTAT	600
	601	AACGTTGGTCTCGATAAGCTCAGAGGCTCTTCCTACGAGT	640
15	641	CTTGGGTGAACTTCAACAGATACAGGAGAGAGATGACCTT	680
	681	GACTGTGCTCGATCTTATCGCACTCTTCCCTTGACGAT	720
	721	GTGAGACTCTACCCAAAGGAAGTGAAAAGTGAAGCTTACCA	760
20	761	GAGACGTGCTCACTGACCCTATTGTGCGAGTCAACAACCT	800
	801	TAGGGGTTATGGAACTACCTTCAGCAATATCGAAAAGTAC	840
25	841	ATTAGGAAACCACATCTCTTCGACTATCTTCACAGAATTC	880
	881	AATTCACACAAGGTTTCAACCAGGATACTATGGTAACGA	920
	921	CTCCTTCAACTATTGGTCCGGTAACTATGTTTCCACCAGA	960
30	961	CCAAGCATTGGATCTAATGACATCATCACATCTCCCTTCT	1000

-151-

1001 ATGGTAACAAGTCCAGTGAACCTGTGCAGAACCTTGAGTT 1040  
5 1041 CAACGGCGAGAAAGTCTATAGAGCCGTCGCAAACACCAAT 1080  
1081 CTCGCTGTGTGGCCATCCGCAGTTTACTCAGGCGTCACAA 1120  
1121 AGGTGGAGTTTAGTCAGTATAACGATCAGACCGATGAGGC 1160  
10 1161 CAGCACCCAGACTTACGACTCCAAACGTAACGTTGGCGCA 1200  
1201 GTCTCTTGGGATTCTATCGACCAATTGCCTCCAGAAACCA 1240  
15 1241 CAGACGAACCATTGGAGAAGGGCTACAGCCACCAACTTAA 1280  
1281 CTATGTGATGTGCTTCTTGATGCAAGGTTCCAGAGGGACC 1320  
1321 ATTCCAGTGTTGACCTGGACACACAAGTCCGTGGACTTCT 1360  
20 1361 TCAACATGATCGATAGCAAGAAGATCACTCAACTTCCCTT 1400  
1401 GGTGAAAGCCTACAAGCTGCAATCTGGTGCTTCCGTTGTC 1440  
1441 GCAGGTCCCAGATTCAGTGGAGGTGACATCATCCAGTGCA 1480  
25 1481 CAGAGAACGGCAGCGCAGCTACTATCTACGTGACACCTGA 1520  
1521 TGTGTCTTACTCTCAGAAGTACAGGGCACGTATTCATTAC 1560  
30 1561 GCATCTACCAGCCAGATCACCTTCACACTCAGCTTGGATG 1600

-152-

1601 GAGCACCCCTTCAACCAGTATTACTTTGACAAGACCATCAA 1640  
5 1641 CAAAGGTGACACTCTCACATAACAATAGCTTCAACTTGGCA 1680  
1681 AGTTTCAGCACACCATTGGAAGTCTCAGGCAACAATCTTC 1720  
1721 AGATCGGCGTCACCGGTCTCAGCGCCGGAGACAAAGTCTA 1760  
10 1761 CATCGACAAGATTGAGTTCATCCCAGTGAAC 1791.

23. A structural gene of Claim 13 which encodes an  
insecticidal protein of *B.t. entomocidus* having the  
15 sequence:

1 ATGGAGGAGAACAACCAAAACCAATGCATTCCATACAAC 40  
41 GCTTGAGTAACCCAGAAGAGGTATTGCTTGATGGAGAACG 80  
20 81 CATTTCAACCGGTAAGTCTTCCATCGACATCTCCTTGTCC 120  
121 TTGGTCCAGTTTCTGGTCAGCAACTTCGTGCCAGGTGGTG 160  
161 GGTTCCCTTGTCGGACTAATTGACTTCGTCTGGGGTATCGT 200  
25 201 TGGTCCATCTCAATGGGATGCATTCCTGGTGCAAATTGAG 240  
241 CAGTTGATCAACGAGAGGATCGCTGAGTTCGCCAGGAACG 280  
30 281 CTGCCATCGCTAACTTGAAGGATTGGGCAATAACTTCAA 320

-153-

	321	CATCTATGTGGAGGCCTTCAAAGAGTGGGAAGAGGACCCT	360
5	361	AACAACCCAGAGACCCGCACTAGGGTGATCGACAGATTCA	400
	401	GAATCTTGGACGGCCTCTTGGAGAGAGATATCCCATCCTT	440
10	441	CAGAATCTCTGGCTTCGAAGTTCCTCTCTTGTCGGTGAC	480
	481	GCTCAAGCAGCTAATCTTCACCTCGCTATCCTTCGAGACA	520
	521	GTGTCATCTTTGGGGAAAGGTGGGGATTGACCACTATCAA	560
15	561	CGTCAATGAGAATTACAACAGACTTATCAGGCACATTGAC	600
	601	GAGTACGCCGACCACTGTGCTAACACCTACAACCGTGGCT	640
	641	TGAACAATCTCCCTAAGTCTACTTATCAAGATTGGATTAC	680
20	681	CTACAACAGGTTGAGGAGAGACTTGACCCTCACAGTTTTG	720
	721	GACATTGCAGCTTTCTTCCCGAACTATGACAACAGGAGAT	760
25	761	ACCCTATCCAACCACTGGGTCAACTTACCAGAGAAGTCTA	800
	801	TACTGACCCACTTATCAACTTCAACCCTCAGTTGCAAAGT	840
	841	GTCGCCCAACTTCCCACATTCAACGTCATGGAGTCCAGCC	880
30	881	GTATCAGGAACCCACACTTGTTTGACATCTTGAACAACCT	920

-154-

	921	TACTATCTTCACCGATTGGTTCAGCGTTGGGCGTAACTTC	960
5	961	TATTGGGGTGGACACAGGGTCATCTCCTCTCTTATTGGAG	1000
	1001	GTGGGAACATTACCTCTCCTATCTATGGACGTGAGGCAA	1040
10	1041	CCAGGAGCCACCACGTAGTTTCACCTTCAACGGTCCAGTC	1080
	1081	TTCAGAACCTTGTCTAACCTACCTTGAGATTGCTCCAGC	1120
	1121	AACCTTGGCCAGCTCCACCTTCAACCTTAGAGGTGTTGA	1160
15	1161	GGGCGTTGAGTTCTCTACTCCTACCAACTCCTTCACTTAC	1200
	1201	AGAGGTAGAGGAACCGTTGATTCCTTGACCGAACTCCCAC	1240
	1241	CAGAGGACAATAGCGTGCCACCCAGGGAAGGCTACTCCCA	1280
20	1281	CAGGTTGTGCCACGCAACCTTCGTGCAGCGTTCCGGAAC	1320
	1321	CCATTCTCTACTACAGGAGTTGTGTTCTCATGGACTGATC	1360
25	1361	GTAGTGCTACTCTCACTAATACCATTGATCCCGAGAGGAT	1400
	1401	CAATCAAATCCCATTGGTCAAGGGTTTCCGTGTGTGGGGA	1440
	1441	GGAAGTTCTGTGTCATCACAGGACCAGGCTTACAGGAGGTG	1480
30	1481	ATATTCTTAGAAGAAACACTTTTGGCGACTTTGTGAGCCT	1520



-155-

	1521	CCAAGTTAACATCAACTCTCCAATTACTCAAAGATATCGT	1560
5	1561	CTCAGGTTTCGTTACGCATCTTCCCGTGACGCTAGAGTCA	1600
	1601	TCGTGCTCACCGGAGCAGCTTCTACCGGTGTCGGTGGACA	1640
	1641	AGTCTCCGTGAACATGCCACTCCAGAAGACTATGGAGATC	1680
10	1681	GGCGAGAACTTGACATCCAGGACCTTCAGATACACCGACT	1720
	1721	TCTCTAACCCTTTTCAGTTTCCGTGCCAACCCTGACATCAT	1760
15	1761	TGGCATTAGCGAACAACCTCTCTTTGGAGCTGGTAGCATC	1800
	1801	TCATCTGGCGAATTGTACATTGACAAGATTGAGATCATTC	1840
	1841	TTGCCGACGCTACCTTCGAGGCTGAGTCTGACCTTGAGAG	1880
20	1881	AGCCCAGAAGGCTGTGAACGCCCTCTTTACCTCCTCTAAT	1920
	1921	CAGATTGGCTTGAAAAC TGACGTTACTGACTATCACATTG	1960
25	1961	ACCAAGTGTCCAAC TTGGTCGACTGCCTTAGCGATGAGTT	2000
	2001	CTGCCTCGACGAGAAGCGTGAAC TCTCCGAGAAAGTTAAA	2040
	2041	CACGCCAAGCGTCTCAGCGACGAGAGGAATCTCTTGCAAG	2080
30	2081	ACCCCAACTTCAGAGGCATCAACAGGCAGCCAGACCGTGG	2120

-156-

	2121	TTGGAGAGGAAGCACCGACATCACCATCCAAGGAGGCGAC	2160
5	2161	GATGTGTTCAAGGAGAACTACGTCACCCTCCCAGGAACTG	2200
	2201	TGGACGAGTGCTACCCTACCTACTTGTACCAGAAGATCGA	2240
	2241	TGAGTCCAAACTCAAAGCCTACACCAGGTATGAACTTAGA	2280
10	2281	GGCTACATCGAAGACAGCCAAGACCTTGAAATCTACCTCA	2320
	2321	TCAGGTACAATGCCAAGCACGAGATCGTGAATGTCCCAGG	2360
15	2361	TACTGGTTCCTCTGGCCACTTTCTGCCCAAATGCCCAT	2400
	2401	GGGAAGTGTGGAGAGCCTAACAGATGCGCTCCACACCTTG	2440
	2441	AGTGGAATCCTGACTTGGA CTGCTCCTGCAGGGATGGCGA	2480
20	2481	GAAGTGTGCCCACCATTCTCATCACTTCACCTTGGACATC	2520
	2521	GATGTGGGATGTACTGACCTGAATGAGGACCTCGGAGTCT	2560
	2561	GGGTCATCTTCAAGATCAAGACCCAAGACGGACACGCAAG	2600
25	2601	ACTTGGCAACCTTGAGTTTCTCGAAGAGAAACCATTGCTC	2640
	2641	GGTGAAGCTCTCGCTCGTGTGAAGAGAGCAGAGAAGAAGT	2680
30	2681	GGAGGGACAAACGTGAGAACTCCAAC TCGAGACTAACAT	2720

-157-

	2721	CGTTTACAAGGAGGCCAAAGAGTCCGTGGATGCTTTGTTC	2760
5	2761	GTGAACTCCCAATATGATAGGTTGCAAGTGGACACCAACA	2800
	2801	TCGCCATGATCCACGCTGCAGACAAACGTGTGCACAGGAT	2840
	2841	TCGTGAGGCTTACTTGCCTGAGTTGTCCGTGATCCCTGGT	2880
10	2881	GTGAACGCTGCCATCTTCGAGGAACTTGAGGGACGTATCT	2920
	2921	TTACCGCATACTCCTTGTACGATGCCAGAAACGTCATCAA	2960
15	2961	GAACGGTGACTTCAACAATGGCCTCTTGTGCTGGAATGTG	3000
	3001	AAAGGTCATGTGGACGTGGAGGAACAGAACAATCACCGTT	3040
	3041	CCGTCCTGGTTATCCCTGAGTGGGAAGCTGAAGTGTCCCA	3080
20	3081	AGAGGTTAGAGTCTGTCCAGGTAGAGGCTACATTCTCCGT	3120
	3121	GTGACCGCTTACAAGGAGGGATACGGTGAGGGTTGCGTGA	3160
	3161	CCATCCACGAGATCGAGGACAACACCGACGAGCTTAAGTT	3200
25	3201	CTCCAACGCGTCGAGGAAGAAGTCTATCCCAACAACACC	3240
	3241	GTTACTTGCAACAACACTACTGGGACCCAGGAAGAGTACG	3280
30	3281	AAGGTACCTACACTAGCCGTAACCAAGGTTACGACGAAGC	3320

-158-

3321 TTACGGAAACAATCCTTCCGTTCTGCTGACTATGCCTCC 3360  
5 3361 GTGTACGAGGAGAAATCCTACACAGATGGCAGACGTGAGA 3400  
3401 ACCCTTGCGAGTCCAACAGAGGTTACGGTGACTACACACC 3440  
10 3441 ACTTCCAGCAGGCTATGTTACCAAGGACCTTGAGTACTTT 3480  
3481 CCTGAGACCGACAAAGTGTGGATCGAGATCGGTGAAACCG 3520  
3521 AGGGAACCTTCATCGTGGACAGCGTGGAGCTTCTCTTGAT 3560  
15 3561 GGAGGAA 3567.

24. A structural gene of Claim 13 which encodes a P2 insecticidal protein having the sequence:

20 1 ATGGACAACAACGTCTTGA ACTCTGGTAGAACAACCATCT 40  
41 GCGACGCATACAACGTCGTGGCTCACGATCCATTCAGCTT 80  
81 CGAACACAAGAGCCTCGACACTATTCAGAAGGAGTGGATG 120  
25 121 GAATGGAAACGTACTGACCACTCTCTCTACGTGCGACCTG 160  
161 TGGTTGGAACAGTGTCCAGCTTCCTTCTCAAGAAGGTCGG 200  
201 CTCTCTCATCGGAAAACGTATCTTGTCCGA ACTCTGGGGT 240  
30

-159-

	241	ATCATCTTTCCATCTGGGTCCACTAATCTCATGCAAGACA	280
5	281	TCTTGAGGGAGACCGAACAGTTTCTCAACCAGCGTCTCAA	320
	321	CACTGATACCTTGGCTAGAGTCAACGCTGAGTTGATCGGT	360
	361	CTCCAAGCAAACATTCGTGAGTTCAACCAGCAAGTGGACA	400
10	401	ACTTCTTGAATCCAACCTCAGAATCCTGTGCCTCTTTCCAT	440
	441	CACTTCTTCCGTGAACACTATGCAGCAACTCTTCCTCAAC	480
15	481	AGATTGCCTCAGTTTCAGATTCAAGGCTACCAGTTGCTCC	520
	521	TTCTTCCACTCTTTGCTCAGGCTGCCAACATGCACTTGTC	560
	561	CTTCATACGTGACGTGATCCTCAACGCTGACGAATGGGGA	600
20	601	ATCTCTGCAGCCACTCTTAGGACATACAGAGACTACTTGA	640
	641	GGAACTACACTCGTGATTACTCCAACCTATTGCATCAACAC	680
25	681	TTATCAGACTGCCTTTCGTGGACTCAATACTAGGCTTCAC	720
	721	GACATGCTTGAGTTCAGGACCTACATGTTCCCTTAACGTGT	760
	761	TTGAGTACGTCAGCATTTGGAGTCTCTTCAAGTACCAGAG	800
30	801	CTTGATGGTGTCCCTCTGGAGCCAATCTCTACGCCTCTGGC	840

-160-

	841	AGTGGACCACAGCAAACCTCAGAGCTTCACAGCTCAGAACT	880
5	881	GGCCATTCTTGTATAGCTTGTTCCAAGTCAACTCCAATA	920
	921	CATTCTCAGTGGTATCTCTGGGACCAGACTCTCCATAACC	960
	961	TTTCCCAACATTGGTGGACTTCCAGGCTCCACTACAACCC	1000
10	1001	ATAGCCTTAACTCTGCCAGAGTGAACCTACAGTGGAGGTGT	1040
	1041	CAGCTCTGGATTGATTGGTGCAACTAACTTGAACCACAAC	1080
15	1081	TTCAATTGCTCCACCGTCTTGCCACCTCTGAGCACACCGT	1120
	1121	TTGTGAGGTCCTGGCTTGACAGCGGTACTGATCGCGAAGG	1160
	1161	AGTTGCTACCTCTACAACTGGCAAACCGAGTCCTTCCAA	1200
20	1201	ACCACTCTTAGCCTTCGGTGTGGAGCTTTCTCTGCACGTG	1240
	1241	GGAATTCAAACCTACTTTCCAGACTACTTCATTAGGAACAT	1280
25	1281	CTCTGGTGTTCCTCTCGTCATCAGGAATGAAGACCTCACC	1320
	1321	CGTCCACTTCATTACAACCAGATTAGGAACATCGAGTCTC	1360
	1361	CATCCGGTACTCCAGGAGGTGCAAGAGCTTACCTCGTGTC	1400
30	1401	TGTCCATAACAGGAAGAACAACATCTACGCTGCCAACGAG	1440

-161-

1441 AATGGCACCATGATTCACCTTGCACCAGAAGATTACACTG 1480  
5 1481 GATTCACCATCTCTCCAATCCATGCTACCCAAGTGAACAA 1520  
1521 TCAGACACGCACCTTCATCTCCGAAAAGTTCGGAAATCAA 1560  
1561 GGTGACTCCTTGAGGTTGAGCAATCCAACACTACCGCTA 1600  
10 1601 GGTACACTTTGAGAGGCAATGGAAACAGCTACAACCTTTA 1640  
1641 CTTGAGAGTTAGCTCCATTGGTAACTCCACCATCCGTGTT 1680  
1681 ACCATCAACGGACGTGTTTACACAGTCTCTAATGTGAACA 1720  
1721 CTACAACGAACAATGATGGCGTTAACGACAACGGAGCCAG 1760  
1761 ATTCAGCGACATCAACATTGGCAACATCGTGGCCTCTGAC 1800  
20 1801 AACACTAACGTTACTTTGGACATCAATGTGACCCTCAATT 1840  
1841 CTGGAACCTCATTGATCTCATGAACATCATGTTTGTGCC 1880  
1881 AACTAACCTCCCTCCATTGTACTAA 1905.  
25

25. A plant transformation vector comprising a plant gene containing a structural gene of Claim 13.

30

-162-

26. A structural gene sequence of Claim 13  
encoding a fusion protein comprising the N-terminal  
610 amino acids of *B.t.k.* HD-1 and the C-terminal 567  
5 amino acids of *B.t.k.* HD-73, said gene having the  
sequence:

```
      1  ATGGACAACAACCCAAACATCAACGAATGCATTCCATACA    40
      41  ACTGCTTGAGTAACCCAGAAGTTGAAGTACTTGGTGGAGA    80
      81  ACGCATTGAAACCGGTTACACTCCCATCGACATCTCCTTG   120
     121  TCCTTGACACAGTTTCTGCTCAGCGAGTTCGTGCCAGGTG   160
     15 161  CTGGGTTTCGTTCTCGGACTAGTTGACATCATCTGGGGTAT   200
     201  CTTTGGTCCATCTCAATGGGATGCATTCTTGGTGCAAATT   240
     241  GAGCAGTTGATCAACCAGAGGATCGAAGAGTTCGCCAGGA   280
     281  ACCAGGCCATCTCTAGGTTGGAAGGATTGAGCAATCTCTA   320
     321  CCAAATCTATGCAGAGAGCTTCAGAGAGTGGGAAGCCGAT   360
     25 361  CCTACTAACCCAGCTCTCCGCGAGGAAATGCGTATTCAAT   400
     401  TCAACGACATGAACAGCGCCTTGACCACAGCTATCCCATT   440
```

30



-163-

441 GTTCGCAGTCCAGAACTACCAAGTTCCTCTCTTGTC CGTG 480  
5 481 TACGTTCAAGCAGCTAATCTTCACCTCAGCGTGCTTCGAG 520  
521 ACGTTAGCGTGTTTGGGCAAAGGTGGGGATTTCGATGCTGC 560  
561 AACCATCAATAGCCGTTACAACGACCTTACTAGGCTGATT 600  
10 601 GGAAACTACACCGACCACGCTGTTCTGTTGGTACAACACTG 640  
641 GCTTGGAGCGTGTCTGGGGTCCTGATTCTAGAGATTGGAT 680  
15 681 TAGATACAACCAGTTCAGGAGAGAATTGACCCTCACAGTT 720  
721 TTGGACATTGTGTCTCTCTTCCCGAACTATGACTCCAGAA 760  
761 CCTACCCTATCCGTACAGTGTCCCAACTTACCAGAGAAAT 800  
20 801 CTATACTAACCCAGTTCTTGAGAACTTCGACGGTAGCTTC 840  
841 CGTGGTTCTGCCCCAAGGTATCGAAGGCTCCATCAGGAGCC 880  
881 CACACTTGATGGACATCTTGAACAGCATAACTATCTACAC 920  
25 921 CGATGCTCACAGAGGAGAGTATTACTGGTCTGGACACCAG 960  
961 ATCATGGCCTCTCCAGTTGGATTTCAGCGGGCCCGAGTTTA 1000  
30 1001 CCTTTCCTCTCTATGGAAGTATGGGAAACGCCGCTCCACA 1040

-164-

	1041	ACAACGTATCGTTGCTCAACTAGGTCAGGGTGTCTACAGA	1080
5	1081	ACCTTGTCTTCCACCTTGTACAGAAGACCCTTCAATATCG	1120
	1121	GTATCAACAACCAGCAACTTCCGTTCTTGACGGAACAGA	1160
10	1161	GTTGCGCTATGGAACCTCTTCTAACTTGCCATCCGCTGTT	1200
	1201	TACAGAAAGAGCGGAACCGTTGATTCCCTTGGACGAAATCC	1240
	1241	CACCACAGAACAACAATGTGCCACCCAGGCAAGGATTCTC	1280
15	1281	CCACAGGTTGAGCCACGTGTCCATGTTCCGTTCCGGATTC	1320
	1321	AGCAACAGTTCCGTGAGCATCATCAGAGCTCCTATGTTCT	1360
	1361	CATGGATTCATCGTAGTGCTGAGTTCAACAATATCATTCC	1400
20	1401	TTCCTCTCAAATCACCCAAATCCCATTGACCAAGTCTACT	1440
	1441	AACCTTGATCTGGAACCTTCTGTCGTGAAAGGACCAGGCT	1480
25	1481	TCACAGGAGGTGATATTCTTAGAAGAACTTCTCCTGGCCA	1520
	1521	GATTAGCACCCCTCAGAGTTAACATCACTGCACCACTTTCT	1560
	1561	CAAAGATATCGTGTGAGGATTGTTACGCATCTACCACTA	1600
30	1601	ACTTGCAATTCCACACCTCCATCGACGGAAGGCCTATCAA	1640

-165-

	1641	TCAGGGTAACTTCTCCGCAACCATGTCAAGCGGCAGCAAC	1680
5	1681	TTGCAATCCGGCAGCTTCAGAACCGTCGGTTTCACTACTC	1720
	1721	CTTTCAACTTCTCTAACGGATCAAGCGTTTTCACCCCTTAG	1760
10	1761	CGCTCATGTGTTCAATTCTGGCAATGAAGTGACATTGAC	1800
	1801	CGTATTGAGTTTGTGCCTGCCGAAGTTACCCTCGAGGCTG	1840
	1841	AGTACAACCTTGAGAGAGCCCAGAAGGCTGTGAACGCCCT	1880
15	1881	CTTTACCTCCACCAATCAGCTTGGCTTGAAAATAACGTT	1920
	1921	ACTGACTATCACATTGACCAAGTGTCCTTGGTACCT	1960
	1961	ACCTTAGCGATGAGTTCTGCCTCGACGAGAAGCGTGAAC	2000
20	2001	CTCCGAGAAAGTTAAACACGCCAAGCGTCTCAGCGACGAG	2040
	2041	AGGAATCTCTTGCAAGACTCCAATTCAAAGACATCAACA	2080
25	2081	GGCAGCCAGAACGTGGTTGGGGTGGAAGCACCGGGATCAC	2120
	2121	CATCCAAGGAGGCGACGATGTGTTCAAGGAGAACTACGTC	2160
	2161	ACCCTCTCCGGAACTTTCGACGAGTGCTACCCTACCTACT	2200
30	2201	TGTACCAGAAGATCGATGAGTCCAAACTCAAAGCCTTCAC	2240

-166-

	2241	CAGGTATCAACTTAGAGGCTACATCGAAGACAGCCAAGAC	2280
5	2281	CTTGAAATCTACTCGATCAGGTACAATGCCAAGCACGAGA	2320
	2321	CCGTGAATGTCCCAGGTACTGGTTCCTCTGGCCACTTTC	2360
	2361	TGCCCAATCTCCCATTGGGAAGTGTGGAGAGCCTAACAGA	2400
10	2401	TGCGCTCCACACCTTGAGTGGAATCCTGACTTGGACTGCT	2440
	2441	CCTGCAGGGATGGCGAGAAGTGTGCCACCATCTCATCA	2480
15	2481	CTTCTCCTTGGACATCGATGTGGGATGTACTGACCTGAAT	2520
	2521	GAGGACCTCGGAGTCTGGGTCATCTTCAAGATCAAGACCC	2560
	2561	AAGACGGACACGCAAGACTTGGCAACCTTGAGTTTCTCGA	2600
20	2601	AGAGAAACCATTGGTCGGTGAAGCTCTCGCTCGTGTGAAG	2640
	2641	AGAGCAGAGAAGAAGTGGAGGGACAAACGTGAGAACTCG	2680
25	2681	AATGGGAAACTAACATCGTTTACAAGGAGGCCAAAGAGTC	2720
	2721	CGTGGATGCTTTGTTTCGTGAACTCCCAATATGATCAGTTG	2760
	2761	CAAGCCGACACCAACATCGCCATGATCCACGCCGCAGACA	2800
30	2801	AACGTGTGCACAGCATTCGTGAGGCTTACTTGCTTGAGTT	2840

-167-

	2841	GTCCGTGATCCCTGGTGTGAACGCTGCCATCTTCGAGGAA	2880
5	2881	CTTGAGGGACGTATCTTTACCGCATTCTCCTTGTACGATG	2920
	2921	CCAGAAACGTCATCAAGAACGGTGA CTTCACAATGGCCT	2960
	2961	CAGCTGCTGGAATGTGAAAGGTCATGTGGACGTGGAGGAA	3000
10	3001	CAGAACAAATCAGCGTTCCGTCCTGGTTGTGCCTGAGTGGG	3040
	3041	AAGCTGAAGTGTCCCAAGAGGTTAGAGTCTGTCCAGGTAG	3080
15	3081	AGGCTACATTCTCCGTGTGACCGCTTACAAGGAGGGATAC	3120
	3121	GGTGAGGGTTGCGTGACCATCCACGAGATCGAGAACAACA	3160
	3161	CCGACGAGCTTAAGTTCTCCA ACTGCGTCGAGGAAGAAAT	3200
20	3201	CTATCCCAACAACACCGT TACTTGCAACGACTACACTGTG	3240
	3241	AATCAGGAAGAGTACGGAGGTGCCTACACTAGCCGTAACA	3280
	3281	GAGGTTACAACGAAGCTCCTTCCGTTCTTGCTGACTATGC	3320
25	3321	CTCCGTGTACGAGGAGAAATCCTACACAGATGGCAGACGT	3360
	3361	GAGAACCCTTGCGAGTTCAACAGAGGTTACAGGGACTACA	3400
30	3401	CACCACTTCCAGTTGGCTATGTTACCAAGGAGCTTGAGTA	3440

-168-

3441 CTTTCCTGAGACCGACAAAGTGTGGATCGAGATCGGTGAA 3480  
5 3481 ACCGAGGGAACCTTCATCGTGGACAGCGTGGAGCTTCTCT 3520  
3521 TGATGGAGGAA 3531.

27. A method of Claim 4 further comprising removal  
10 of sequences comprising more than five consecutive A+T  
or G+C bases.

28. A structural gene sequence of Claim 13  
comprising a majority of plant preferred codons.

29. A structural gene encoding the coat protein of  
15 potato leaf roll virus, said gene having the sequence:

1 ATGAGTACTGTCGTGGTTAAGGGAAACGTGAACGGTGGTG 40  
41 TTCAACAACCTAGAAGGAGAAGAAGGCAATCCCTTCGTAG 80  
20 81 GAGAGCTAACAGAGTTCAGCCAGTGGTTATGGTCACTGCT 120  
121 CCTGGGCAACCAAGAAGGAGAAGAAGGAGAAGAGGAGGTA 160  
161 ATCGCAGATCAAGAAGAACTGGAGTTCCCAGAGGAAGAGG 200  
25 201 TTCAAGCGAGACATTCGTGTTTACAAAGGACAACCTCGTG 240  
241 GGCAACTCCCAAGGAAGTTTACCTTCGGACCAAGTGTTT 280  
30 281 CAGACTGTCCAGCATTCAAGGATGGAATACTCAAGGCTTA 320

-169-

321 CCATGAGTACAAGATCACAAGTATCTTGCTTCAGTTCGTC 360  
 5 361 AGCGAGGCCTCTTCCACCTCTCCAGGCTCCATCGCTTATG 400  
 401 AGTTAGATCCACATTGCAAAGTTTCATCCCTCCAGTCCTA 440  
 441 CGTCAACAAGTTCCAAATCACAAAGGGTGGTGCTAAGACC 480  
 10 481 TATCAAGCTCGTATGATCAACGGAGTTGAATGGCACGATT 520  
 521 CTTCTGAGGATCAGTGCAGAATCCTTTGGAAAGGAAATGG 560  
 15 561 AAAGTCTTCAGATCCAGCTGGATCTTTCAGAGTTACCATC 600  
 601 AGAGTTGCTCTTCAAAACCCAAAG 624.

20 30. A chimeric plant gene which comprises a structural coding sequence encoding an insecticidal protein of *Bacillus thuringiensis*, said structural coding sequence being modified to reduce the number of putative polyadenylation signals within said structural coding sequence.

25 31. A chimeric plant gene of Claim 30 in which the polyadenylation signals are selected from the group consisting of AATAAA, AATAAT, AACCAA, ATATAA, AATCAA, ATACTA, ATAAAA, ATGAAA, AAGCAT, ATTAAT, ATACAT, AAAATA, ATTAAA, AATTAA, AATACA and CATAAA.

30

-170-

5       32. A chimeric plant gene of Claim 31 in which said structural coding sequence is further modified to reduce the number of ATTTA sequences within said structural coding sequence.

      33. A chimeric plant gene of Claim 32 in which said structural coding sequence is substantially devoid of polyadenylation signals and ATTTA sequences.

10       34. A transformed plant cell containing a gene of Claim 33.

      35. A transformed plant cell of Claim 34 selected from the group consisting of soybean, cotton, alfalfa, oilseed rape, flax, tomato, sugarbeet, sunflower, potato, tobacco, maize, rice and wheat.

15       36. A plant comprising transformed plant cells of Claim 34.

      37. A plant of Claim 36 which comprises plant cells of Claim 35.

      38. A seed produced by a plant of Claim 36.

20

25

30



DETERMINATION OF DNA REGIONS IN GENES TO MODIFY BY SITE-DIRECTED  
MUTAGENESIS FOR INCREASED EXPRESSION IN PLANTS

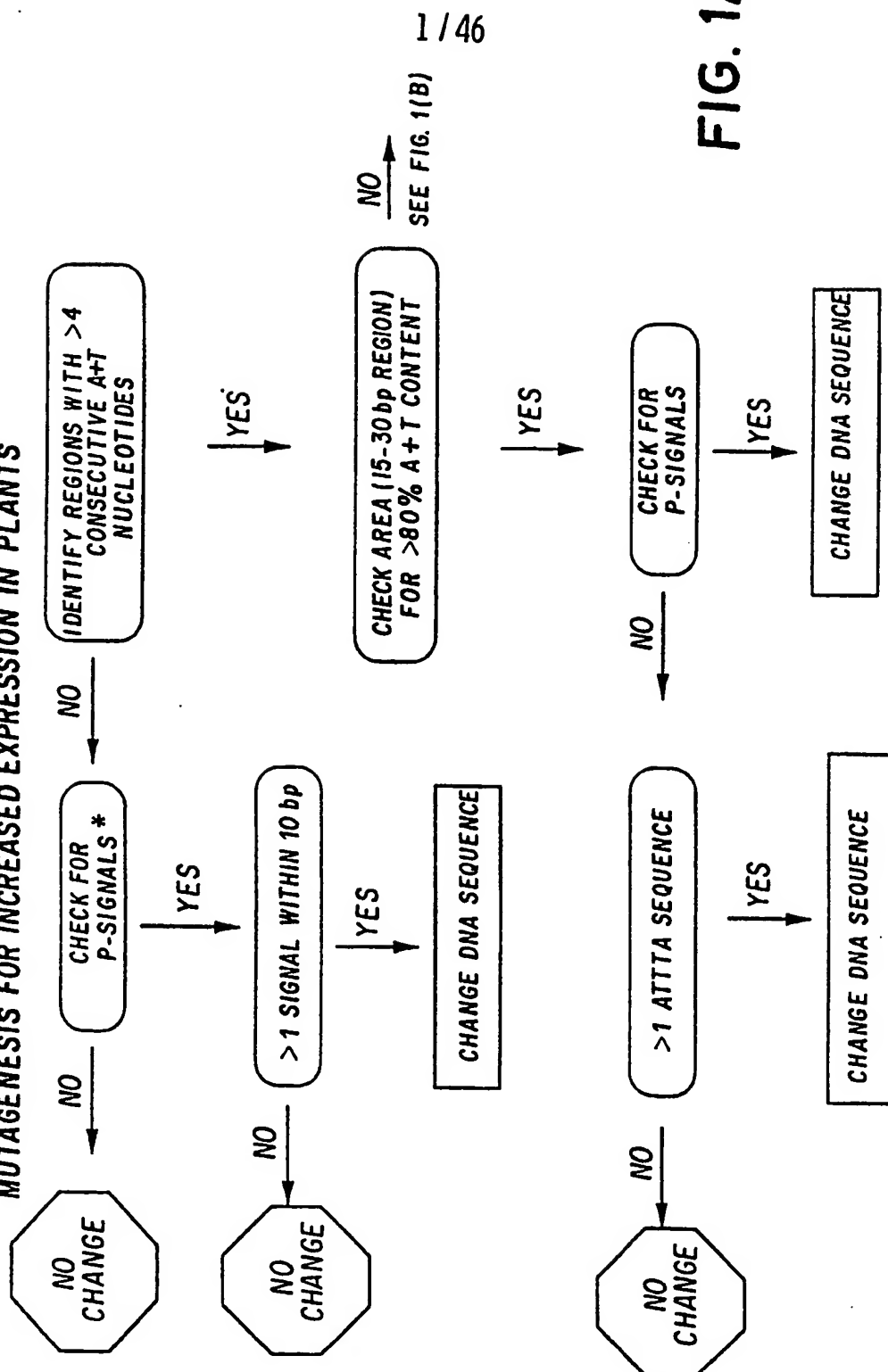


FIG. 1A

\* POLYADENYLATION SIGNAL SEQUENCES

DETERMINATION OF DNA REGIONS IN GENES TO MODIFY BY SITE-DIRECTED  
MUTAGENESIS FOR INCREASED EXPRESSION IN PLANTS

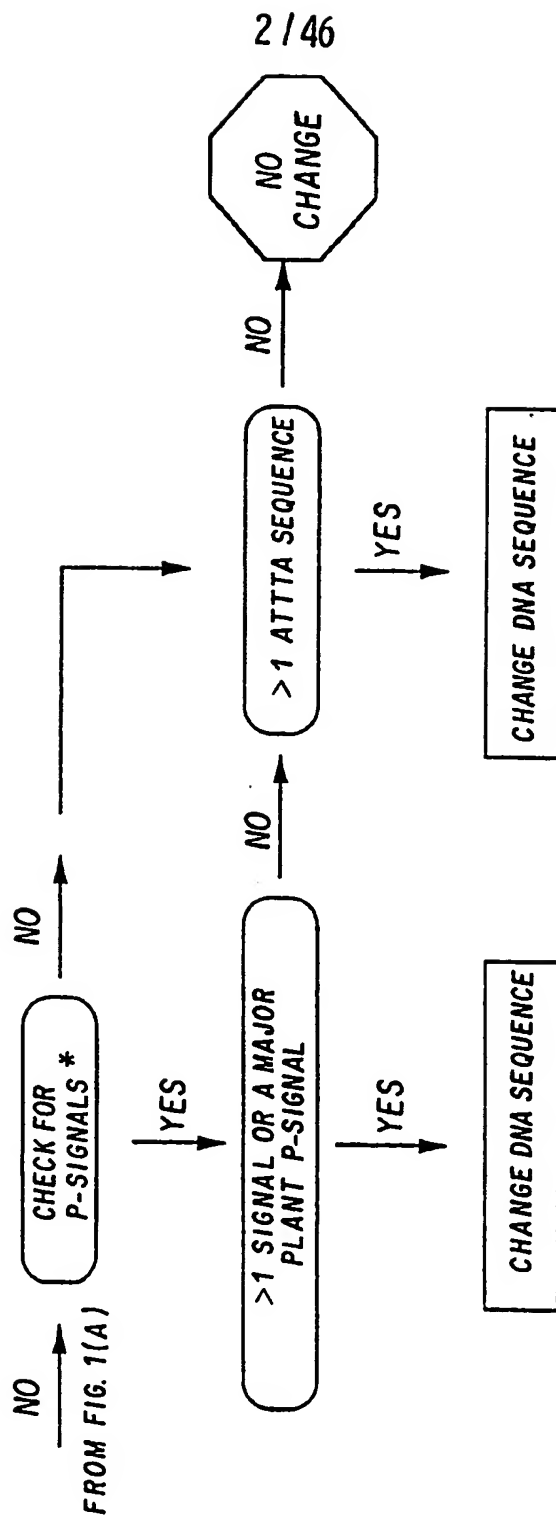


FIG. 1B

\* POLYADENYLATION SIGNAL SEQUENCES

3/46

1	ATGGCTATAGAACTGGTTACACCCCAATCGATATTCCT	40
41	TGTCGCTAACGCAATTTCTTTTGAGTGAATTTGTTCCCGG	80
81	TGCTGGATTTGTGTTAGGACTAGTTGATATAATATGGGGA T  C	120
121	ATTTTTGGTCCCTCTCAATGGGACGCATTTCTTGACAAA	160
161	TTGAACAGTTAATTAACCAAAGAATAGAAGAATTCGCTAG C  C  C          G          C          G	200
201	GAACCAAGCCATTTCTAGATTAGAAGGACTAAGCAATCTT T	240
241	TATCAAATTTACGCAGAATCTTTTAGAGAGTGGGAAGCAG	280
281	ATCCTACTAATCCAGCATTAAAGAGAAGAGATGCGTATTCA	320
321	ATTCAATGACATGAACAGTGCCCTTACAACCGCTATTCCT	360
361	CTTTTTGCAGTTCAAAATTATCAAGTTCCTCTTTTATCAG CC  C  C	400
401	TATATGTTCAAGCTGCAAATTTACATTTATCAGTTTGTAG G  C                  C  CC  C  CC  C	440
441	AGATGTTTCAGTGTTTGGACAAAGGTGGGGATTGATGCC	480
481	GCGACTATCAATAGTCGTTATAATGATTTAACTAGGCTTA	520
521	TTGGCAACTATACAGATCATGCTGTACGCTGGTACAATAC	560
561	GGGATTAGAGCGTGTATGGGGACCGGATTCTAGAGATTGG	600
601	ATAAGATATAATCAATTTAGAAGAGAATTAACACTAACTG C  G  C  C  G  C                  GC  T	640
641	TATTAGATATCGTTTCTCTATTTCCGAACATGATAGTAG	680
681	AACGTATCCAATTCGAACAGTTTCCCAATTAACAAGAGAA	720

FIG. 2A

SUBSTITUTE SHEET

4 / 46

721	ATTTATACAAACCCAGTATTAGAAAATTTTGATGGTAGTT	760
761	TTCGAGGCTCGGCTCAGGGCATAGAAGGAAGTATTAGGAG	800
801	TCCACATTTGATGGATATACTTAATAGTATAACCATCTAT	840
841	ACGGATGCTCATAGAGGAGAAATATTATTGGTCAGGGCATC C C C T C	880
881	AAATAATGGCTTCTCCTGTAGGGTTTTTCGGGGCCAGAATT G C	920
921	CACTTTTCCGCTATATGGAACCTATGGGAAATGCAGCTCCA	960
961	CAACAACGTATTGTTGCTCAACTAGGTCAGGGCGTGTATA	1000
1001	GAACATTATCGTCCACCTTATATAGAAGACCTTTTAATAT C	1040
1041	AGGGATAAATAATCAACAACCTATCTGTTCTTGACGGGACA C C C C	1080
1081	GAATTGCTTATGGAACCTCCTCAAATTTGCCATCCGCTG	1120
1121	TATACAGAAAAAGCGGAACGGTAGATTGCTGGATGAAAT	1160
1161	ACCGCCACAGAATAACAACGTGCCACCTAGGCAAGGATT	1200
1201	AGTCATCGATTAAGCCATGTTTCAATGTTTCGTTCAAGGCT	1240
1241	TTAGTAATAGTAGTGTAAGTATAATAAGAGCTCCTATGTT	1280
1281	CTCTTGGATACATCGTAGTGCTGAATTTAATAATATAATT G C C C C C	1320
1321	CCTTCATCACAAATTACACAAATACCTTTAACAAATCTA C C C AC C C G	1360
1361	CTAATCTTGGCTCTGGAACCTTCTGTCGTTAAAGGACCAGG	1400

FIG. 2B

SUBSTITUTE SHEET

5 / 46

1401	ATTTACAGGAGGAGATATTCTTCGAAGAACTTCACCTGGC	1440
1441	CAGATTTCAACCTTAAGAGTAAATATTACTGCACCATTAT	1480
1481	CACAAAGATATCGGGTAAGAATTCGCTACGCTTCTACCAC	1520
1521	AAATTTACAATTCATACATCAATTGACGGAAGACCTATT CC T G C	1560
1561	AATCAGGGGAATTTTTCAGCAACTATGAGTAGTGGGAGTA	1600
1601	ATTTACAGTCCGGAAGCTTTAGGACTGTAGGTTTTACTAC	1640
1641	TCCGTTTAACTTTTCAAATGGATCAAGTGTATTTACGTTA	1680
1681	AGTGCTCATGTCTTCAATTCAGGCAATGAAGTTTATATAG	1720
1721	ATCGAATTGAATTTGTTCCGGCA	1743

FIG. 2C

SUBSTITUTE SHEET

6 / 46

1	ATGGATAACAATCCGAACATCAATGAATGCATTCCTTATA	40
	C C A C A C	
41	ATTGTTTAAAGTAACCCTGAAGTAGAAGTATTAGGTGGAGA	80
	C C G A T C T	
81	AAGAATAGAACTGGTTACACCCCAATCGATATTTTCCTTG	120
	C C T C T C C C	
121	TCGCTAACGCAATTTCTTTGAGTGAATTTGTTCCCGGTG	160
	CT G A G GC C C G C G A	
161	CTGGATTGTGTTAGGACTAGTTGATATAATATGGGGAAT	200
	G C TC C C C C T	
201	TTTTGGTCCCTCTCAATGGGACGCATTTCTTGTACAAATT	240
	C A T C G G	
241	GAACAGTTAATTAACCAAAGAATAGAAGAATTCGCTAGGA	280
	G G C G G C G C	
281	ACCAAGCCATTTCTAGATTAGAAGGACTAAGCAATCTTTA	320
	G C G G T G C	
321	TCAAATTTACGCAGAATCTTTTAGAGAGTGGAAGCAGAT	360
	C C T GAGC C C	
361	CCTACTAATCCAGCATTAAAGAGAAGAGATGCGTATTCAAT	400
	C TC CC C G A	
401	TCAATGACATGAACAGTGCCCTTACAACCGCTATTCCTCT	440
	C C T G C A C AT	
441	TTTTGCAGTTCAAAATTATCAAGTTCCTCTTTTATCAGTA	480
	G C C G C C C G C G	
481	TATGTTCAAGCTGCAAATTTACATTTATCAGTTTTGAGAG	520
	C A T C T CC CAGC GC TC	
521	ATGTTTCAGTGTTTGGACAAAGGTGGGGATTTGATGCCGC	560
	C AGC G C T	
561	GACTATCAATAGTCGTTATAATGATTTAAGTAGGCTTATT	600
	A C C C C CC T G	
601	GGCAACTATACAGATcATGCTGTaCGCTGGTACAATACGG	640
	A C C CC C T T C T	
641	GATTAGAGCGTGTATGGGGACCGGATTCTAGAGATTGGAT	680
	C G C T T	

FIG. 3A

SUBSTITUTE SHEET

7 / 46

681	AAGATATAATCAATTTAGAAGAGAATTAACACTAACTGT	720
	T C C G C G G C C A T	
721	TTAGATATCGTTTCTCTATTTCCGAAGTATGATAGTAGAA	760
	G C T G C C CTCC	
761	CGTATCCAATTCGAACAGTTTCCCAATTAACAAGAGAAAT	800
	C C T C T G C T C	
801	TTATACAAACCCAGTATTAGAAAATTTTGATGGTAGTTTT	840
	C T TC T G C C C C C	
841	CGAGGCTCGGCTCAGGGCATAGAAGGAAGTATTAGGAGTC	880
	T T T C A T C CTCC C C	
881	CACATTTGATGGATATACTTAATAGTATAACCATCTATAC	920
	C C CT G C C T C	
921	GGATGCTCATAGAGGAGAATATTATTGGTCAGGGCATCAA	960
	C C G C T A C G	
961	ATAATGGCTTCTCCTGTAGGGTTTTTCGGGGCCAGAATTCA	1000
	C C A T A CAGC C G T	
1001	CTTTTCCGCTATATGGAAGTATGGGAAATGCAGCTCCACA	1040
	C T C C C	
1041	ACAACGTATTGTTGCTCAACTAGGTCAGGGCGTGTATAGA	1080
	C T C C	
1081	ACATTATCGTCCACCTTATATAGAAGACCTTTTAATATAG	1120
	C G T G C C C C	
1121	GGATAAATAATCAACAACCTATCTGTTCTTGACGGGACAGA	1160
	T C C C G T C A	
1161	ATTTGCTTATGGAACCTCCTCAAATTTGCCATCCGCTGT	1200
	G C C T T C T	
1201	TACAGAAAAAGCGGAACGGTAGATTCGCTGGATGAAATAC	1240
	G C T CT C C	
1241	CGCCACAGAATAACAACGTGCCACCTAGGCAAGGATTTAG	1280
	A C T C CTC	
1281	TCATCGATTAAAGCCATGTTTCAATGTTTCGTTTCAGGCTTT	1320
	C CA G G C G C C C A C	
1321	AGTAATAGTAGTGTAAGTATAATAAGAGCTCCTATGTTCT	1360
	C C TCC G C C C	
1361	CTTGGATACATCGTAGTGCTGAATTTAATAATATAATTCC	1400
	A T G C C C	

FIG. 3B

SUBSTITUTE SHEET

8 / 46

1401	TTCATCACAAATTACACAAATACCTTTAACAAAATCTACT	1440
	C T C C C A G C G	
1441	AATCTTGGCTCTGGAACCTTCTGTCGTTAAAGGACCAGGAT	1480
	C A G C	
1481	TTACAGGAGGAGATATTCTTCGAAGAACTTCACCTGGCCA	1520
	C T A T	
1521	GATTTCAACCTTAAGAGTAAATATTACTGCACCATTATCA	1560
	AGC C C T C C C T T	
1561	CAAAGATATCGGGTAAGAATTCGCTACGCTTCTACCACAA	1600
	T C G T A A	
1601	ATTTACAATTCATACATCAATTGACGGAAGACCTATTAA	1640
	C G C C C C G C	
1641	TCAGGGGAATTTTTCAGCAACTATGAGTAGTGGGAGTAAT	1680
	T C C C C TCA C C C C	
1681	TTACAGTCCGGAAGCTTTAGGACTGTAGGTTTTACTACTC	1720
	G A C C A C C C	
1721	CGTTTAACTTTTCAAATGGATCAAGTGTATTTACGTTAAG	1760
	T C C T C C T C C C T	
1761	TGCTCATGTCTTCAATTCAGGCAATGAAGTTTATATAGAT	1800
	C G T G C T C	
1801	CGAATTGAATTTGTTCCGGCAGAAGTAACCTTTGAGGCAG	1840
	T G G T C T C T	
1841	AATAT	1845
	G C	

FIG. 3C

SUBSTITUTE SHEET



9 / 46

1	ATGGATAACAATCCGAACATCAATGAATGCATTTCCTTATA	40
	C C A C A C	
41	ATTGTTTAAGTAACCCTGAAGTAGAAGTATTAGGTGGAGA	80
	C C G A T C T	
81	AAGAATAGAACTGGTTACACCCCAATCGATATTCCTTG	120
	C C T C T C C C	
121	TCGCTAACGCAATTTCTTTTGAGTGAATTTGTTCCCGGTG	160
	CT G A G GC C C G C G A	
161	CTGGATTTGTGTTAGGACTAGTTGATATAATATGGGGAAT	200
	G C TC C C C C T	
201	TTTTGGTCCCTCTCAATGGGACGCATTTCTTGTACAAATT	240
	C A T C G G	
241	GAACAGTTAATTAACCAAAGAATAGAAGAATTCGCTAGGA	280
	G G C G G C G C	
281	ACCAAGCCATTTCTAGATTAGAAGGACTAAGCAATCTTTA	320
	G C G G T G C	
321	TCAAATTTACGCAGAATCTTTTAGAGAGTGGAAGCAGAT	360
	C C T GAGC C C	
361	CCTACTAATCCAGCATTAAAGAGAAGAGATGCGTATTCAAT	400
	C TC CC C G A	
401	TCAATGACATGAACAGTGCCTTACAACCGCTATTCCTCT	440
	C C T G C A C AT	
441	TTTTGCAGTTCAAAATTATCAAGTTCCTCTTTTATCAGTA	480
	G C C G C C C G C G	
481	TATGTTCAAGCTGCAAATTTACATTTATCAGTTTTGAGAG	520
	C A T C T CC CAGC GC TC	
521	ATGTTTCAGTGTGTTGGACAAAGGTGGGGATTGATGCCGC	560
	C AGC G C T	
561	GACTATCAATAGTCGTTATAATGATTTAACTAGGCTTATT	600
	A C C C C CC T G	
601	GGCAACTATACAGATTATGCTGTACGCTGGTACAATACGG	640
	A C C CC C T T C T	
641	GATTAGAACGTGTATGGGGACCGGATTCTAGAGATTGGGT	680
	C G G C T T A	

FIG. 4A

SUBSTITUTE SHEET

10 / 46

681	AAGGTATAATCAATTTAGAAAGAGAATTAACACTAACTGTA	720
	T A C C G C G G C C A T	
721	TTAGATATCGTTGCTCTGTTCCCGAATTATGATAGTAGAA	760
	G C T GT C C CTCC	
761	GATATCCAATTTCGAACAGTTTCCCAATTAACAAGAGAAAT	800
	CC C T C T G C T C	
801	TTATACAAACCCAGTATTAGAAAATTTTGATGGTAGTTTT	840
	C T TC T G C C C C C	
841	CGAGGCTCGGCTCAGGGCATAGAAAGAAGTATTAGGAGTC	880
	T T T C A T C G CTCC C C	
881	CACATTTGATGGATATACTTAACAGTATAACCATCTATAC	920
	C C CT G C T C	
921	GGATGCTCATAGGGGTTATTATTATTGGTCAGGGCATCAA	960
	C C A AG G C T A C G	
961	ATAATGGCTTCTCCTGTAGGGTTTTCGGGGCCAGAATTCA	1000
	C C A T A CAGC C G T	
1001	CTTTTCCGCTATATGGAACATATGGGAAATGCAGCTCCACA	1040
	C T C C C	
1041	ACAACGTATTGTTGCTCAACTAGGTCAGGGCGTGTATAGA	1080
	C T C C	
1081	ACATTATCGTCCACTTTATATAGAAGACCTTTTAATATAG	1120
	C G T C G C C C C	
1121	GGATAAATAATCAACAACCTATCTGTTCTTGACGGGACAGA	1160
	T C C C G T C A	
1161	ATTTGCTTATGGAACCTCCTCAAATTTGCCATCCGCTGTA	1200
	G C C T T C T	
1201	TACAGAAAAAGCGGAACGGTAGATTGCTGGATGAAATAC	1240
	G C T CT C C	
1241	CGCCACAGAATAACAACGTGCCACCTAGGCAAGGATTTAG	1280
	A C T C CTC	
1281	TCATCGATTAAAGCCATGTTTCAATGTTTCGTTTCAGGCTTT	1320
	C CA G G C G C C C A C	
1321	AGTAATAGTAGTGTAAAGTATAATAAGAGCTCCTATGTTCT	1360
	C C TCC G C C C	
1361	CTTGATACATCGTAGTGCTGAATTTAATAATATAATTGC	1400
	C G C C C C C	

FIG. 4B

SUBSTITUTE SHEET

11/46

1401	ATCGGATAGTATTACTCAAATCCCTGCAGTGAAGGGAAAC	1440
	C	
1441	TTTCTTTTAAATGGTTCTGTAATTCAGGACCAGGATTTA	1480
	C C C C C	
1481	CTGGTGGGGACTTAGTTAGATTAAATAGTAGTGGAATAA	1520
	A C C C C C C	
1521	CATTCAGAATAGAGGGTATATTGAAGTTCCAATTCCTTC	1560
1561	CCATCGACATCTACCAGATATCGAGTTCGTGTACGGTATG	1600
	C A GA	
1601	CTTCTGTAACCCCGATTACCTCAACGTTAATTGGGGTAA	1640
	G T	
1641	TTCATCCATTTTTCCTCAATACAGTACCAGCTACAGCTACG	1680
	C C T C	
1681	TCATTAGATAATCTACAATCAAGTGATTTTGGTTATTTTG	1720
	C G C C C C C	
1721	AAAGTGCCAATGCTTTTACATCTTCATTAGGTAATATAGT	1760
	C C C C	
1761	AGGTGTTAGAAATTTTAGTGGGACTGCAGGAGTGATAATA	1800
	G C T C	
1801	GACAGATTTGAATTTATTCCAGTTACTGCAACACTCGAGG	1840
	C G C	
1841	CTGAATATAATCTGGAAAGAGCGCAGAAGGCGGTGAATGC	1880
	A TGCG	
1881	GCTGTTTACGTCTACAAACCAACTAGGGCTAAAAACAAAT	1920
	CTGT ACGTCTACA C AGCT G ACTC G CA TG	
1921	G 1921	

FIG. 4C

SUBSTITUTE SHEET

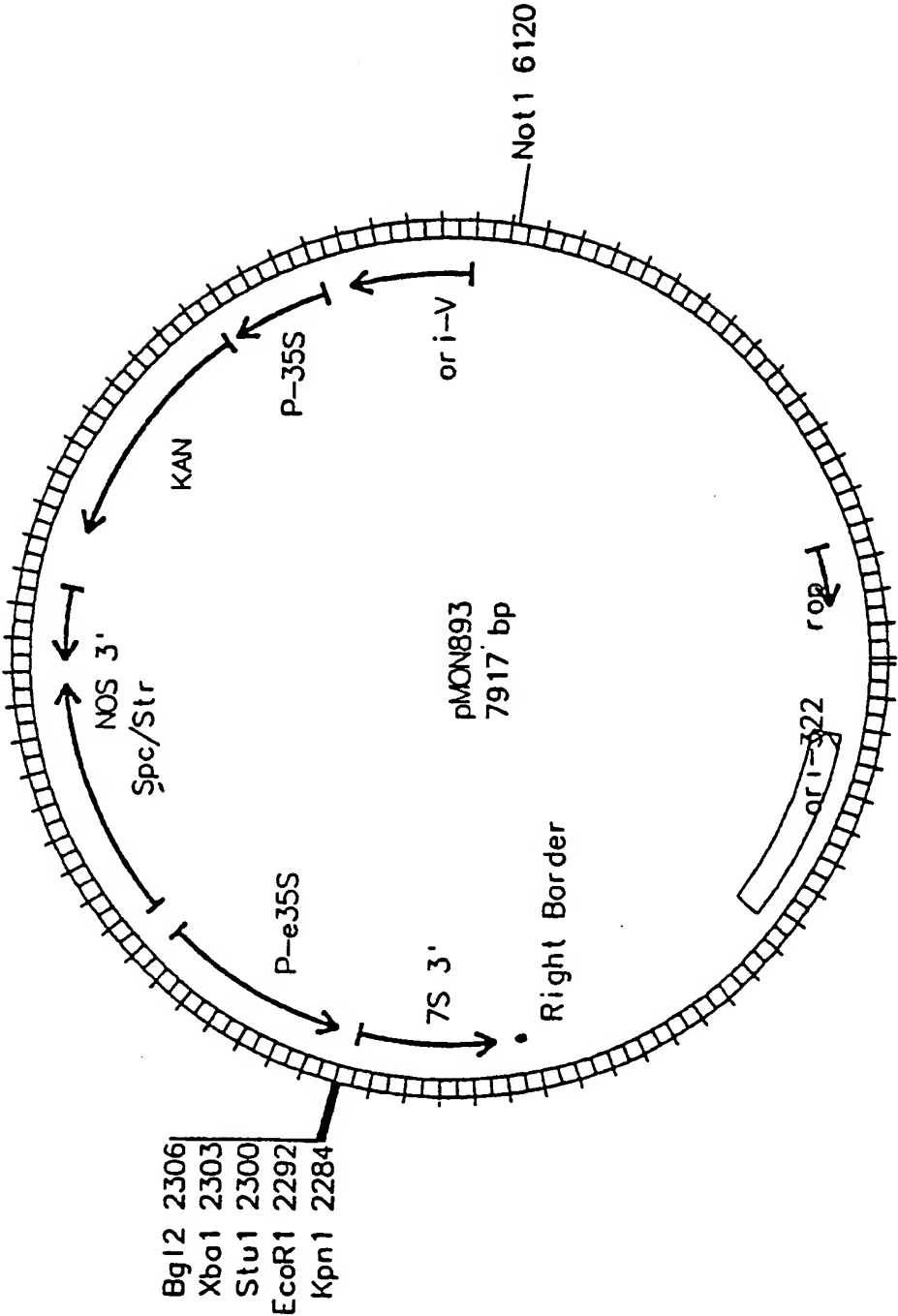
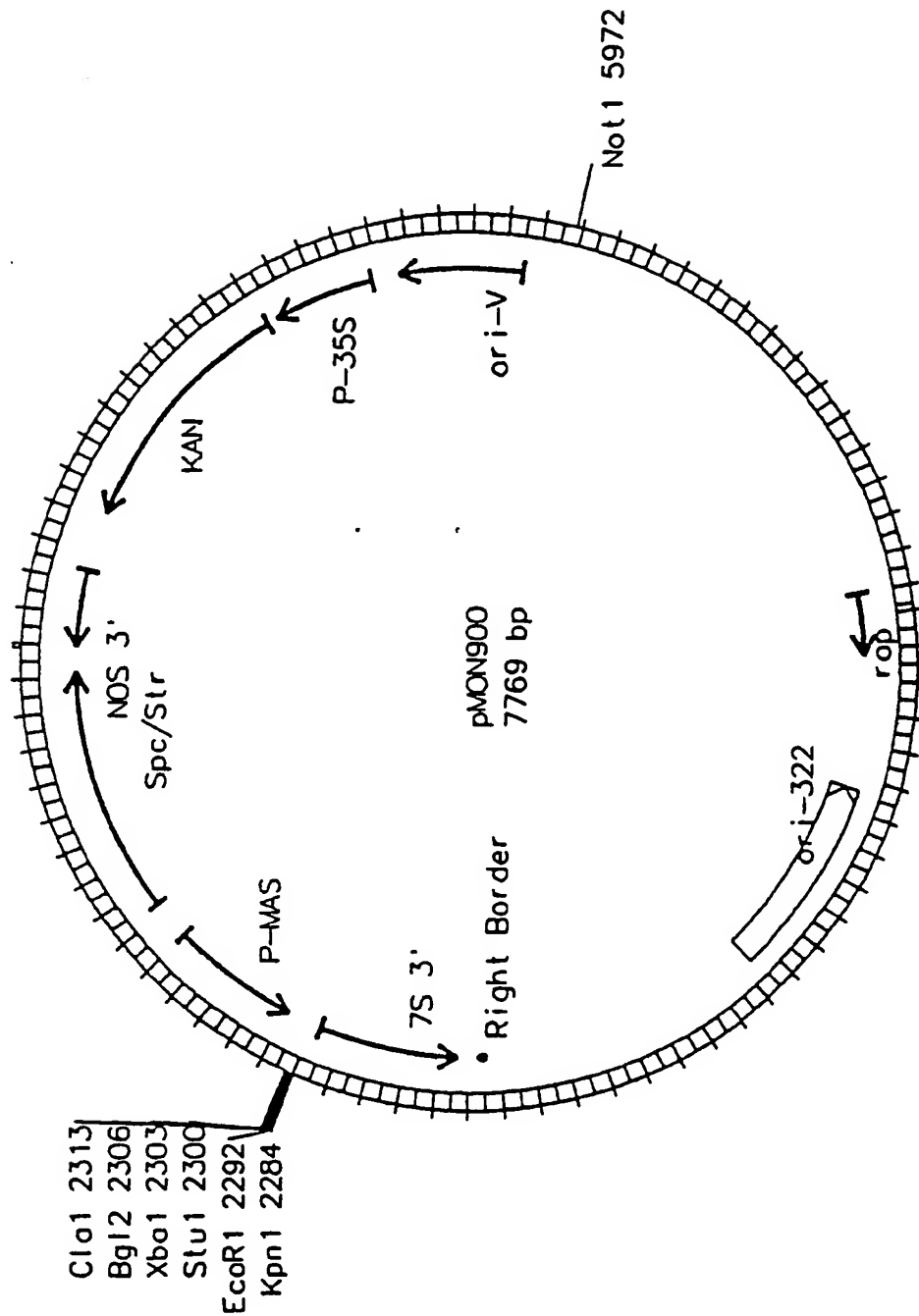


FIG. 5



**FIG. 6**

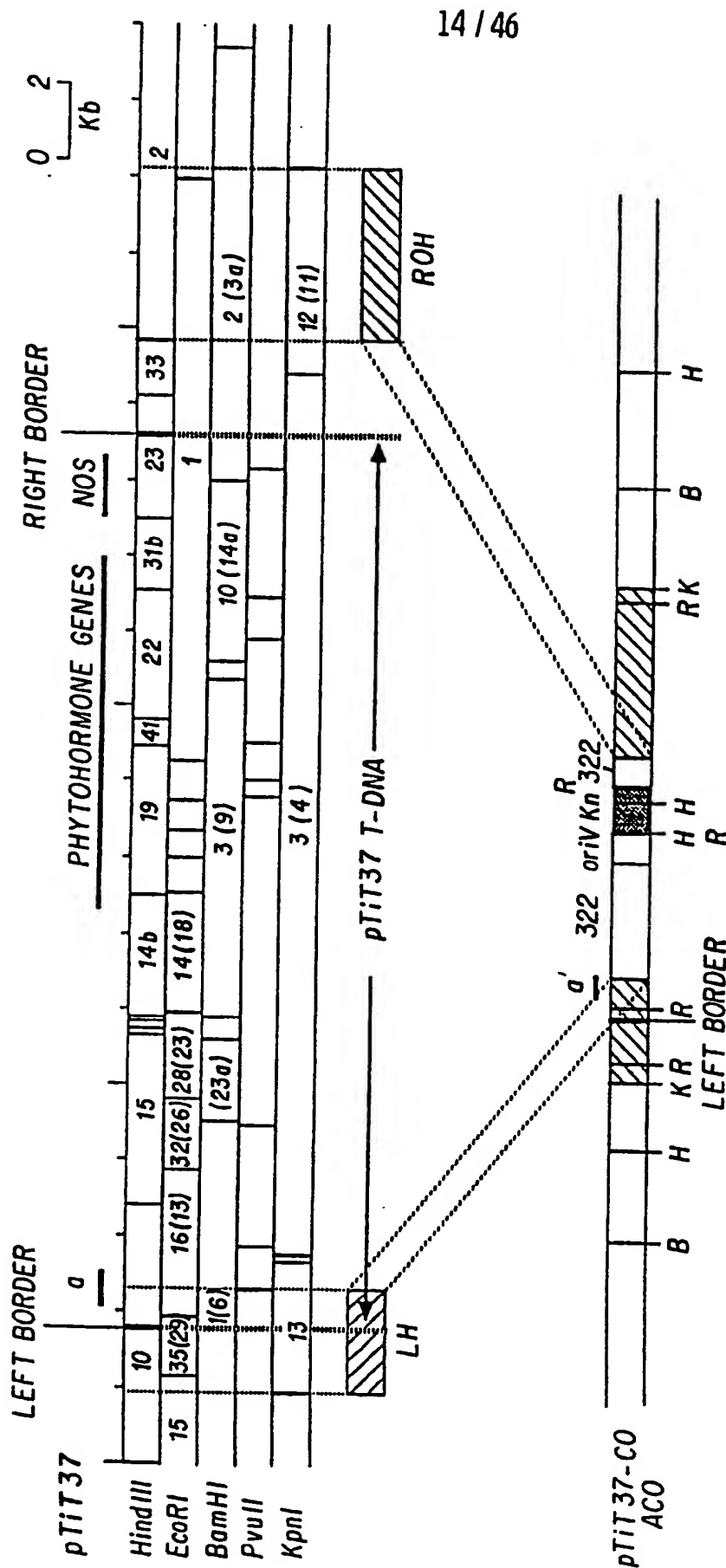


FIG. 7

15 / 46

1	GAAAGAATAGAACTGGTTACACCCCAATCGATATTTTCCT	40
	ATGGCC T C T C C C	
41	TGTCGCTAACGCAATTTCTTTTGAGTGAATTTGTTCCCGG	80
	CT G A G GC C C G C G A	
81	TGCTGGATTTGTGTTAGGACTAGTTGATATAATATGGGGA	120
	G C TC C C C C T	
121	ATTTTTGGTCCCTCTCAATGGGACGCATTTCTTGACAAA	160
	C A T C G G	
161	TTGAACAGTTAATTAACCAAAGAATAGAAGAATTCGCTAG	200
	G G C G G C G C	
201	GAACCAAGCCATTTCTAGATTAGAAGGACTAAGCAATCTT	240
	G C G G T G C	
241	TATCAAATTTACGCAGAATCTTTTAGAGAGTGGGAAGCAG	280
	C C T GAGC C C	
281	ATCCTACTAATCCAGCATTAAAGAGAAGAGATGCGTATTCA	320
	C TC CC C G A	
321	ATTCAATGACATGAACAGTGCCCTTACAACCGCTATTCCT	360
	C C T G C A C A	
361	CTTTTTGCAGTTCAAATTATCAAGTTCCTCTTTTATCAG	400
	T G C C G C C C G C	
401	TATATGTTCAAGCTGCAAATTTACATTTATCAGTTTTGAG	440
	G C A T C T CC CAGC GC TC	
441	AGATGTTTCAGTGTTTGGACAAAGGTGGGGATTTGATGCC	480
	C AGC G C T	
481	GCGACTATCAATAGTCGTTATAATGATTTAACTAGGCTTA	520
	A C C C C CC T G	
521	TTGGCAACTATACAGATTATGCTGTACGCTGGTACAATAC	560
	A C C CC C T T C	
561	GGGATTAGAACGTGTATGGGGACCGGATTCTAGAGATTGG	600
	T C G G C T T	
601	GTAAGGTATAATCAATTTAGAAGAGAATTAACTAACTG	640
	A T A C C G C G G C C A	
641	TATTAGATATCGTTGCTCTGTTCCCGAATTATGATAGTAG	680
	T G C T GT C C CTCC	

FIG.8A

SUBSTITUTE SHEET

16 / 46

681	AAGATATCCAATTTCGAACAGTTTCCCAATTAAACAAGAGAA	720
	CC C T C T G C T C	
721	ATTTATACAAACCCAGTATTAGAAAATTTTGATGGTAGTT	760
	C T TC T G C C C C	
761	TTCGAGGCTCGGCTCAGGGCATAGAAAGAAGTATTAGGAG	800
	C T T T C A T C G CTCC C	
801	TCCACATTTGATGGATATACTTAACAGTATAACCATCTAT	840
	C C C CT G C T C	
841	ACGGATGCTCATAGGGGTTATTATTATTGGTCAGGGCATC	880
	C C A AG G C T A C	
881	AAATAATGGCTTCTCCTGTAGGGTTTTTCGGGGCCAGAATT	920
	G C C A T A CAGC C G	
921	CACTTTTCCGCTATATGGAACCTATGGGAAATGCAGCTCCA	960
	T C T C C C	
961	CAACAACGTATTGTTGCTCAACTAGGTCAGGGCGTGTATA	1000
	C T C C	
1001	GAACATTATCGTCCACTTTATATAGAAGACCTTTTAATAT	1040
	C G T C G C C C	
1041	AGGGATAAATAATCAACAACCTATCTGTTCTTGACGGGACA	1080
	C T C C C G T C A	
1081	GAATTTGCTTATGGAACCTCCTCAAATTTGCCATCCGCTG	1120
	G C C T T C	
1121	TATACAGAAAAAGCGGAACGGTAGATTTCGCTGGATGAAAT	1160
	T G C T CT C	
1161	ACCGCCACAGAATAACAACGTGCCACCTAGGCAAGGATTT	1200
	C A C T C C	
1201	AGTCATCGATTAAGCCATGTTTCAATGTTTCGTTTCAGGCT	1240
	TCC CA G G C G C C C A	
1241	TTAGTAATAGTAGTGTAAGTATAATAAGAGCTCCTATGTT	1280
	C C C TCC G C C C	
1281	CTCTTGGATACATCGTAGTGCTGAATTTAATAATATAATT	1320
	C G C C C C C	
1321	GCATCGGATAGTATTACTCAAATCCCTGCAGTGAAGGGAA	1360
	C	
1361	ACTTTCTTTTAAATGGTTCTGTAATTTTCAGGACCAGGATT	1400
	C C C C	

FIG. 8B

SUBSTITUTE SHEET



17 / 46

1401	TACTGGTGGGGACTTAGTTAGATTAAATAGTAGTGAAAT	1440
	C            A    C C            C C C C	
1441	AACATTCAGAATAGAGGGTATATTGAAGTTCCAATTCAC	1480
1481	TCCCATCGACATCTACCAGATATCGAGTTCGTGTACGGTA	1520
	C                            A                    GA	
1521	TGCTTCTGTAAACCCCGATTACCTCAACGTTAATTGGGGT	1560
	G            T	
1561	AATTCATCCATTTTTTCCAATACAGTACCAGCTACAGCTA	1600
	C C                            T	
1601	CGTCATTAGATAATCTACAATCAAGTGATTTTGGTTATTT	1640
	C C G                    C    C C            C C	
1641	TGAAAGTGCCAATGCTTTTACATCTTCATTAGGTAATATA	1680
	C C            C C	
1681	GTAGGTGTTAGAAATTTTAGTGGGACTGCAGGAGTGATAA	1720
	G                    C                            T	
1721	TAGACAGATTGAATTTATTCCAGTTACTGCAACACTCGA	1760
	C                    C G C	
1761	GGCTGAA	1767
	G	

FIG. 8C

SUBSTITUTE SHEET

18 / 46

1	ATGGATAACAATCCGAACATCAATGAATGCATTTCCTTATA	40
	C C A C A C	
41	ATTGTTTAAGTAACCCTGAAGTAGAAGTATTAGGTGGAGA	80
	C C G A T C T	
81	AAGAATAGAACTGGTTACACCCCAATCGATATTTTCCTTG	120
	C C T C T C C C	
121	TCGCTAACGCAATTTCTTTTGAGTGAATTTGTTCCCGGTG	160
	CT G A G GC C C G C G A	
161	CTGGATTGTGTAGGACTAGTTGATATAATATGGGGAAT	200
	G C TC C C C C T	
201	TTTTGGTCCCTCTCAATGGGACGCATTTCTTGTACAAATT	240
	C A T C G G	
241	GAACAGTTAATTAACCAAAGAATAGAAGAATTCGCTAGGA	280
	G G C G G C G C	
281	ACCAAGCCATTTCTAGATTAGAAGGACTAAGCAATCTTTA	320
	G C G G T G C	
321	TCAAATTTACGCAGAATCTTTTAGAGAGTGGAAGCAGAT	360
	C C T GAGC C C	
361	CCTACTAATCCAGCATTAAAGAGAAGAGATGCGTATTCAAT	400
	C TC CC C G A	
401	TCAATGACATGAACAGTGCCCTTACAACCGCTATTCCTCT	440
	C C T G C A C AT	
441	TTTTGCAGTTCAAAATTATCAAGTTCCTCTTTTATCAGTA	480
	G C C G C C C G C G	
481	TATGTTCAAGCTGCAAATTTACATTTATCAGTTTTGAGAG	520
	C A T C T CC CAGC GC TC	
521	ATGTTTCAGTGTTTGGACAAAGGTGGGGATTGATGCCGC	560
	C AGC G C T	
561	GACTATCAATAGTCGTTATAATGATTAACTAGGCTTATT	600
	A C C C C CC T G	
601	GGCAACTATACAGATTATGCTGTACGCTGGTACAATACGG	640
	A C C CC C T T C T	
641	GATTAGAACGTGTATGGGGACCGGATTCTAGAGATTGGGT	680
	C G G C T T A	

FIG. 9A

SUBSTITUTE SHEET

19 / 46

681	AAGGTATAATCAATTTAGAAGAGAATTAACACTAACTGTA	720
	T A C C G C G G C C A T	
721	TTAGATATCGTTGCTCTGTTCCCGAATTATGATAGTAGAA	760
	G C T GT C C CTCC	
761	GATATCCAATTCGAACAGTTTCCCAATTAACAAGAGAAAT	800
	CC C T C T G C T C	
801	TTATACAAACCCAGTATTAGAAAATTTTGATGGTAGTTTT	840
	C T TC T G C C C C C	
841	CGAGGCTCGGCTCAGGGCATAGAAAGAAGTATTAGGAGTC	880
	T T T C A T C G CTCC C C	
881	CACATTTGATGGATATACTTAACAGTATAACCATCTATAC	920
	C C CT G C T C	
921	GGATGCTCATAGGGGTTATTATTATTGGTCAGGGCATCAA	960
	C C A AG G C T A C G	
961	ATAATGGCTTCTCCTGTAGGGTTTTTCGGGGCCAGAATTCA	1000
	C C A T A CAGC C G T	
1001	CTTTTCCGCTATATGGAACATATGGGAAATGCAGCTCCACA	1040
	C T C C C	
1041	ACAACGTATTGTTGCTCAACTAGGTCAGGGCGTGTATAGA	1080
	C T C C	
1081	ACATTATCGTCCACTTTATATAGAAGACCTTTTAATATAG	1120
	C G T C G C C C C	
1121	GGATAAATAATCAACAACCTATCTGTTCTTGACGGGACAGA	1160
	T C C C G T C A	
1161	ATTTGCTTATGGAACCTCCTCAAATTTGCCATCCGCTGTA	1200
	G C C T T C T	
1201	TACAGAAAAAGCGGAACGGTAGATTCGCTGGATGAAATAC	1240
	G C T CT C C	
1241	CGCCACAGAATAACAACGTGCCACCTAGGCAAGGATTTAG	1280
	A C T C CTC	
1281	TCATCGATTAAAGCCATGTTTCAATGTTTCGTTTCAGGCTTT	1320
	C CA G G C G C C C A C	
1321	AGTAATAGTAGTGTAAGTATAATAAGAGCTCCTATGTTCT	1360
	C C TCC G C C C	
1361	CTTGGATACATCGTAGTGCTGAATTTAATAATATAATTGC	1400
	C G C C C C C	

FIG. 9B

SUBSTITUTE SHEET

20 / 46

1401	ATCGGATAGTATTACTCAAATCCCTGCAGTGAAGGGAAAC	1440
	C	
1441	TTTCTTTTTAATGGTTCTGTAATTTTCAGGACCAGGATTTA	1480
	C C C C C	
1481	CTGGTGGGGACTTAGTTAGATTAAATAGTAGTGGAATAA	1520
	A C C C C C C	
1521	CATTCAGAATAGAGGGTATATTGAAGTTCCAATTCCTTC	1560
1561	CCATCGACATCTACCAGATATCGAGTTCGTGTACGGTATG	1600
	C A GA	
1601	CTTCTGTAACCCCGATTACCTCAACGTTAATTGGGGTAA	1640
	G T	
1641	TTCATCCATTTTTTCCAATACAGTACCAGCTACAGCTACG	1680
	C C T C	
1681	TCATTAGATAATCTACAATCAAGTGATTTTGGTTATTTTG	1720
	C G C C C C C	
1721	AAAGTGCCAATGCTTTTACATCTTCATTAGGTAATATAGT	1760
	C C C C	
1761	AGGTGTTAGAAATTTTAGTGGGACTGCAGGAGTGATAATA	1800
	G C T C	
1801	GACAGATTTGAATTTATTCCAGTTACTGCAACACTCGAGG	1840
	C G C	
1841	CTGAATATAATCTGGAAAGAGCGCAGAAGGCGGTGAATGC	1880
1881	GCTGTTTACGTCTACAAACCAACTAGGGCTAAAAACAAAT	1920
1921	GTAACGGATTATCATATTGATCAAGTGTCCAATTTAGTTA	1960
1961	CGTATTTATCGGATGAATTTTGTCTGGATGAAAAGCGAGA	2000
2001	ATTGTCCGAGAAAGTCAAACATGCGAAGCGACTCAGTGAT	2040
2041	GAACGCAATTTACTCCAAGATTCAAATTTCAAAGACATTA	2080
2081	ATAGGCAACCAGAACGTGGGTGGGGCGGAAGTACAGGGAT	2120

FIG.9C

SUBSTITUTE SHEET

21 / 46

2121	TACCATCCAAGGAGGGGATGACGTATTTAAAGAAAATTAC	2160
2161	GTCACACTATCAGGTACCTTTGATGAGTGCTATCCAACAT	2200
2201	ATTTGTATCAAAAAATCGATGAATCAAAATTAAAGCCTT	2240
2241	TACCCGTTATCAATTAAGAGGGTATATCGAAGATAGTCAA	2280
2281	GACTTAGAAATCTATTTAATTCGCTACAATGCAAACATG	2320
2321	AAACAGTAAATGTGCCAGGTACGGGTTCTTATGGCCGCT	2360
2361	TTCAGCCCAAAGTCCAATCGGAAAGTGTGGAGAGCCGAAT	2400
2401	CGATGCGCGCCACACCTTGAATGGAATCCTGACTTAGATT	2440
2441	GTTCTGTAGGGATGGAGAAAAGTGTGCCCATCATTCGCA	2480
2481	TCATTTCTCCTTAGACATTGATGTAGGATGTACAGACTTA	2520
2521	AATGAGGACCTAGGTGTATGGGTGATCTTTAAGATTAAGA	2560
2561	CGCAAGATGGGCACGCAAGACTAGGGAATCTAGAGTTTCT	2600
2601	CGAAGAGAAACCATTAGTAGGAGAAGCGCTAGCTCGTGTG	2640
2641	AAAAGAGCGGAGAAAAAATGGAGAGACAAACGTGAAAAAT	2680
2681	TGGAATGGGAAACAAATATCGTTTATAAAGAGGCAAAGA	2720
2721	ATCTGTAGATGCTTTATTTGTAAACTCTCAATATGATCAA	2760
2761	TTACAAGCGGATACGAATATTGCCATGATTCATGCGGCAG	2800
2801	ATAAACGTGTTTCATAGCATTCGAGAAGCTTATCTGCCTGA	2840

FIG. 9D

SUBSTITUTE SHEET

22/46

2841	GCTGTCTGTGATTCCGGGTGTCAATGCGGCTATTTTGTAA	2880
2881	GAATTAGAAGGGCGTATTTTCACTGCATTCTCCCTATATG	2920
2921	ATGCGAGAAATGTCATTAAAAATGGTGATTTTAATAATGG	2960
2961	CTTATCCTGCTGGAACGTGAAAGGGCATGTAGATGTAGAA	3000
3001	GAACAAAACAACCAACGTTCCGGTCCTTGTTGTTCCGGAAT	3040
3041	GGGAAGCAGAAGTGTCAACAAGAAGTTCGTGTCTGTCCGGG	3080
3081	TCGTGGCTATATCCTTCGTGTCACAGCGTACAAGGAGGGA	3120
3121	TATGGAGAAGGTTGCGTAACCATTCATGAGATCGAGAACAA	3160
3161	ATACAGACGAACTGAAGTTTAGCAACTGCGTAGAAGAGGA	3200
3201	AATCTATCCAAATAACACGGTAACGTGTAATGATTATACT	3240
3241	GTAAATCAAGAAGAATACGGAGGTGCGTACACTTCTCGTA	3280
3281	ATCGAGGATATAACGAAGCTCCTTCCGTACCAGCTGATTA	3320
3321	TGCGTCAGTCTATGAAGAAAAATCGTATACAGATGGACGA	3360
3361	AGAGAGAATCCTTGTGAATTTAACAGAGGGTATAGGGATT	3400
3401	ACACGCCACTACCAGTTGGTTATGTGACAAAAGAATTAGA	3440
3441	ATACTTCCCAGAAACCGATAAGGTATGGATTGAGATTGGA	3480
3481	GAAACGGAAGGAACATTTATCGTGGACAGCGTGGAATTAC	3520
3521	TCCTTATGGAGGAA	3534

FIG. 9E

SUBSTITUTE SHEET

23 / 46

1	ATGGATAACAATCCGAACATCAATGAATGCATTTCCTTATA	40
	C C A C A C	
41	ATTGTTTAAGTAACCCCTGAAGTAGAAGTATTAGGTGGAGA	80
	C C G A T C T	
81	AAGAATAGAACTGGTTACACCCCAATCGATATTCCTTG	120
	C C T C T C C C	
121	TCGCTAACGCAATTTCTTTTGAGTGAATTTGTTCCCGGTG	160
	CT G A G GC C C G C G A	
161	CTGGATTGTGTAGGACTAGTTGATATAATATGGGGAAT	200
	G C TC C C C C T	
201	TTTTGGTCCCTCTCAATGGGACGCATTTCTTGTAACAATT	240
	C A T C G G	
241	GAACAGTTAATTAACCAAGAATAGAAGAATTCGCTAGGA	280
	G G C G G C G C	
281	ACCAAGCCATTTCTAGATTAGAAGGACTAAGCAATCTTTA	320
	G C G G T G C	
321	TCAAATTTACGCAGAATCTTTTAGAGAGTGGGAAGCAGAT	360
	C C T GAGC C C	
361	CCTACTAATCCAGCATTAAGAGAAGAGATGCGTATTCAAT	400
	C TC CC C G A	
401	TCAATGACATGAACAGTGCCCTTACAACCGCTATTCCTCT	440
	C C T G C A C AT	
441	TTTTGCAGTTCAAAATTATCAAGTTCCTCTTTTATCAGTA	480
	G C C G C C C G C G	
481	TATGTTCAAGCTGCAAATTTACATTTATCAGTTTGGAGAG	520
	C A T C T CC CAGC GC TC	
521	ATGTTTCAGTGTGGACAAAGGTGGGGATTTGATGCCGC	560
	C AGC G C T	
561	GACTATCAATAGTCGTTATAATGATTTAACTAGGCTTATT	600
	A C C C C CC T G	
601	GGCAACTATACAGATTATGCTGTACGCTGGTACAATACGG	640
	A C C CC C T T C T	

FIG. 10A

SUBSTITUTE SHEET

24 / 46

641	GATTAGAACGTGTATGGGGACCGGATTCTAGAGATTGGGT	680
	C G G C T T A	
681	AAGGTATAATCAATTTAGAAGAGAATTAACACTAACTGTA	720
	T A C C G C G G C C A T	
721	TTAGATATCGTTGCTCTGTTCCCGAATTATGATAGTAGAA	760
	G C T GT C C CTCC	
761	GATATCCAATTCTGAACAGTTTCCCAATTAACAAGAGAAAT	800
	CC C T C T G C T C	
801	TTATACAAACCCAGTATTAGAAAATTTTGATGGTAGTTTT	840
	C T TC T G C C C C C	
841	CGAGGCTCGGCTCAGGGCATAGAAAGAAGTATTAGGAGTC	880
	T T T C A T C G CTCC C C	
881	CACATTTGATGGATATACTTAACAGTATAACCATCTATAC	920
	C C CT G C T C	
921	GGATGCTCATAGGGGTTATTATTATTGGTCAGGGCATCAA	960
	C C A AG G C T A C G	
961	ATAATGGCTTCTCCTGTAGGGTTTTTCGGGGCCAGAATTCA	1000
	C C A T A CAGC C G T	
1001	CTTTTCCGCTATATGGAACCTATGGGAAATGCAGCTCCACA	1040
	C T C C C	
1041	ACAACGTATTGTTGCTCAACTAGGTCAGGGCGTGTATAGA	1080
	C T C C	
1081	ACATTATCGTCCACTTTATATAGAAGACCTTTTAATATAG	1120
	C G T C G C C C C	
1121	GGATAAATAATCAACAACCTATCTGTTCTTGACGGGACAGA	1160
	T C C C G T C A	
1161	ATTTGCTTATGGAACCTCCTCAAATTTGCCATCCGCTGTA	1200
	G C C T T C T	
1201	TACAGAAAAAGCGGAACGGTAGATTTCGCTGGATGAAATAC	1240
	G C T CT C C	
1241	CGCCACAGAATAACAACGTGCCACCTAGGCAAGGATTTAG	1280
	A C T C CTC	
1281	TCATCGATTAAAGCCATGTTTCAATGTTTCGTTCAAGGCTTT	1320
	C CA G G C G C C C A C	
1321	AGTAATAGTAGTGTAAGTATAATAAGAGCTCCTATGTTCT	1360
	C C TCC G C C C	

FIG. 10B

SUBSTITUTE SHEET



25 / 46

1361	CTTGGATACATCGTAGTGC <sup>•</sup> TGAATT <sup>•</sup> TAATAATATAATTGC <sup>•</sup>	1400
	C                  G  C  C  C  C  C	
1401	ATCGGATAGTATTACTCAAATCCCTGCAGTGAAGGGAAAC <sup>•</sup>	1440
	C	
1441	TTTCTTTT <sup>•</sup> TAATGGTTCTGTAAT <sup>•</sup> TCAGGACCAGGATT <sup>•</sup> TA	1480
	C  C  C                  C                  C	
1481	CTGGTGGGGACTTAGTTAGATTAAATAGTAGTGGAAATAA <sup>•</sup>	1520
	A  C  C                  C  C  C  C	
1521	CATTCAGAA <sup>•</sup> TAGAGGGTATATTGAAGTTCCAATTCACTTC <sup>•</sup>	1560
1561	CCATCGACATCTACCAGATATCGAGTTCGTGTACGGTATG <sup>•</sup>	1600
	C                          A                  GA	
1601	CTTCTGTAACCCCGATTCACTCAACGTTAATTGGGGTAA <sup>•</sup>	1640
	G          T	
1641	TTCATCCATTTT <sup>•</sup> TTCCAATACAGTACCAGCTACAGCTACG <sup>•</sup>	1680
	C  C                  T                  C	
1681	TCATTAGATAATCTACAATCAAGTGATTTTGGTTATTTTG <sup>•</sup>	1720
	C  G                  C  C  C          C  C	
1721	AAAGTGCCAATGCTTTTACATCTTCATTAGGTAATATAGT <sup>•</sup>	1760
	C  C                  C  C	
1761	AGGTGTTAGAAATTTTAGTGGGACTGCAGGAGTGATAATA <sup>•</sup>	1800
	G                  C                          T  C	
1801	GACAGATTTGAATTTATTCCAGTTACTGCAACACTCGAGG <sup>•</sup>	1840
	C  G  C	
1841	CTGAATATAATCTGGAAAGAGCGCAGAAGGCGGTGAATGC <sup>•</sup>	1880
1881	GCTGTTTACGTCTACAAACCAACTAGGGCTAAAAACAAAT <sup>•</sup>	1920
	G  C  C  C  G  C	
1921	GTAACGGATTATCATATTGATCAAGTGTCCAATTTAGTTA <sup>•</sup>	1960
	G                          C  G  G	
1961	CGTATTTATCGGATGAATTTTGTCTGGATGAAAAGCGAGA <sup>•</sup>	2000
	C  CC  CAGC          G  C	
2001	ATTGTCCGAGAAAGTCAAACATGCGAAGCGACTCAGTGAT <sup>•</sup>	2040
2041	GAACGCAATTTACTCCAAGATTCAAATTTCAAAGACATT <sup>•</sup> A	2080

FIG.10C

SUBSTITUTE SHEET

26 / 46

2081	ATAGGCAACCAGAACGTGGGTGGGGCGGAAGTACAGGGAT	2120
2121	TACCATCCAAGGAGGGGATGACGTATTTAAAGAAAATTAC	2160
	G T C G C G G C	
2161	GTCACACTATCAGGTACCTTTGATGAGTGCTATCCAACAT	2200
2201	ATTTGTATCAAAAAATCGATGAATCAAAATTAAAGCCTT	2240
	CC C C G G C G C G G	
2241	TACCCGTTATCAATTAAGAGGGTATATCGAAGATAGTCAA	2280
2281	GACTTAGAAATCTATTTAATTCGCTACAATGCAAACATG	2320
	C C G CC C C	
2321	AAACAGTAAATGTGCCAGGTACGGGTTCCCTTATGGCCGCT	2360
2361	TTCAGCCCAAAGTCCAATCGGAAAGTGTGGAGAGCCGAAT	2400
2401	CGATGCGCGCCACACCTTGAATGGAATCCTGACTTAGATT	2440
2441	GTTCGTGTAGGGATGGAGAAAAGTGTGCCCATCATTTCGA	2480
2481	TCATTTCTCCTTAGACATTGATGTAGGATGTACAGACTTA	2520
2521	AATGAGGACCTAGGTGTATGGGTGATCTTTAAGATTAAGA	2560
2561	CGCAAGATGGGCACGCAAGACTAGGGAATCTAGAGTTTCT	2600
2601	CGAAGAGAAACCATTAGTAGGAGAAGCGCTAGCTCGTGTG	2640
2641	AAAAGAGCGGAGAAAAAATGGAGAGACAAACGTGAAAAAT	2680
	G G	
2681	TGGAATGGGAAACAAATATCGTTTATAAAGAGGCAAAAGA	2720
	G C C C C	
2721	ATCTGTAGATGCTTTATTTGTAAACTCTCAATATGATCAA	2760
2761	TTACAAGCGGATACGAATATTGCCATGATTCATGCGGCAG	2800

FIG. 10D

SUBSTITUTE SHEET

27 / 46

2801	ATAAACGTGTTTCATAGCATTTCGAGAAGCTTATCTGCCTGA	2840
2841	GCTGTCTGTGATTCCGGGTGTCAATGCGGCTATTTTGA	2880
2881	GAATTAGAAGGGCGTATTTTCTACTGCATTCTCCCTATATG	2920
2921	ATGCGAGAAATGTCATTAAAAATGGTGATTTTAATAATGG	2960
2961	CTTATCCTGCTGGAACGTGAAAGGGCATGTAGATGTAGAA	3000
3001	GAACAAAACAACCAACGTTTCGGTCCTTGTGTGTTCCGGAAT	3040
3041	GGGAAGCAGAAGTGTCAACAAGAAGTTCGTGTCTGTCCGGG	3080
3081	TCGTGGCTATATCCTTCGTGTCACAGCGTACAAGGAGGGA	3120
3121	TATGGAGAAGGTTGCGTAACCATTTCATGAGATCGAGAACA	3160
3161	ATACAGACGAACTGAAGTTTAGCAACTGCGTAGAAGAGGA	3200
3201	AATCTATCCAAATAACACGGTAACGTGTAATGATTATACT	3240
3241	GTAAATCAAGAAGAATACGGAGGTGCGTACACTTCTCGTA	3280
3281	ATCGAGGATATAACGAAGCTCCTTCCGTACCAGCTGATTA	3320
3321	TGCGTCAGTCTATGAAGAAAATCGTATACAGATGGACGA	3360
3361	AGAGAGAATCCTTGTGAATTTAACAGAGGGTATAGGGATT	3400
3401	ACACGCCACTACCAGTTGGTTATGTGACAAAAGAATTAGA	3440
3441	ATACTTCCCAGAAACCGATAAGGTATGGATTGAGATTGGA	3480
3481	GAAACGGAAGGAACATTTATCGTGGACAGCGTGAATTAC	3520
3521	TCCTTATGGAGGAA	3534

**FIG. 10E**

# SUBSTITUTE SHEET

28 / 46

1	ATGGATAACAATCCGAACATCAATGAATGCATTCCTTATA	40
	C C A C A C	
41	ATTGTTTAAGTAACCCTGAAGTAGAAGTATTAGGTGGAGA	80
	C C G A T C T	
81	AAGAATAGAACTGGTTACACCCCAATCGATATTTCTTG	120
	C C T C T C C C	
121	TCGCTAACGCAATTTCTTTTGAGTGAATTTGTTCCCGGTG	160
	CT G A G GC C C G C G A	
161	CTGGATTTGTGTTAGGACTAGTTGATATAATATGGGGAAT	200
	G C TC C C C C T	
201	TTTTGGTCCCTCTCAATGGGACGCATTTCTTGTACAAATT	240
	C A T C G G	
241	GAACAGTTAATTAACCAAAGAATAGAAGAATTCGCTAGGA	280
	G G C G G C G C	
281	ACCAAGCCATTTCTAGATTAGAAGGACTAAGCAATCTTTA	320
	G C G G T G C	
321	TCAAATTTACGCAGAATCTTTTAGAGAGTGGAAGCAGAT	360
	C C T GAGC C C	
361	CCTACTAATCCAGCATTAAAGAGAAGAGATGCGTATTCAAT	400
	C TC CC C G A	
401	TCAATGACATGAACAGTGCCCTTACAACCGCTATTCCTCT	440
	C C T G C A C AT	
441	TTTTGCAGTTCAAAATTATCAAGTTCCTCTTTTATCAGTA	480
	G C C G C C C G C G	
481	TATGTTCAAGCTGCAAATTTACATTTATCAGTTTGGAGAG	520
	C A T C T CC CAGC GC TC	
521	ATGTTTCAGTGTTTGGACAAAGGTGGGGATTTGATGCCGC	560
	C AGC G C T	
561	GACTATCAATAGTCGTTATAATGATTTAACTAGGCTTATT	600
	A C C C C CC T G	
601	GGCAACTATACAGATTATGCTGTACGCTGGTACAATACGG	640
	A C C CC C T T C T	
641	GATTAGAACGTGTATGGGGACCGGATTCTAGAGATTGGGT	680
	C G G C T T A	

FIG. 11A

SUBSTITUTE SHEET

29 / 46

681	AAGGTATAATCAATTTAGAAGAGAATTAACACTAACTGTÄ	720
	T A C C G C G G C C A T	
721	TTAGATATCGTTGCTCTGTTCCCGAATTATGATAGTAGAA	760
	G C T GT C C CTCC	
761	GATATCCAATTCGAACAGTTTCCCAATTAACAAGAGAAAT	800
	CC C T C T G C T C	
801	TTATACAAACCCAGTATTAGAAAATTTTGATGGTAGTTTT	840
	C T TC T G C C C C C	
841	CGAGGCTCGGCTCAGGGCATAGAAAGAAGTATTAGGAGTC	880
	T T T C A T C G CTCC C C	
881	CACATTTGATGGATATACTTAACAGTATAACCATCTATAC	920
	C C CT G C T C	
921	GGATGCTCATAGGGGTTATTATTATTGGTCAGGGCATCAA	960
	C C A AG G C T A C G	
961	ATAATGGCTTCTCCTGTAGGGTTTTTCGGGGCCAGAATTCA	1000
	C C A T A CAGC C G T	
1001	CTTTTCCGCTATATGGAACATATGGGAAATGCAGCTCCACA	1040
	C T C C C	
1041	ACAACGTATTGTTGCTCAACTAGGTCAGGGCGTGTATAGA	1080
	C T C C	
1081	ACATTATCGTCCACTTTATATAGAAGACCTTTTAATATAG	1120
	C G T C G C C C C	
1121	GGATAAATAATCAACAACCTATCTGTTCTTGACGGGACAGA	1160
	T C C C G T C A	
1161	ATTTGCTTATGGAACCTCCTCAAATTTGCCATCCGCTGTÄ	1200
	G C C T T C T	
1201	TACAGAAAAAGCGGAACGGTAGATTCGCTGGATGAAATAC	1240
	G C T CT C C	
1241	CGCCACAGAATAACAACGTGCCACCTAGGCAAGGATTTAG	1280
	A C T C CTC	
1281	TCATCGATTAAAGCCATGTTTCAATGTTTCGTTTCAGGCTTT	1320
	C CA G G C G C C C A C	
1321	AGTAATAGTAGTGTAAGTATAATAAGAGCTCCTATGTTCT	1360
	C C TCC G C C C	
1361	CTTGGATACATCGTAGTGCTGAATTTAATAATATAATTGC	1400
	C G C C C C C	

FIG. 11B

SUBSTITUTE SHEET

30 / 46

1401	ATCGGATAGTATTACTCAAATCCCTGCAGTGAAGGGAAAC	1440
	C	
1441	TTTCTTTTAAATGGTCTGTAATTCAGGACCAGGATTTA	1480
	C C C C	
1481	CTGGTGGGGACTTAGTTAGATTAAATAGTAGTGGAATAA	1520
	A C C C C C	
1521	CATTCAGAATAGAGGGTATATTGAAGTTCCAATTCCTTC	1560
1561	CCATCGACATCTACCAGATATCGAGTTCGTGTACGGTATG	1600
	C A GA	
1601	CTTCTGTAACCCCGATTCACTCAACGTTAATTGGGGTAA	1640
	G T	
1641	TTCATCCATTTTTTCCAATACAGTACCAGCTACAGCTACG	1680
	C C T C	
1681	TCATTAGATAATCTACAATCAAGTGATTTTGGTTATTTTG	1720
	C G C C C C	
1721	AAAGTGCCAATGCTTTTACATCTTCATTAGGTAATATAGT	1760
	C C C C	
1761	AGGTGTTAGAAATTTTAGTGGGACTGCAGGAGTGATAATA	1800
	G C T C	
1801	GACAGATTTGAATTTATTCCAGTTACTGCAACACTCGAGG	1840
	C G C	
1841	CTGAATATAATCTGGAAAGAGCGCAGAAGGCGGTGAATGC	1880
	G C C T G C T C	
1881	GCTGTTTACGTCTACAAACCACTAGGGCTAAAAACAAAT	1920
	C C C C C T G T CT G T C	
1921	GTAACGGATTATCATATTGATCAAGTGTCCTCAATTTAGTTA	1960
	T T C C C C G C	
1961	CGTATTTATCGGATGAATTTGTCTGGATGAAAAGCGAGA	2000
	C CC TAGC G C C C C G T	
2001	ATTGTCCGAGAAAGTCAAACATGCGAAGCGACTCAGTGAT	2040
	C C T C C T C C	
2041	GAACGCAATTTACTCCAAGATTCAAATTTCAAAGACATTA	2080
	GA G C CT G C C C C	
2081	ATAGGCAACCAGAACGTGGGTGGGGCGGAAGTACAGGGAT	2120
	C G T T C C	

FIG. 11C

SUBSTITUTE SHEET

31 / 46

2121	TACCATCCAAGGAGGGGATGACGTATTTAAAGAAAATTAC	2160
	C C C T G C G G C	
2161	GTCACACTATCAGGTACCTTTGATGAGTGCTATCCAACAT	2200
	C C C A T C C C T C	
2201	ATTTGTATCAAAAAATCGATGAATCAAAATTAAAAGCCTT	2240
	C C G G G C C C	
2241	TACCCGTTATCAATTAAGAGGGTATATCGAAGATAGTCAA	2280
	C A G C T C C C C	
2281	GACTTAGAAATCTATTTAATTCGCTACAATGCAAAACATG	2320
	C T C CG CA G C G C	
2321	AAACAGTAAATGTGCCAGGTACGGGTTCTTATGGCCGCT	2360
	G C G C T C C A	
2361	TTCAGCCCAAAGTCCAATCGGAAAGTGTGGAGAGCCGAAT	2400
	T TC C T G T C	
2401	CGATGCGCGCCACACCTTGAATGGAATCCTGACTTAGATT	2440
	A T G G C	
2441	GTTCTGTAGGGATGGAGAAAAGTGTGCCCATCATTCGCA	2480
	C C C C G C T	
2481	TCATTTCTCCTTAGACATTGATGTAGGATGTACAGACTTA	2520
	C G C G T C G	
2521	AATGAGGACCTAGGTGTATGGGTGATCTTTAAGATTAAGA	2560
	C A C C C C	
2561	CGCAAGATGGGCACGCAAGACTAGGGAATCTAGAGTTTCT	2600
	C C A T C C T	
2601	CGAAGAGAAACCATTAGTAGGAGAAGCGCTAGCTCGTGTG	2640
	G C T T C	
2641	AAAAGAGCGGAGAAAAATGGAGAGACAAACGTGAAAAAT	2680
	G A G G G G C	
2681	TGGAATGGGAAACAAATATCGTTTATAAAGAGGCAAAAGA	2720
	C T C C G C	
2721	ATCTGTAGATGCTTTATTTGTAAACTCTCAATATGATCAA	2760
	G C G G C G C G	
2761	TTACAAGCGGATACGAATATTGCCATGATTCATGCGGCAG	2800
	G C C C C C C C C	
2801	ATAAACGTGTTTCATAGCATTTCGAGAAGCTTATCTGCCTGA	2840
	C G C T G CT	

FIG. 11D

SUBSTITUTE SHEET

32 / 46

2841	GCTGTCTGTGATTCCGGGTGTCAATGCGGCTATTTTGA	2880
	T C C T G C T C C C G	
2881	GAATTAGAAGGGCGTATTTTCACTGCATTCTCCCTATATG	2920
	C T G A C T C T G C	
2921	ATGCGAGAAATGTCATTAAAAATGGTGATTTTAATAATGG	2960
	C C C G C C C C	
2961	CTTATCCTGCTGGAACGTGAAAGGGCATGTAGATGTAGAA	3000
	C CAG T T G C G G	
3001	GAACAAAACAACCAACGTTCCGGTCCTTGTTGTTCCGGAAT	3040
	G T G C G G T G	
3041	GGGAAGCAGAAGTGTCAACAAGAAGTTCGTGTCTGTCCGGG	3080
	T C G A A A	
3081	TCGTGGCTATATCCTTCGTGTCACAGCGTACAAGGAGGGA	3120
	A A C T C G C T	
3121	TATGGAGAAGGTTGCGTAACCATTTCATGAGATCGAGAACA	3160
	C T G G C C	
3161	ATACAGACGAACTGAAGTTTAGCAACTGCGTAGAAGAGGA	3200
	C C G T CTC C G A	
3201	AATCTATCCAAATAACACGGTAACGTGTAATGATTATACT	3240
	C C C T T C C C C	
3241	GTAAATCAAGAAGAATACGGAGGTGCGTACACTTCTCGTA	3280
	G G G C AGC	
3281	ATCGAGGATATAACGAAGCTCCTTCCGTACCAGCTGATTA	3320
	CA T C T T C	
3321	TGCGTCAGTCTATGAAGAAAATCGTATACAGATGGACGA	3360
	C C G C G G C C CA	
3361	AGAGAGAATCCTTGTGAATTTAACAGAGGGTATAGGGATT	3400
	C T C C G C T C C	
3401	ACACGCCACTACCAGTTGGTTATGTGACAAAAGAATTAGA	3440
	A T C T C G GC T	
3441	ATACTTCCCAGAAACCGATAAGGTATGGATTGAGATTGGA	3480
	G T T G C A G C C T	
3481	GAAACGGAAGGAACATTTATCGTGGACAGCGTGGAATTAC	3520
	C G C C GC T	
3521	TCCTTATGGAGGAA	3534
	T G	

FIG. 11E

SUBSTITUTE SHEET



33/46

1	ATGACTGCAGATAATAATACGGAAGCACTAGATAGCTCTA	40
	C C C C C C C T	
41	CAACAAAAGATGTCATTCAAAAAGGCATTTCCTAGTAGG	80
	C T G T C G G T C T G	
81	TGATCTCCTAGGCGTAGTAGGTTCCCGTTTGGTGGAGCG	120
	A C T G G T A T C C C	
121	CTTGTTTCGTTTTATACAACTTTTTAAATACTATTGGC	160
	C GAGC C C C C C	
161	CAAGTGAAGACCCGTGGAAGGCTTTTATGGAACAAGTAGA	200
	C G T A A C G T	
201	AGCATTGATGGATCAGAAAATAGCTGATTATGCAAAAAT	240
	TC T G T A C G C	
241	AAAGCTCTTGCGAGTTACAGGGCCTTCAAAATAATGTCG	280
	G T G AC C G C G	
281	AAGATTATGTGAGTGCATTGAGTTCATGGCAAAAAATCC	320
	G C C TCCAGC G G C	
321	TGTGAGTTCACGAAATCCACATAGCCAGGGGCGGATAAGA	360
	T C CA T C A TA C	
361	GAGCTGTTTTCTCAAGCAGAAAGTCATTTTCGTAATTCAA	400
	T C C TCC C CA A C	
401	TGCCTTCGTTTGCAATTTCTGGATACGAGGTTCTATTTCT	440
	AGC T C C T T C	
441	AACAACATATGCACAAGCTGCCAACACACATTTATTTTTTA	480
	C T C T C C G C C	
481	CTAAAAGACGCTCAAATTTATGGAGAAGAATGGGGATACG	520
	T G C G	
521	AAAAAGAAGATATTGCTGAATTTTATAAAAGACAATAAA	560
	G G C G C C GC T T	
561	ACTTACGCAAGAATATACTGACCATTGTGTCAAATGGTAT	600
	G C C G C C G	
601	AATGTTGGATTAGATAAATTAAGAGGTTTCATCTTATGAAT	640
	C TC C GC C C T C C G	
641	CTTGGGTAAACTTTAACCGTTATCGCAGAGAGATGACATT	680
	G C A A CA G C	

FIG. 12A

SUBSTITUTE SHEET

34 / 46

681	AACAGTATTAGATTTAATTGCACTATTTCCATTGTATGAT	720
	G T GC C C T C C C C	
721	GTTCGGCTATACCCAAAAGAAGTTAAAACCGAATTAACAA	760
	GA A C G G T GC T C	
761	GAGACGTTTTTAACAGATCCAATTGTCGGAGTCAACAACCT	800
	GC C T C T	
801	TAGGGGCTATGGAACAACCTTCTCTAATATAGAAAATTAT	840
	T T AGC C C C	
841	ATTCGAAAACACATCTATTTGACTATCTGCATAGAATTC	880
	A G C C T C	
881	AATTCACACGCGGTTCCAACCAGGATATTATGGAAATGA	920
	C AA T C T C	
921	CTCTTTCAATTATTGGTCCGTAATTATGTTTCAACTAGA	960
	C C C C C	
961	CCAAGCATAGGATCAAATGATATAATCACATCTCCATTCT	1000
	T T C C C	
1001	ATGGAAATAAATCCAGTGAACCTGTACAAAATTTAGAATT	1040
	T C G G G CC T G	
1041	TAATGGAGAAAAAGTCTATAGAGCCGTAGCAAATACAAAT	1080
	C C C G C C C	
1081	CTTGCGGTCTGGCCGTCCGCTGTATATTCAGGTGTTACAA	1120
	C T G A A T C C C	
1121	AAGTGAATTTAGCCAATATAATGATCAAACAGATGAAGC	1160
	G G T G C G C G	
1161	AAGTACACAAACGTACGACTCAAAAAGAAATGTTGGCGCG	1200
	C C C G T C C T C A	
1201	GTCAGCTGGGATTCTATCGATCAATTGCCTCCAGAAACAA	1240
	TCT C C	
1241	CAGATGAACCTCTAGAAAAGGGATATAGCCATCAACTCAA	1280
	C AT G G C C C T	
1281	TTATGTAATGTGCTTTTTAATGCAGGGTAGTAGAGGAACA	1320
	C G C G A TCC G C	
1321	ATCCCAGTGTTAACTTGGACACATAAAAGTGTAGACTTTT	1360
	T G C C GTCC G C	
1361	TTAACATGATTGATTGCAAAAAATTACACAACCTCCGTT	1400
	C C AGC G G C T C	

FIG.12B

SUBSTITUTE SHEET

35 / 46

1401	AGTAAAGGCATATAAGTTACAATCTGGTGCTTCGTTGTC	1440
	G G A C C C G	
1441	GCAGGTCCTAGGTTTACAGGAGGAGATATCATTCAATGCA	1480
	C A C T T C C G	
1481	CAGAAAATGGAAGTGCGGCAACTATTTACGTTACACCGGA	1520
	G C C C A T C G T	
1521	TGTGTCGTA CTCTCAAAAATATCGAGCTAGAATTCATTAT	1560
	T G G C A G A C T C	
1561	GCTTCTACATCTCAGATAACATTTACACTCAGTTTAGACG	1600
	A CAGC C C C C G T	
1601	GGGCACCATTTAATCAATACTATTTTCGATAAAACGATAAA	1640
	A C C C G T C T C G C C	
1641	TAAAGGAGACACATTAACGTATAATTCATTTAATTTAGCA	1680
	C T T C C A C A G C C C G	
1681	AGTTTCAGCACACCATTTCGAATTATCAGGGAATAACTTAC	1720
	T C C C C T C T	
1721	AAATAGGCGTCACAGGATTAAAGTGCTGGAGATAAAGTTTA	1760
	G C C T C C C C C C	
1761	TATAGACAAAATTGAATTTATTCCAGTGAAT	1791
	C C G G C C C	

FIG.12C

SUBSTITUTE SHEET

36 / 46

1	ATG AATAATGTATTGAATAGTGGAAGAACAACACTATTT	40
	GAC C C C CTC T C C	
41	GTGATGCGTATAATGTAGTAGCCCATGATCCATTTAGTTT	80
	C C A C C C G T C C C	
81	TGAACATAAATCATTAGATACCATCCAAAAGAATGGATG	120
	C C GAGCC C C T T G G G	
121	GAGTGGAAAAGAACAGATCATAGTTTATATGTAGCTCCTG	160
	A C T T C CTC C C C C A	
161	TAGTCGGAACGTGTCTAGTTTTTTGCTAAAGAAAGTGGG	200
	G T A C C CC T C G C	
201	GAGTCTTATTGGAAAAAGGATATTGAGTGAATTATGGGGG	240
	CTC C C C T C TCC C C T	
241	ATAATATTTCTAGTGGTAGTACAAATCTAATGCAAGATA	280
	C C ATC GTCC T C C	
281	TTTAAAGGGAGACAGAACAATTCCTAAATCAAAGACTTAA	320
	C G C G T C C GC T C	
321	TACAGATACCCTTGCTCGTGTAATGCAGAATTGATAGGG	360
	C T T G A A C C T G C T	
361	CTCCAAGCGAATATAAGGGAGTTTAATCAACAAGTAGATA	400
	A C TC T C C G G C	
401	ATTTTTTAAACCCTACTCAAAACCCTGTTCTTTATCAAT	440
	C C G T A G T G C T C	
441	AACTTCTTCGGTTAATACAATGCAGCAATTATTTCTAAAT	480
	C C G C T C C C C C	
481	AGATTACCCAGTTCAGATACAAGGATACCAGTTGTTAT	520
	G T T T C C CC	
521	TATTACCTTTATTTGCACAGGCAGCCAATATGCATCTTTC	560
	TC T AC C T T C CT G	
561	TTTTATTAGAGATGTTATTCTTAATGCAGATGAATGGGGT	600
	C C AC T C G C C C T C A	
601	ATTTTCAGCAGCAACATTACGTACGTATCGAGATTACCTGA	640
	C T C TC TA G A CA C T	
641	GAAATTATACAAGAGATTATTCTAATTATTGTATAAATAC	680
	G C C TC T C C C C C C	

FIG. 13A

SUBSTITUTE SHEET

37 / 46

681	GTATCAA <sup>.</sup> ACTGCGTTT <sup>.</sup> AGAGGGT <sup>.</sup> TAAACACCCGTTT <sup>.</sup> TACAC <sup>.</sup>	720
	T     G     C     C T   AC C   T   TA GC T	
721	GATATGTTAGA <sup>.</sup> AATTTAGA <sup>.</sup> ACATATATGTTT <sup>.</sup> TAAATGTAT <sup>.</sup>	760
	C   C T   G   C   G   C   C     CC T   C   G	
761	TTGAATATGTATCCATT <sup>.</sup> TGGTCATTGTTT <sup>.</sup> TAAATATCAGAG <sup>.</sup>	800
	G   C   CAG     AGTC C   C   G   C	
801	TCTTATGGTATCTTCTGGCGCTAATTTATATGCTAGCGGT <sup>.</sup>	840
	CT G     G   C     A   C   C C   C   CTCT   C	
841	AGTGGACCACAGCAGACACAATCATT <sup>.</sup> TACAGCACAAA <sup>.</sup> ACT <sup>.</sup>	880
	A   T   GAGC   C     T   G	
881	GGCCATTTTATATTCTCTTTTCCAAGTTAATTCGAATTA <sup>.</sup>	920
	C   G   AGCT G     C   C   C   C	
921	TATATTATCTGGTATTAGTGGTACTAGGCTTTCTATTACC <sup>.</sup>	960
	C   TC CAG     CTC   G   C   A   C   C   A	
961	TTCCCTAATATTGGTGGTTTACCGGGTAGTACTACA <sup>.</sup> ACTC <sup>.</sup>	1000
	T   C   C     AC T   A   CTCC     C	
1001	ATTCATTGAATAGTGCCAGGGTTAATTATAGCGGAGGAGT <sup>.</sup>	1040
	AGCC T   CTC     A   G   C   C   T     T	
1041	TTCATCTGGTCTCATAGGGGCGACTAATCTCAATCACAAC <sup>.</sup>	1080
	CAGC     AT G   T   T   A     CT G   C	
1081	TTTAATTGCAGCACGGTCCTCCCTCCTTTATCAACACCAT <sup>.</sup>	1120
	C     TC   C   T   G   A   C   GAGC     G	
1121	TTGTTAGAAGTTGGCTGGATT <sup>.</sup> CAGGTACAGATCGAGAGGG <sup>.</sup>	1160
	G   GTCC     T   CAGC     T     C   A	
1161	CGTTGCTACCTCTACGAATTGGCAGACAGAATCCTTTCAA <sup>.</sup>	1200
	A     A   C     A   C   G     C	
1201	ACAAC <sup>.</sup> TTAAGTTTAAGGTGTGGTGCTTTTTCAGCCCGTG <sup>.</sup>	1240
	C   C T   CC TC     A     C   T   A	
1241	GAAATTCAA <sup>.</sup> ACTATTTCCCAGATTATTTTATCCGTAATAT <sup>.</sup>	1280
	G     C   T     C   C   C   TA G   C	
1281	TTCTGGGGTTCTTTAGTTATTAGAAACGAAGATCTAACA <sup>.</sup>	1320
	C     T     C C C   C   G   T     C   C   C	
1321	AGACCGTTACACTATAACCAAATAAGAAATATAGAAAGTC <sup>.</sup>	1360
	C T   AC T   T   C     G   T   G   C   C   GTC	
1361	CTTCGGGAACACCTGGTGGAGCACGGGCCTATTTGGTATC <sup>.</sup>	1400
	A   C   T   T   A   A   T   A A   T   CC C   G	

FIG.13B

SUBSTITUTE SHEET

38 / 46

1401	TGTGCATAACAGAAAAAATAATATCTATGCCGCTAATGAA	1440
	C G G C C C T C C G	
1441	AATGGTACTATGATCCATTTGGCGCCAGAAGATTATACAG	1480
	C C T CC T A C T	
1481	GATTTACTATATCGCCAATACATGCCACTCAAGTGAATAA	1520
	C C C T C T C C	
1521	TCAAACTCGAACATTTATTTCTGAAAAATTTGGAAATCAA	1560
	G A C C C C C G C	
1561	GGTGATTCCTTAAGATTTGAACAAAGCAACACGACAGCTC	1600
	C G G C G TC T C A	
1601	GTTATACGCTTAGAGGGAATGGAAATAGTTACAATCTTTA	1640
	G C TT G C C C C	
1641	TTTAAGAGTATCTTCAATAGGAAATTCAACTATTCGAGTT	1680
	C G TAGC C T T C C C C T	
1681	ACTATAAACGGTAGAGTTTATACTGTTTCAAATGTTAATA	1720
	C C AC T C A C T G C	
1721	CCACTACAAATAACGATGGAGTTAATGATAATGGAGCTCG	1760
	T A G C T C C C C CA	
1761	TTTTTCAGATATTAATATCGGTAATATAGTAGCAAGTGAT	1800
	A CAGC C C C T C C C G CTC C	
1801	AATACTAATGTAACGCTAGATATAAATGTGACATTAACT	1840
	C C T TT G C C CC C T	
1841	CCGGTACTCCATTTGATCTCATGAATATTATGTTTGTGCC	1880
	T A C C	
1881	AACTAATCTTCCACCACTTTAT	1902
	C C T T G C	

FIG. 13C

SUBSTITUTE SHEET

39 / 46

1	ATGGAGGAAAATAATCAAAATCAATGCATACCTTACAATT	40
	G C C C T A C	
41	GTTTAAGTAATCCTGAAGAAGTACTTTTGGATGGAGAACG	80
	C G C A G T GC T	
81	GATATCAACTGGTAATTCATCAATTGATATTTCTCTGTCA	120
	C T C C T C C C C CT C	
121	CTTGTTTCAGTTTCTGGTATCTAACTTTGTACCAGGGGGAG	160
	T G C CAGC C G T T	
161	GATTTTATAGTTGGATTAATAGATTTTGTATGGGGAATAGT	200
	G CC T C C T C C C T C	
201	TGGCCCTTCTCAATGGGATGCATTTCTAGTACAAATTGAA	240
	T A C G G G	
241	CAATTAATTAATGAAAGAATAGCTGAATTTGCTAGGAATG	280
	G G C C G G C G C C C	
281	CTGCTATTGCTAATTTAGAAGGATTAGGAAACAATTTCAA	320
	C C C G G C T C	
321	TATATATGTGGAAGCATTTAAAGAATGGGAAGAAGATCCT	360
	C C G C C G G C	
361	AATAATCCAGAAACCAGGACCAGAGTAATTGATCGCTTTC	400
	C G C C T G G C CA A CA	
401	GTATACTTGATGGGCTACTTGAAAGGGACATTCCTTCGTT	440
	A CT G C C CT G G A T C A C	
441	TCGAATTTCTGGATTTGAAGTACCCCTTTTATCCGTTTAT	480
	CA C C C T T C G G C	
481	GCTCAAGCGGCAATCTGCATCTAGCTATATTAAGAGATT	520
	A T T C C CC TC CA	
521	CTGTAATTTTGGAGAAAGATGGGGATTGACAACGATAAA	560
	G C C G G C T C	
561	TGTCAATGAAACTATAATAGACTAATTAGGCATATTGAT	600
	C G T C C T C C C	
601	GAATATGCTGATCACTGTGCAAATACGTATAATCGGGGAT	640
	G C C C T C C C C T C	
641	TAAATAATTTACCGAAATCTACGTATCAAGATTGGATAAC	680
	G C C C T G T T	
681	ATATAATCGATTACGGAGAGACTTAACATTGACTGTATT	720
	C C CA G GA G CC C A T G	

FIG. 14A

SUBSTITUTE SHEET

40 / 46

721	GATATCGCCGCTTTCTTTCCAACTATGACAATAGGAGAT	760
	C T A C G C	
761	ATCCAATTCAGCCAGTTGGTCAACTAACAAGGGAAGTTTA	800
	C T C A G T C A C	
801	TACGGACCCATTAATTAATTTTAATCCACAGTTACAGTCT	840
	T C T C C C T G AAG	
841	GTAGCTCAATTACCTACTTTTAACGTTATGGAGAGCAGCC	880
	C C C T C A C C TC	
881	GAATTAGAAATCCTCATTATTGATATATTGAATAATCT	920
	T C G C A C G C C C C	
921	TACAATCTTTACGGATTGGTTTAGTGTTGGACGCAATTTT	960
	T C C C C G T C C	
961	TATTGGGGAGGACATCGAGTAATATCTAGCCTTATAGGAG	1000
	T CA G C C CTCT T	
1001	GTGGTAACATAACATCTCCTATATATGGAAGAGAGGCGAA	1040
	G T C C C T A	
1041	CCAGGAGCCTCCAAGATCCTTTACTTTTAATGGACCGGTA	1080
	A C TAGT C C C C T A C	
1081	TTTAGGACTTTATCAAATCCTACTTTACGATTATTACAGC	1120
	C A C G T C C GA GC C	
1121	AACCTTGGCCAGCGCCACCATTTAATTTACGTGGTGTGA	1160
	T T C CC TA A	
1161	AGGAGTAGAATTTTCTACACCTACAAATAGCTTTACGTAT	1200
	G C T G C T C CTC C T C	
1201	CGAGGAAGAGGTACGGTTGATTCTTTAACTGAATTACCGC	1240
	A T A C C G C C C A	
1241	CTGAGGATAATAGTGTGCCACCTCGCGAAGGATATAGTCA	1280
	A C C CA G C CTCC	
1281	TCGTTTATGTCATGCAACTTTTGTTCAAAGATCTGGAACA	1320
	CA G G C C C C G GC T C T	
1321	CCTTTTTTAACAACCTGGTGTAGTATTTTCTTGGACCGATC	1360
	A CC C T A A T G C A T	
1361	GTAGTGCAACTCTTACAAATACAATTGATCCAGAGAGAAT	1400
	T C T C C G	

FIG. 14B

SUBSTITUTE SHEET



41 / 46

1401	TAATCAAATACCTTTAGTGAAAGGATTTAGAGTTTGGGGG	1440
	C C A G C G T CC T G A	
1441	GGCACCTCTGTCATTACAGGACCAGGATTTACAGGAGGGG	1480
	A T C C C T	
1481	ATATCCTTCGAAGAAATACCTTTGGTGATTTTGTATCTCT	1520
	T A C T C C GAGC	
1521	ACAAGTCAATATTAATTCACCAATTACCCAAAGATACCGT	1560
	C T C C C T T T	
1561	TTAAGATTTCTGTTACGCTTCCAGTAGGGATGCACGAGTTA	1600
	C C G A TTCC T C TA C	
1601	TAGTATTAACAGGAGCGGCATCCACAGGAGTGGGAGGCCA	1640
	C GC C C C A T T C T C T A	
1641	AGTTAGTGTAATATGCCTCTTCAGAAAACATATGGAAATA	1680
	CTCC G C A C G G C	
1681	GGGGAGAACTTAACATCTAGAACATTTAGATATACCGATT	1720
	C G C G C C C C	
1721	TTAGTAATCCTTTTTCATTTAGAGCTAATCCAGATATAAT	1760
	CTC C CAGT CC T C C T C C	
1761	TGGGATAAGTGAACAACCTCTATTTGGTGCAGGTTCTATT	1800
	C T C C A T AGC C	
1801	AGTAGCGGTGAACCTTATATAGATAAAATTGAAATTATTC	1840
	TCATCT C T G C T C G G C	
1841	TAGCAGATGCAACATTTGAAGCAGAATCTGATTTAGAAAG	1880
	T C C T CC C G T G ACA CC T G	
1881	AGCACAAAAGGCGGTGAATGCCCTGTTTACTTCTTCCAAT	1920
	C G T C C C CA	
1921	CAAATCGGGTTAAAAACCGATGTGACGGATTATCATATTG	1960
	GC T C G TA C T T C C	
1961	ATCAAGTATCCAATTTAGTGGATTGTTTATCAGATGAATT	2000
	C G C G CACC ACC TAGC G	
2001	TTGTCTGGATGAAAAGCGAGAATTGTCCGAGAAAGTCAA	2040
	C C C C G T C C T	
2041	CATGCGAAGCGACTCAGTGATGAGCGGAATTTACTTCAAG	2080
	C C T C C A C CT G	
2081	ATCCAAACTTCAGAGGGATCAATAGACAACCAGACCGTGG	2120
	CT C A AC C G G A	

FIG. 14C

SUBSTITUTE SHEET

42 / 46

2121	CTGGAGAGGAAGTACAGATATTACCATCCAAGGAGGAGAT	2160
	T G T C C GG C C C	
2161	GACGTATTCAAAGAGAATTACGTCACACTACCGGGTACCG	2200
	T G G C C CT C A TT	
2201	TTGATGAGTGCTATCCAACGTATTTATATCAGAAAATAGA	2240
	C C C T C C G C G C	
2241	TGAGTCGAAATTAAAAGCTTATACCCGTTATGAATTAAGA	2280
	C C C C TC A G C C T	
2281	GGGTATATCGAAGATAGTCAAGACTTAGAAATCTATTTGA	2320
	C C C C C T C C	
2321	TCCGTTACAATGCAAAACACGAAATAGTAAATGTGCCAGG	2360
	A G C G G CC G C	
2361	CACGGGTTCCTTATGGCCGCTTTCAGCCCAAATGCCAATC	2400
	T T C C A T TCT C T	
2401	GGAAAGTGTGGAGAACCGAATCGATGCGCGCCACACCTTG	2440
	G G T CA T	
2441	AATGGAATCCTGATCTAGATTGTTCTGCAGAGACGGGGA	2480
	G CT G C C G T C	
2481	AAAATGTGCACATCATTCCCATCATTTACCTTGATATT	2520
	G G C C T C T C C	
2521	GATGTTGGATGTACAGACTTAAATGAGGACTTAGGTGTAT	2560
	G T C G C C A C	
2561	GGGTGATATTCAAGATTAAGACGCAAGATGGCCATGCAAG	2600
	C C C C C A C	
2601	ACTAGGGAATCTAGAGTTTCTCGAAGAGAAACCATTATTA	2640
	T C C T GG C	
2641	GGGGAAGCACTAGCTCGTGTGAAAAGAGCGGAGAAGAAGT	2680
	T T C G A	
2681	GGAGAGACAAACGAGAGAAACTGCAGTTGGAAACAAATAT	2720
	G T CG A G T C	
2721	TGTTTATAAAGAGGCAAAAGAATCTGTAGATGCTTTATTT	2760
	C C G C G C G G C	
2761	GTAAACTCTCAATATGATAGATTACAAGTGGATACGAACA	2800
	G C CAG G CC C C	
2801	TCGCCATGATTCATGCGGCAGATAAACGCGTTCATAGAAT	2840
	C C C C T G C C	

FIG. 14D

SUBSTITUTE SHEET

43 / 46

2841	CCGGAAGCGTATCTGCCAGAGTTGTCTGTGATTCCAGGT	2880
	T T G T CT T C C T	
2881	GTCAATGCGGCCATTTTCGAAGAATTAGAGGGACGTATTT	2920
	G C T C G C T C	
2921	TTACAGCGTATTCTTATATGATGCGAGAAATGTCATTAA	2960
	C A TC G C C C C	
2961	AAATGGCGATTTCAATAATGGCTTATTATGCTGGAACGTG	3000
	G C T C C C CAGC T	
3001	AAAGGTCATGTAGATGTAGAAGAGCAAACAACCACCGTT	3040
	G C G G A G T G	
3041	CGGTCCTTGTTATCCCAGAATGGGAGGCAGAAGTGTACAC	3080
	C G G G T G A T C	
3081	AGAGGTTCGTGTCTGTCCAGGTCGTGGCTATATCCTTCGT	3120
	A A A A C T C	
3121	GTCACAGCATATAAAGAGGGATATGGAGAGGGCTGCGTAA	3160
	G C T C G C T T T G	
3161	CGATCCATGAGATCGAAGACAATACAGACGAACTGAAATT	3200
	C C GA C C G T G	
3201	CAGCAACTGTGTAGAAGAGGAAGTATATCCAAACAACACA	3240
	TC C C G A A C C C	
3241	GTAACGTGTAATAATTATACTGGGACTCAAGAAGAATATG	3280
	T T C CG C C T A G G C	
3281	AGGGTACGTACACTTCTCGTAATCAAGGATATGACGAAGC	3320
	GA G C AGC CAG T CA	
3321	CTATGGTAATAACCCTTCCGTACCAGCTGATTACGCTTCA	3360
	TCC TCXXXXXXXXXX T T C T C C	
3361	GTCTATGAAGAAAATCGTATACAGATGGACGAAGAGAGA	3400
	G C G G C C CA C T	
3401	ATCCTTGTGAATCTAACAGAGGCTATGGGGATTACACACC	3440
	C C G TC T CA C	
3441	ACTACCGGCTGGTTATGTAACAAAGGATTTAGAGTACTTC	3480
	T A T C T C GC T T	
3481	CCAGAGACCGATAAGGTATGGATTGAGATCGGAGAAACAG	3520
	T C A G C T C	
3521	AAGGAACATTCATCGTGGATAGCGTGGAATTACTCCTTAT	3560
	G C C GC T T G	
3561	GGAGGAA	3567

FIG.14E

SUBSTITUTE SHEET

44 / 46

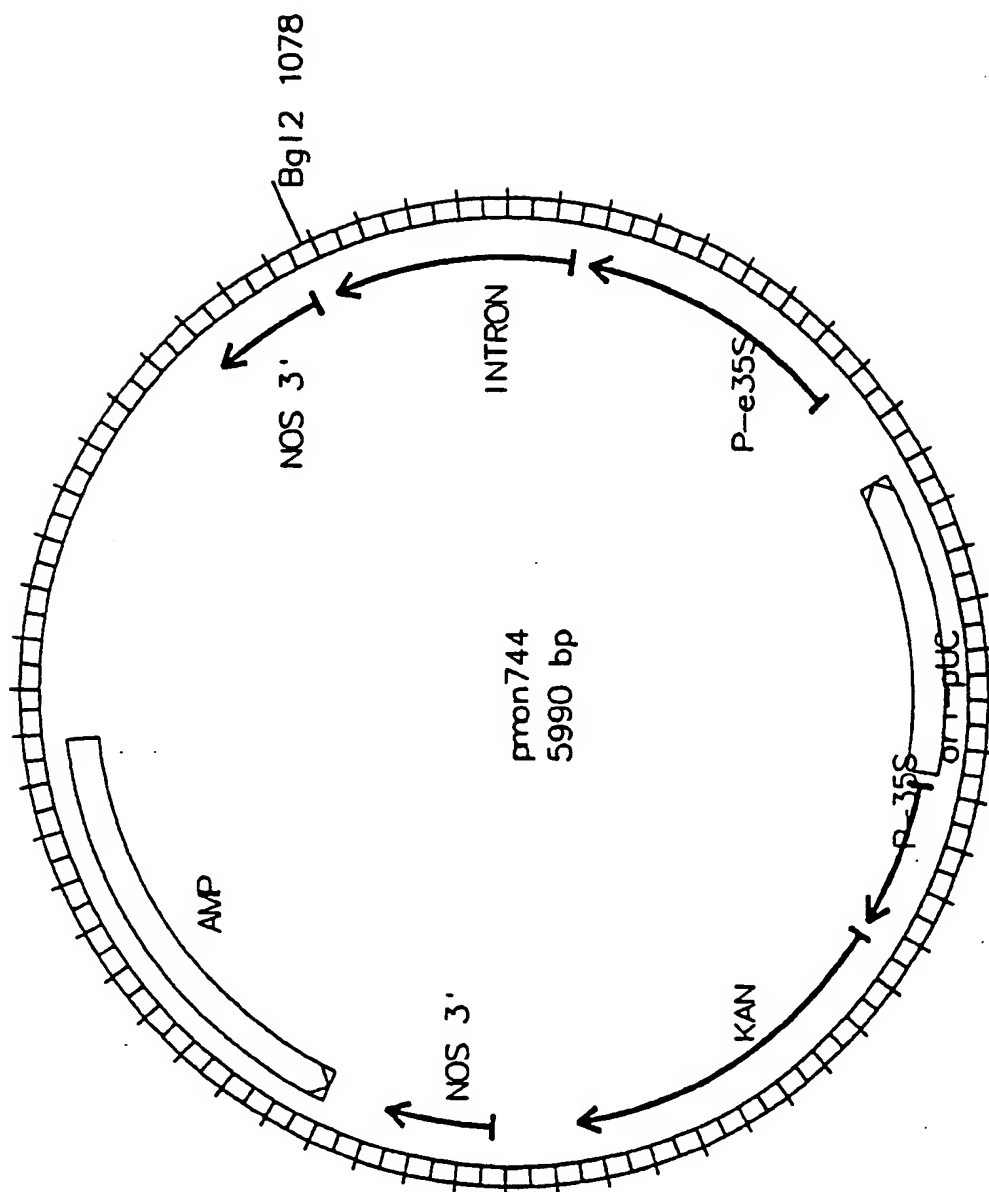


FIG.15

45 / 46

1 AGATCTAGAGGTAATTGTTATGAGTACTGTCGTGGTTAAG 40  
GATC

41 GGAAACGTCAACGGTGGTGTACAACAACCTAGAAGGAGGA 80  
G T A

81 GAAGGCAATCCCTTCGCAGGAGGGCTAACAGAGTACAGCC 120  
T A T

121 AGTGGTTATGGTCACTGCTCCTGGCGAACCAGGAGGAGG 160  
GC A A A

161 AGACGCAGAAGAGGAGGCAATCGCAGGTCAAGAAGAAGT 200  
A G T A

201 GAGTTCACAGGGGAAGGGGCTCAAGCGAGACATTCGTGTT 240  
A A T

241 TACAAAGGACAACCTCGTGGGCAACTCCCAAGGAAGTTTC 280

281 ACCTTCGGACCAAGTGTATCAGACTGTCCAGCATTCAAGG 320  
T

321 ATGGAATACTCAAGGCCTACCATGAGTACAAGATCACAAG 360  
T

361 TATCCTTCTTCAGTTCGTCAGCGAGGCCTCTTCCACCTCA 400  
T G T

401 CCAGGATCCATCGCTTATGAGTTGGACCCACATTGCAAAG 440  
C A T

441 TATCATCCCTCCAGTCCTACGTCAACAAGTTCCAAATCAC 480  
T

481 AAAGGGAGGAGCTAAGACCTATCAAGCTAGGATGATCAAC 520  
T T C T

521 GGAGTAGAATGGCACGATTCATCTGAGGATCAGTGCAGGA 560  
T T A

561 TACTTTGGAAAGGAAGTGGAAAATCTTCAGACCCAGCAGG 600  
C A G T T

601 ATCTTTTCAGAGTCACCATCAGAGTGGCTCTTCAAACCCC 640  
T T A

641 AAGTAATAGACTCCGGATCAGAGCCTGGTCCAAGCCCACA 680  
A T

FIG. 16A

SUBSTITUTE SHEET

46 / 46

681 ACCAACACCCACTCCAACCTCCCCAAAAGCATGAGCGATT 720  
721 ATTGCTTACGTCGGCATACCTATGCTGACCATTCAAGAAT 760  
761 TC 762

**FIG. 16B****SUBSTITUTE SHEET**

# INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 90/00778

## I. CLASSIFICATION OF SUBJECT MATTER (if several classification symbols apply, indicate all) \*

According to International Patent Classification (IPC) or to both National Classification and IPC

IPC<sup>5</sup>: C 12 N 15/82, 15/32, 15/67, 15/40, 5/10, A 01 H 5/00

## II. FIELDS SEARCHED

Minimum Documentation Searched <sup>7</sup>

Classification System |

Classification Symbols

IPC<sup>5</sup>

C 12 N, A 01 H

Documentation Searched other than Minimum Documentation  
to the Extent that such Documents are Included in the Fields Searched <sup>8</sup>

## III. DOCUMENTS CONSIDERED TO BE RELEVANT \*

Category <sup>9</sup>	Citation of Document, <sup>11</sup> with Indication, where appropriate, of the relevant passages <sup>12</sup>	Relevant to Claim No. <sup>13</sup>
Y	EP, A, 0142924 (LUBRIZOL GENETICS INC.) 29 May 1985 see page 26, lines 20-30 --	1,30,31
Y	UCLA Symp. Mol. Cell. Biol., New Ser. volume 48, 1987, Molecular Strategies for Crop Protection, Alan R. Liss, Inc., M.J. Adang et al.: "Expression of a <u>Bacillus thuringiensis</u> insecticidal crystal protein gene in tobacco plants", pages 345-353 see page 351, last paragraph --	1,30,31
A	Plant Physiol., volume 85, 1987, K.A. Barton et al.: "Bacillus thuringiensis $\delta$ -endotoxin expressed in transgenic <i>Nicotiana tabacum</i> provides resistance to lepidopteran insects", pages 1103-1109 see page 1109, left-hand column, last two paragraphs --	3-28
	./.	

\* Special categories of cited documents: <sup>10</sup>

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier document but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

"&" document member of the same patent family

## IV. CERTIFICATION

Date of the Actual Completion of the International Search

5th June 1990

Date of Mailing of this International Search Report

09.07.90

International Searching Authority

EUROPEAN PATENT OFFICE

Signature of Authorized Officer

M. Pez

M. PEIS

III. DOCUMENTS CONSIDERED TO BE RELEVANT (CONTINUED FROM THE SECOND SHEET)		
Category *	Citation of Document, " with indication, where appropriate, of the relevant passages	Relevant to Claim No.
O,A	Biological Abstracts/RRM, BR 35:107674, AN 88:477784, M. Adang et al.: "Engineering crop plants for insect resistance", & 154th National American Association for the Advancement of Science, Annual meeting, Boston, Massachusetts, USA, February 11-15, 1988 see the abstract --	3-28
A	EP, A, 0275957 (HOECHST) 27 July 1988 see the whole document --	4
A	Nucleic Acids Research, volume 17, no. 2, 1989, IRL Press, (Oxford, GB), E.E. Murray et al.: "Codon usage in plant genes", pages 477-498 see the whole document --	4
A	EP, A, 0223452 (MONSANTO) 27 May 1987 see example 11 --	28
E	EP, A, 0359472 (LUBRIZOL GENETICS) 21 March 1990 see page 22, lines 1-14; claim 12 -----	1-14,25, 28,30-38



# ANNEX TO THE INTERNATIONAL SEARCH REPORT ON INTERNATIONAL PATENT APPLICATION NO.

US 9000778

SA 34761

This annex lists the patent family members relating to the patent documents cited in the above-mentioned international search report. The members are as contained in the European Patent Office EDP file on 29/06/90  
The European Patent Office is in no way liable for these particulars which are merely given for the purpose of information.

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
EP-A- 0142924	29-05-85	AU-B- 574101	30-06-88
		AU-A- 3347384	28-03-85
		JP-A- 60094041	27-05-85
EP-A- 0275957	27-07-88	DE-A- 3737918	02-03-89
		AU-A- 1061988	28-07-88
		JP-A- 63273479	10-11-88
		ZA-A- 8800368	05-07-88
EP-A- 0223452	27-05-87	AU-A- 6452886	30-04-87
		JP-A- 62201527	05-09-87
EP-A- 0359472	21-03-90	AU-A- 4118289	15-03-90

**THIS PAGE BLANK (USPTO)**

**This Page is Inserted by IFW Indexing and Scanning  
Operations and is not part of the Official Record**

**BEST AVAILABLE IMAGES**

Defective images within this document are accurate representations of the original documents submitted by the applicant.

Defects in the images include but are not limited to the items checked:

- ☒ **BLACK BORDERS**
- ☒ **IMAGE CUT OFF AT TOP, BOTTOM OR SIDES**
- ☒ **FADED TEXT OR DRAWING**
- ☐ **BLURRED OR ILLEGIBLE TEXT OR DRAWING**
- ☐ **SKEWED/SLANTED IMAGES**
- ☐ **COLOR OR BLACK AND WHITE PHOTOGRAPHS**
- ☐ **GRAY SCALE DOCUMENTS**
- ☒ **LINES OR MARKS ON ORIGINAL DOCUMENT**
- ☐ **REFERENCE(S) OR EXHIBIT(S) SUBMITTED ARE POOR QUALITY**
- ☐ **OTHER:** \_\_\_\_\_

**IMAGES ARE BEST AVAILABLE COPY.**

**As rescanning these documents will not correct the image problems checked, please do not report these problems to the IFW Image Problem Mailbox.**

**THIS PAGE BLANK (USPTO)**